RESEARCH ARTICLE



Limitations of invasive snake control tools in the context of a new invasion on an island with abundant prey

Shane R. Siers¹, Melia G. Nafus², Jeried E. Calaor^{3*}, Rachel M. Volsteadt¹, Matthew S. Grassi^{3*}, Megan Volsteadt⁴, Aaron F. Collins⁵, Patrick D. Barnhart⁵, Logan T. Huse⁶, Amy A. Yackel Adams⁶, Diane L. Vice⁴

1 U.S. Department of Agriculture, Wildlife Services, National Wildlife Research Center, Barrigada, Guam, USA 2 U.S. Geological Survey Pacific Island Ecosystems Research Center, Hawai'i National Park, Hilo, Hawai'i, USA 3 Research Corporation of the University of Guam, Mangilao, Guam, USA 4 Guam Department of Agriculture, Division of Aquatic and Wildlife Resources, Mangilao, Guam, USA 5 U.S. Department of Agriculture, Wildlife Services, Guam State Office, Barrigada, Guam, USA 6 U.S. Geological Survey Fort Collins Science Center, Fort Collins, Colorado, USA

Corresponding author: Shane R. Siers (shane.r.siers@usda.gov)

Academic editor: J. Jeschke | Received 6 March 2023 | Accepted 24 November 2023 | Published 5 January 2024

Citation: Siers SR, Nafus MG, Calaor JE, Volsteadt RM, Grassi MS, Volsteadt M, Collins AF, Barnhart PD, Huse LT, Yackel Adams AA, Vice DL (2024) Limitations of invasive snake control tools in the context of a new invasion on an island with abundant prey. NeoBiota 90: 1–33. https://doi.org/10.3897/neobiota.90.103041

Abstract

In October 2020, a new population of invasive brown treesnakes (*Boiga irregularis*) was discovered on the 33-ha Cocos Island, 2.5 km off the south coast of Guam. Cocos Island is a unique conservation resource, providing refuge for many lizards and birds, including endangered species, which were extirpated from mainland Guam by invasive predators including brown treesnakes. We sought to evaluate the usefulness of toxic baiting with acetaminophen-treated carrion baits and cage trapping, common tools for the control of brown treesnakes on mainland Guam, as potential eradication tools on Cocos Island. We evaluated multiple bait types and bait presentations: on the ground, suspended in the canopy emulating aerial bait applications and in four plastic-tube bait station configurations intended to exclude non-target species. We monitored all baits with time-lapse cameras. Despite improved exclusion of non-targets by bait station design, most baits were quickly removed by non-target species, particularly coconut crabs (*Birgus latro*) and Mariana monitors (*Varanus tsukamotoi*). Monitoring of 1,250 baits available for 2,427 bait nights resulted in no observations of brown treesnakes taking any bait. Subsequently, we tested two trap types commonly used on Guam and compared trapping success with live versus dead mouse lures. In 10,553 trap nights using live and dead mouse lures, we only captured one brown treesnake, in a trap with a live mouse

^{*} Contracted to USDA Wildlife Services, National Wildlife Research Center.

lure. These baiting and trapping rates are so low as to be ineffectual for all practical purposes. Concurrent visual searching and hand capture of brown treesnakes during initial rapid response efforts demonstrates that these low baiting and trapping success rates are not a result of low snake density. We make a case for our assumption that the ineffectiveness of these tools on Cocos Island is due to the context of extremely high abundance of preferred live prey, primarily large geckos and birds. Our results have profound conservation ramifications, because any future island invasions by brown treesnakes are likely to occur within similarly prey-rich environments where these baiting and trapping methods might be similarly ineffective.

Keywords

bait stations, *Boiga irregularis*, camera traps, conservation, eradication feasibility, incipient population, non-target species, trapping

Introduction

The pace and scale of the introduction and spread of non-native reptiles continues to increase, as does recognition of the attendant ecological and economic harms they cause (Kraus 2009; Reed and Kraus 2010; Kraus 2015; Capinha et al. 2017). Snakes comprise a considerable proportion of these reptile invasions, many of which occur on islands where already-imperilled native species are at risk of extirpation or extinction by snake predation; examples include wolf snakes (*Lycodon aulicus*) on Mauritius and Réunion Island (Deso and Probst 2007), corn snakes (*Pantherophis guttatus*) and boa constrictors (*Boa constrictor*) on multiple Caribbean islands, California kingsnakes (*Lampropeltis californiae*) in the Canary Islands and multiple colubrids in the Balearic Islands (Tonge 1990; Perry et al. 2003; Bushar et al. 2015; Monzón-Argüello et al. 2015; Silva-Rocha et al. 2015). Invasive snake problems can be particularly intractable because of snakes' cryptic nature and ability to withstand long periods without feeding (Durso et al. 2011; Siers et al. 2018a; Yackel Adams et al. 2018; Boback et al. 2020; Nafus et al. 2020). To date, there are no known examples of eradication of an invasive snake population at a scale larger than 1 ha (Campbell et al. 2012; DIISE 2023).

The most well-known and well-studied example of an island snake invasion is that of the brown treesnake (*Boiga irregularis*) on the island of Guam in the Western Pacific. Accidentally transported from the Admiralty Islands to Guam in shipments of military equipment following World War II (Rodda and Savidge 2007; Richmond et al. 2015), this slender, nocturnal, arboreal predator spread throughout the entire island by the mid-1980s (Savidge 1987) and achieved densities unprecedented for any natural non-aggregating snake population (Rodda et al. 1999a). The spread of brown treesnakes across Guam was followed by a wave of negative impacts to all native vertebrate taxa (Wiles 1987; Fritts and Rodda 1998) including collapses in nearly all native bird populations, resulting in the extirpation or extinction of 12 of 15 native forest birds and the functional extinction of the island's entire forest avifauna (Savidge 1987; Wiles et al. 2003). This bird loss was followed by cascading effects on plants, invertebrates and ecological processes (Perry and Morton 1999; Rogers et al. 2012; Caves et al. 2013; Fricke et al. 2014; Freedman et al. 2018). Socioeconomic damages caused by the brown treesnake invasion of Guam include 'home invasions' and painful bites to humans including infants (Fritts 1988; Fritts et al. 1990, 1994), predation on domestic animals including the loss of small-scale poultry production (Fritts and McCoid 1991; Rodda and Savidge 2007), declines in tourism (Hall 1996; Shwiff et al. 2010) and costs of power outages caused by snakes short-circuiting transmission lines (Fritts 2002).

Methods and strategies for brown treesnake control are being developed, tested and implemented for the protection and restoration of Guam's native flora and fauna (e.g. Aguon et al. (2002); Siers et al. (2017a, 2020a, b); Clark et al. (2018); Klug et al. (2021a, b); Pollock et al. (2021)). However, the first and highest priority for invasive brown treesnake control has been interdiction-the prevention of further spread of this harmful predator from Guam to other vulnerable locations throughout the Pacific (Hall 1996; Stanford and Rodda 2007; Perry and Vice 2009; Clark et al. 2018; Engeman et al. 2018). The Commonwealth of the Northern Mariana Islands (CNMI, especially Saipan, Rota and Tinian) and the Hawaiian Archipelago are at particularly high risk of invasion and severe ecological and economic consequences (Fritts 1988; Shwiff et al. 2010; BTSTWG 2015; Yackel Adams et al. 2021). Before the implementation of a full interdiction programme on Guam, live brown treesnakes were too-commonly found in cargo from Guam to Saipan, Hawaii and other destinations; since a USDA Wildlife Services operational control programme began in 1993, such encounters have dropped to nearly zero and Saipan continues to be considered snake-free (Hall 1996; Stanford and Rodda 2007; Yackel Adams et al. 2018, 2021).

Cocos Island (CHamoru name: Islan Dåno') is a small atoll island situated approximately 2.5 km off the southern tip of the main island of Guam. Cocos Island was considered to comprise the majority of remaining snake-free habitat in Guam and is home to many vertebrates susceptible to brown treesnake predation, including some species that no longer persist on mainland Guam. Guam rails (*Hypotaenidia owstoni*: ko'ko'), once extinct in the wild due to brown treesnake predation, were introduced to Cocos Island where they have reproduced and thrived (Medina and Aguon 2000). The endangered Mariana skink (*Emoia slevini*) was extirpated from mainland Guam by brown treesnake predation, but a remnant population was recently rediscovered on Cocos Island (USFWS 2019). Other species, including regionally endemic lizards and birds, also persist on Cocos Island (Rodda and Fritts 1992).

While the high volume of commercial and military cargo and vessels originating from central and northern Guam has been scrupulously inspected for stowaway snakes, traffic between southern Guam and Cocos Island has received relatively little attention. A biosecurity plan was developed for the Island (USDA Wildlife Services 2009) to monitor for incursions of cats, rodents and snakes on Cocos Island, as well as control snakes in high traffic areas, i.e. vessels that visited Cocos Island daily for business. An awareness campaign targeted staff and visitors to Cocos Island to report sightings and conduct boat inspections. The implementation of personal craft inspections was voluntary with no regulatory enforcement.

In October 2020, a local fisherperson reported killing snakes on Cocos Island during a night-time visit to the atoll. Subsequent search efforts by the U.S. Geological Survey's (USGS) Brown Treesnake Rapid Response Team (RRT; Stanford and Rodda 2007) confirmed a population of brown treesnakes on Cocos Island (Guam Department of Agriculture 2020; Barnhart et al. 2022). The RRT intermittently continued night-time searches through September 2021, with additional training exercises ongoing through September 2023 and had sighted 64 brown treesnakes (58 of which were captured and euthanised; U.S. Geological Survey 2023). As of September 2023, the volunteer group Friends of Islan Dåno had captured and removed 36 brown treesnakes (Martin Kastner, Friends of Islan Dåno, written communication, 2023) and USDA Wildlife Services had removed an additional 23 (Alyssa Taitano, USDA, written communication, 2023). Currently, these search and removal efforts continue intermittently while agencies plan for a more comprehensive response, potentially including an eradication effort (USDA Wildlife Services 2021a, b). Preliminary USGS data reflect an apparently reproductive population, with representatives of all size classes captured and much larger and heavier snakes than found in similar samples from Guam (Barnhart et al. 2022).

Several tools and techniques have been developed and continue to be improved for management of invasive brown treesnakes on Guam (Clark et al. 2018). The common human pharmaceutical acetaminophen (paracetamol) has been identified as an effective oral toxicant for brown treesnakes (Savarie et al. 2000; Siers et al. 2021) with a relatively low environmental risk profile (Johnston et al. 2002). A tablet containing 80 mg of acetaminophen has been registered with the U.S. Environmental Protection Agency as a vertebrate pesticide for brown treesnake control (Reg. No. 56228-34). Coupled with carrion baits (typically 4–6 g dead neonatal mice), acetaminophen baiting has been demonstrated to reduce brown treesnake abundance on a landscape scale (Savarie et al. 2001; Clark and Savarie 2012; Siers et al. 2020a, b), has become a mainstay of interdiction operations (Clark et al. 2012; Clark et al. 2018; Engeman et al. 2018) and is suggested to be capable of eradicating brown treesnakes on Guam within snake barriers, as part of an integrated pest management strategy (Nafus et al. 2022). Baiting can be more cost-effective than traditional trapping methods (Clark et al. 2012) and, as such, was thought to be a desirable eradication tool for managing the established brown treesnake population on Cocos Island.

Since the early 1990s, live trapping with cage traps has been the primary method of brown treesnake removal and continues to be a foundational tool for research and management programmes (Engeman and Linnell 1998; Tyrrell et al. 2009; Clark et al. 2018; Engeman et al. 2018). Current trap designs are modifications of crayfish or minnow traps, composed of a cylindrical wire mesh trap body with a funnel at each end. Stock funnel openings are widened and covered with a wire mesh one-way flap to allow snakes access to the trap body, but blocking escape. A live mouse in a protective chamber is the lure that entices snakes into them. Although these brown treesnake traps are considered the most efficient snake traps in the world (Rodda et al. 1999b), care and provisioning of live mice is costly and infrared photography has indicated that many snakes that encounter traps fail to enter the trap (Yackel Adams et al. 2019).

Moreover, as food resources become more abundant, live mouse traps may have decreasing efficacy, which is of potential importance in rapid response settings (Gragg et al. 2007; Stanford and Rodda 2007). Traps are also expensive and prone to damage by non-target species, particularly coconut crabs (*Birgus latro*) and Mariana monitors (*Varanus tsukamotoi* [nee *indicus*]). Due to these drawbacks, trapping is seen as an effective brown treesnake removal tool, but probably with greatest value when integrated with acetaminophen baiting (Nafus et al. 2022).

It is important to conduct pilot evaluations of the utility of potential control tools to establish their effectiveness prior to substantial investments in planning eradication projects (Genovesi 2001; Clout and Williams 2009). Identifying the limitations of control tools is critical for preliminary feasibility assessments and managing eradication costs. Prior to planning for an eradication attempt on Cocos Island, we sought to evaluate the practicality of acetaminophen baiting and live trapping in the context of the Island's prey-rich environment which is similar to possible scenarios if brown treesnakes were to successfully arrive and establish a population in other areas vulnerable to invasion. Our general objectives for both methods were to evaluate: 1) brown treesnake removal rates with various tool implementation methods; 2) interference by non-target species with brown treesnake removal methods; and 3) potential harm to native species from brown treesnake removal methods.

Methods

Study location

Cocos Island (33.6 ha; Fig. 1) is centred at approximately 13.238°N, 144.653°E and located 2.5 km southwest from the southern coast of Guam, forming part of the Merizo Barrier Reef surrounding Cocos Lagoon. The vegetation is described in detail in Fosberg (1960). The substrate is deep, well-drained loamy sand and the flora is primarily *Casuarina equisetifolia* forest in the northeast, while the south-western portion of the island is primarily mixed strand forest comprising *Cocos nucifera*, *Hernandia sonora*, *Guettarda speciosa*, *Merrilliodendron megacarpum*, *Morinda citrifolia*, *Intsia bijuga*, *Casuarina equisetifolia*, *Terminalia catappa*, *Tournefortia argentea*, *Carica papaya*, *Barringtonia asiatica*, *Hibiscus tiliaceus*, *Leucaena leucocephala* and *Thespesia populnea*. Vegetation along the south-eastern shore is dominated by *Pemphis acidula* and *Scaevola sericea*, while the north-western coastline is mostly open sand.

The north-eastern 80% of the Island is under private ownership and the southwestern 20% is owned by the Government of Guam and managed by the Guam Department of Parks and Recreation. The Island is uninhabited, but Cocos Island Resort operates as a day resort offering water-sports, trail walks and food and beverages. The resort closed when the Governor of Guam declared an island-wide public emergency shutdown in response to the COVID-19 pandemic on 13 March 2020. The resort has not yet reopened since then.



Figure 1. Map of Cocos Island 2.5 km from the southern tip of Guam, USA. Orange lines indicate locations of trails used for bait applications and trapping. The top of the image is orientated to the north. Image: Maxar Intelligence 2021.

To minimise disturbance of threatened and endangered species present on the Island, our activities were limited to the edges of existing cart paths and footpaths as per the conditions of our U.S. Fish and Wildlife Service Endangered Species Act consultation (Fig. 1). Cart paths and trails on Cocos Island receive very little maintenance or traffic and are primarily under continuous canopy and do not substantially alter the surrounding forest structure. The majority of mainland Guam brown treesnake research is conducted along road edges and maintained transects (e.g. Christy et al. (2010); Siers et al. (2017b)) so we believe it unlikely that limiting activities to these areas would bias results.

Acetaminophen baiting

We sought to evaluate the relative merits of a variety of potential baits and bait presentation methods on Cocos Island. Preliminary evidence from mainland Guam indicates that brown treesnakes with recent experience feeding on birds may be preferentially attracted to dead bird baits over dead rodents (Nafus et al. 2021). To evaluate bait preferences, we offered three sizes of dead mice and two sizes of dead bird chicks as baits, which where suitable to the presentation method: 4–6-gram dead neonatal mice (DNM); 10–17-g small mice (SM); 18–35-g large mice (LM); 10–14-g small bird baits (hatchling quail, SB); and 25–35-g large bird baits (hatchling chickens, LB) (Fig. 2). All baits were monitored with commercial infrared game cameras (H68, Apeman, Shenzhen, China) set to time-lapse with one image recorded every 30 seconds for 24 hours per day.



Figure 2. Dead animal baits used in this study. Left to right: 4–6-g dead neonatal mouse (DNM); 10–17-g small mouse (SM); 18–35-g large mouse (LM); 10–14-g small bird (quail chick, SB); and 25–35-g large bird (chicken chick, LB).

Canopy presentation: USDA Wildlife Services has engineered an Aerial Delivery System (ADS) for the automated assembly and aerial distribution of bait cartridges containing a DNM treated with a tablet containing 80 mg of acetaminophen (Siers et al. 2019a, 2020b, 2021; Goetz et al. 2020, 2021). These cartridges open upon ejection from the aircraft, exposing a ribbon to cause entanglement in the forest canopy where arboreal treesnakes forage, preventing baits from falling to the forest floor where they can be taken by terrestrial non-target species, such as crabs. We emulated aerial bait applications by positioning opened ADS cartridges on a simulated branch (45-cm wooden dowel) with the DNM hanging in the field of view (FOV) of a camera mounted atop a painter's pole, extended into the forest canopy and temporarily lashed to natural vegetation with bungee cords (Fig. 3A).

Ground presentation: As some ADS baits fail to tangle in the canopy or DNM may become unstuck from the cartridge and fall through to the forest floor, we sought to evaluate the fate of DNM on the ground. Additionally, recent evidence



Figure 3. Camera orientations for canopy and ground bait monitoring **A** infrared camera set-up mimicking aerial application of dead neonatal mouse (DNM) baits via the USDA Wildlife Services Aerial Delivery System (ADS) for landscape-scale brown treesnake control; the camera and bait are elevated into the forest canopy atop a telescoping painter's pole **B** ground bait monitoring set-up, with an infrared camera mounted directly over a large mouse (LM) bait on a tripod constructed from extruded metal tubing (conduit); the bait and a lightweight PVC background with circular size standards were lashed to a 0.9-kg lead diving weight to prevent small crabs from dragging the bait out of the camera's field of view.

indicates that ADS treatments might not adequately expose large brown treesnakes to baits; larger brown treesnakes on prev-depleted Guam are more prone to foraging on the ground, might be preferentially attracted to larger baits and might require greater doses of acetaminophen for effective removal (Rodda and Reed 2007; Nafus et al. 2020; Goetz et al. 2021; Siers et al. 2021). Moreover, the estimated take rates for large ground baits in a treated population on Guam is greater than for standard ADS baits (Nafus et al. 2022). For these reasons, USDA Wildlife Services has considered an alternative aerially delivered bait system comprising a slightly larger mouse (SM) placed in the same bait cartridge tube, but without the ribbon assembly, which would let the bait fall to the forest floor where larger snakes may be more effectively targeted (Siers et al. 2021). We also considered that even larger baits placed on the forest floor, potentially containing larger doses of acetaminophen, could more effectively target larger ground-foraging snakes, so we incorporated LM and LB into ground presentations. Ground baits were placed on small, thin PVC plastic platforms printed with 20-cm size standards and lashed to a 0.9-kg. lead diving weight to prevent small crabs from dragging the bait out of the field of view of the cameras. Cameras were mounted directly overhead on tripods fashioned from inexpensive extruded metal tubing (conduit; Fig. 3B). Brown treesnake head measurements taken from overhead images containing a size standard can be used to estimate snake size (Siers 2021). As both canopy and ground baits are not protected from being taken by non-target species, we did not treat these baits with acetaminophen tablets for this pilot evaluation.

Bait station presentations: Polyvinyl chloride (PVC) tube bait stations ('bait tubes') are intended to exclude non-target species that might interfere with baits, making them unavailable to brown treesnakes and to protect native species from unintentional exposure to acetaminophen intoxication. Standard operational baiting methods include placing a DNM treated with a tablet containing 80 mg of acetaminophen into a 5-cm diameter, 30-cm long PVC bait tube, with 6.35-mm bolts crossing the openings at the ends to further prevent ingress by non-targets. Bait tubes are usually suspended horizontally by two lengths of paracord from existing vegetation or structures, such as fence lines (Savarie et al. 2001; Clark et al. 2012; Lardner et al. 2013; Clark et al. 2018). Based on the average size of brown treesnakes recovered from Cocos Island prior to the testing of these tools (Barnhart et al. 2022), we elected to deploy larger baits (SM, LM, SB and LB) within our bait stations on Cocos Island. We evaluated standard 5 × 30-cm horizontal bait tubes (Fig. 4A), as well as alternative designs intended to more reliably exclude non-target species, such as crabs and Mariana monitors (e.g. Mathies et al. (2011)): longer 5 × 45-cm horizontal bait tubes (Fig. 4B); vertical 5-cm diameter × 30-cm long bait tubes capped at the top end (Fig. 4C); and capped vertical 10-cm diameter × 30-cm long bait tubes (Fig. 4D). Both horizontal bait tube designs included bolts across the openings, whereas vertical tubes did not to prevent use of the bolts by non-targets to assist climbing into the tubes.



Figure 4. Camera orientations and bait station configurations for bait monitoring **A** standard 5-cm diameter \times 30-cm long polyvinyl chloride (PVC) horizontal bait station suspended by nylon paracord from a wooden dowel armature with infrared camera positioned with the bait in the field of view; the two ends of the armature are temporarily lashed to natural vegetation with elastic cords **B** extended 5 \times 45-cm horizontal bait tube **C** capped 5 \times 30-cm vertical bait tube **D** capped 10 \times 30-cm vertical bait tubes. Baits in horizontal bait tubes were held in place by gravity, while baits were held in the caps of vertical bait tubes by spring clamps on one foot of the bait. As baits in vertical bait tubes were not visible to the camera, a length of biodegradable flagging was tied to one leg of the bait and pulled through a small hole in the side of the tube for ease of identifying when the bait was taken during camera image review.

We suspended all bait station types by paracord from a wooden dowel armature to which the trail camera was affixed, locking the bait in the FOV of the camera. We temporarily attached these armatures to existing vegetation by elastic cords, with minor vegetation pruning to ensure that leaves or branches did not obstruct the view of the bait. We placed baits in the centre of horizontal tubes where they were held in place by gravity and friction alone. Within vertical bait tubes, we attached baits within the caps clipping one foot with a small metal spring clamp, holding baits in place, but making them easily removed with a slight tug from snakes or non-targets. As cameras could not directly view baits in the vertical tubes, we tied a length of white biodegradable flagging tape to one leg of each bait and pulled the tape through a small hole in the side of the tube; when the bait was removed, the flag disappeared and the animal within the FOV of the camera at that time was attributed with the bait removal.

As bait stations offered some degree of protection from non-target interference, we treated baits in bait stations with tablets containing 80 mg of acetaminophen inserted into the body of the bait via the oral cavity, to remove any brown treesnakes that took baits.

We spaced bait placements at approximately 20-m intervals along existing paths and trails on Cocos Island (Fig. 1); entry into the forest was not authorised prior to a formal endangered species consultation process with the U.S. Fish and Wildlife Service. Paths were subdivided into nine, segments (transects). Each week we placed 36 bait monitoring stations along one transect, alternating each of three presentation types (12 stations each). We monitored each transect of 36 stations for one week, with the bait checked and replaced once mid-week with a fresh bait, for baiting intervals of 3 to 4 days. Beyond 3 to 4 days, baits are degraded through putrefaction and consumption by ants and fly larvae and are no longer considered viable for take by brown treesnakes. During Weeks 1-9 (12 December 2020-16 February 2021), we placed canopy, ground and standard 5 × 30-cm horizontal tubes (Figs 3A, B, 4A) in alternating positions along each transect, with bait types alternated as appropriate to the objectives of the presentation type. During Weeks 10-18 (18 February 2021-22 April 2021), we alternated 5×45 -cm horizontal tubes and the two vertically orientated tube types $(5 \times 30 \text{ and } 10 \times 30 \text{ cm}; \text{ Fig. 4B-D})$, along with alternating bait types. At bait checks, if a bait appeared to have been taken, we reviewed camera images to identify the time of bait removal and the species taking the bait.

This portion of the study was performed during Guam's cooler, drier months. Average daily temperatures for Guam ranged from highs of 30.4 °C (standard deviation = 0.903 °C) to lows of 24.7 °C (SD = 1.03 °C) and rainfall averaged 2.16 mm/day (SD = 5.3 mm/day, max = 48 mm/day), based on National Oceanic and Atmospheric Administration data (www.weather.gov).

Live trapping

After years of experimentation with multiple live trap designs, a modified crayfish or minnow trap was adopted as the standard brown treesnake live trap used on Guam. The original trap is a two-piece dual-funnel design of galvanised wire mesh with the entrances modified with a PVC ring holding a one-way wire mesh flap that allows access to the trap body, but blocks escape by snakes (Fig. 5, left). Rodda et al. (1999b) showed this trap to be the most effective trap known for any snake at the time. Snakes are lured into the traps by a live mouse in a protected wire mesh chamber held within the trap body. This two-piece design was later adapted for operational purposes by creating a single-piece body of more durable stainless steel, accessed by removing one of the funnel ends and incorporating the mouse chamber into the trap body so that the mouse can be serviced without opening the trap; this version of the trap is referred to as the USDA Wildlife Services "WS Standard" (Fig. 5, centre; Vice et al. (2005)). In both trap types, mice are provisioned with a custom-made block of commercial seed and pellet mix embedded in a paraffin wax matrix which prevents exposure to the elements until the mouse chews through the wax (Fig. 5, right). A piece of fresh potato provides the necessary water.

The use of live mouse lures is less than desirable due to maintenance expense and perceptions regarding animal welfare; however, despite extensive efforts, no trap lure has been found to be nearly as effective and practical as a live mouse (Chiszar 1990; Shivik and Clark 1997; Shivik 1998, 1999; Lindberg et al. 2000). Prior to verification of brown treesnake presence on the Island, Guam's Division of Aquatic and Wildlife Resources (GDAWR) performed surveillance trapping using dead mouse and rat lures due to concerns about escaped live mice establishing a population on the Island (D. Vice, GDAWR, written communication, 2023) but no snakes were ever captured in these traps. Prior to this study, there were no reported head-to-head tests of live versus dead mouse lures in brown treesnake traps.

To evaluate differences in efficacy and durability between trap types and capture success between live and dead mouse lures, we alternated 99 one-piece WS Standard and 99 original two-piece traps approximately every 20 m along the same existing trails as the previous baiting trials (Fig. 1). We alternated live and dead mouse lures between



Figure 5. Two types of traps used. Left: Galvanised wire mesh two-piece trap with separate live mouse lure chamber within the trap body. Centre: Stainless steel one-piece Wildlife Services Standard trap with integrated live mouse lure chamber. Right: Integrated lure chamber in one-piece trap showing live mouse, feed block of pellets and seeds immersed in paraffin wax and piece of fresh potato to provide moisture; mice in both trap types are provisioned in this manner.

every pair of two trap types (repeating the order of one-piece/live, two-piece/live, onepiece/dead, two-piece/dead etc.) for a total of 100 traps with live mouse lures and 98 with dead mice. Following the typical use patterns of these two trap types, we hung one-piece traps on nylon paracord and two-piece traps on metal tie wire, at about waist to chest height on existing vegetation. We checked traps twice weekly (every 3 or 4 days) for 55 nights, provisioning live mouse lures and replacing dead mouse lures with fresh dead mice. We recorded brown treesnake captures, non-target captures and trap damage caused by non-target species. We also recorded traps as non-functional when missing lures, with funnel flaps stuck open or closed, with large holes due to crab damage or with other defects making them unlikely to capture or prevent escape by snakes. As traps were confirmed to be functional at the beginning of each trap-checking interval, we assumed traps became non-functional approximately mid-interval, on average, so reduced our tally of effective trap nights by one-half of the checking interval per non-functioning trap, similar to methods of Nelson and Clark (1973). Trapping results are reported as captures per unit effort (CPUE) or snake captures per night per trap.

We monitored a subset of 20 traps via infrared game cameras (Hyperfire 2, Reconyx, Holmen, Wisconsin). We distributed 10 cameras evenly along a rock retaining wall and another 10 along a transect through a bird roosting area. We positioned half of these cameras on traps with live mouse lures and the other half on traps with dead mice. We recorded time-lapse images (one photo every 60 seconds) between 1800 and 0600 h to observe for brown treesnakes investigating traps, but failing to enter, as has been documented on mainland Guam (Yackel Adams et al. 2019; Amburgey et al. 2021).

We performed trapping from 17 June to 12 August, 2021, earlier months of Guam's warmer, rainier season. Guam daytime highs averaged 31.4 °C (SD = 1.04) with night-time lows of 25.6 °C (SD = 0.969) and rainfall of 9.22 mm/day (SD = 13.5mm, max = 71.9mm) (www.weather.gov). We measured snout-vent length (SVL, mm) of trapped snakes by gently stretching them along a flexible tape and measured weight using handheld spring scales with maximum ranges from 10 g (0.1 g precision) to 1000 g (10 g precision) (Pesola, Schindellegi, Switzerland). We determined sex by probing for inverted hemipenes with steel sexing probes (Reed and Tucker 2012).

Visual detection during rapid response

Throughout the evaluation of control tools, USGS conducted nocturnal visual searches and hand-removal of brown treesnakes (December 2020 through July 2021). Methods followed those applied on Guam in which individuals surveyed transects after dusk using powerful headlamps (Wilma, Lupine Lighting System, Lebanon, PA, USA) walking a slow searcher pace, such that each transect (~ 400 m) lasted approximately 1 hour. During snake searches, observations of potential prey items (lizards, birds and bats) were recorded. Visual survey data are available for download (U.S. Geological Survey 2023). These searches were also limited to the cart paths and trails depicted in Fig. 1. We recorded SVL, weight and sex for captured snakes as above.

Statistical methods

All summary statistics, statistical tests and graphing were performed in the R environment for statistical computing, Version 4.2.2 (R Core Team 2021). We evaluated the likelihood that an unobserved bait take due to camera malfunction could have been taken by a snake by describing the 95% confidence interval of brown treesnake bait takes given the successfully observed baits (binom.confint, method = "exact"). We evaluated differences in duration of bait availability amongst presentation types with Cox proportional hazard survival models function coxph), with trials ending when the bait was taken by a non-target or when the three to four days monitoring period was over. Differences in trap capture rates amongst trap types and trap lure types were calculated using Fisher's exact tests with 95% confidence intervals (fisher.test).

Ethics statement

This study was carried out in compliance with relevant laws and guidelines. All animal use was approved by the USDA National Wildlife Research Center Institutional Animal Care and Use Committee under protocols QA-3106 and QA-3340 and USGS Institutional Animal Care and Use Committee protocol 2021-02. Compliance with the Endangered Species Act was ensured through informal consultation with the U. S. Fish and Wildlife Service (01EPIF00-2021-I-0087 and 01EPIF00-2021-I-0087-R001).

Results

Acetaminophen baiting

After eliminating incomplete trials (camera failure etc.), we successfully monitored a total of 1,250 baits between December 2020 and April 2021. During these trials, we observed no baits being investigated or taken by brown treesnakes. As there were no bait takes by brown treesnakes, we could not make comparisons of bait take rates amongst bait types or presentation types. Of the 701 baits that were taken, we could not identify the species in 30 (4.3%) of the cases. The 95% binomial confidence interval for brown treesnake takes for the 671 baits for which a species ID was confirmed is 0–0.548%; if this rate were applied to the 30 unknown takes, the upper confidence limit for brown treesnake bait takes would be 0.164 baits; thus, we consider it highly unlikely that any of the unknown animals taking these 30 baits was a brown treesnake.

A high overall proportion of the baits (56.1%) were taken by non-target species (Fig. 6), primarily by coconut crabs and Mariana monitors. Canopy baits were removed by coconut crabs (11.5%), Mariana monitors (12%) and insects (11.5%, mostly ants and fly larvae), while 20.7% fell from the simulated bait cartridge due to putrefaction or consumption by insects. One bait was removed from the bait cartridge by a large gecko (*Gehyra oceanica*). Of the DNM baits in the canopy, 43.8% remained available (i.e. were not taken by non-targets), but were not observed being investigated or taken by brown

treesnakes. All but three baits placed directly on the ground were taken by non-targets, mostly coconut crabs and Mariana monitors, with some take by hermit crabs (*Coenobita* spp.) and land crabs (likely *Cardisoma carnifex* or *Discoplax* spp.). One DNM ground bait was taken by a Pacific reef heron (*Egretta sacra*) and another by a Guam rail. Of baits offered in the standard 5×30 -cm horizontal tubes, 66.3% were taken by Mariana monitors and coconut crabs. The other bait station designs successfully repelled almost all coconut crabs; Mariana monitors continued to be the primary challenge for all bait station types, but the 5×45 -cm horizontal tube reduced monitor takes to only 15.8% (Fig. 6).

Coconut crabs and Mariana monitors were by far the most common consumers of baits. Plotting the recorded time of bait takes by coconut crabs and Mariana monitors (Fig. 7) reflected clear patterns of nocturnal activity for coconut crabs and diurnal activity for Mariana monitors, with bimodal peaks of monitor activity in early morning and late afternoon.



Figure 6. Fates of carrion baits applied on Cocos Island. No brown treesnakes were observed taking any baits. DNM = dead neonatal mouse (4–6 g); SM = small mouse (10–14 g); LM = large mouse (25–35 g); SB = small birds (10–14 g quail chick); LB = large bird (25–35 g chicken chick). Sample size is the number of baits successfully monitored after eliminating incomplete trials.



Figure 7. Times of bait removal by the primary non-target consumers on Cocos Island. Most baits were removed by coconut crabs (*Birgus latro*) and Mariana monitors (*Varanus tsukamotoi*). Light grey strips indicate changes in sunrise and sunset over the study period.

We recorded the duration of bait availability before being taken by non-targets or removed at the end of the trial and subjected these data to survival analysis (Fig. 8). Non-targets removed almost all ground baits within 24 hours. The horizontal 5×45 - cm bait tube clearly outperformed all other presentation types, with 80% of the baits remaining at the end of the observation period.

Summing all the time that baits were available to brown treesnakes before being taken by non-targets, we recorded a total of 2,427 "bait days" with no takes by brown treesnakes. This amounts to an overall daily estimated brown treesnake bait take rate of 0.000 per 100 bait days with an upper 95% binomial confidence interval of 0.151 baits per 100 bait days.

Trapping

We operated 198 traps (99 one-piece, 99 two-piece) with mouse lures (100 live, 98 dead) for 55 nights, for a total of 10,890 trap nights (Table 1) between and June and



Figure 8. Persistence of all baits offered in various presentation types. Curves depict reduction in proportion of baits available over time as non-targets remove them. Baits typically decline in viability through putrefaction and consumption by insects after 48 to 96 hours. Shaded areas represent 95% confidence intervals.

Table 1. Trapping effort (trap nights) by trap type and lure type. Values reflect overall effort and effort adjusted for non-functioning traps (e.g. lure missing, entrance flaps stuck open or closed, holes due to crab damage). Traps were functional at the previous check, so non-functional traps were presumed to be functional for one-half of the check interval, on average.

Trap type	Overall effort (trap nights)			Adjusted for non-functioning traps			
	Live mouse	Dead mouse	Total	Live mouse	Dead mouse	Total	
One-piece	2,750	2,695	5,445	2,682.5	2,620.0	5,302.5	
Two-piece	2,750	2,695	5,445	2,663.0	2,588.0	5,251.0	
TOTAL	5,500	5,390	10,890	5,345.5	5,208.0	10,553.5	

August of 2021. After adjusting for non-functioning traps, we achieved an effort of 10,553.5 effective trap nights.

During this effort, we captured only one brown treesnake: a 1,249-mm SVL male weighing 360 g (Fig. 9). The snake was captured in a two-piece galvanised trap with a live mouse lure. One capture in 10,553 trap nights yields a combined CPUE of 9.47^{e-5} or 0.00947 captures per 100 trap nights. Considering CPUE only for traps with live mouse lures increased CPUE to 0.0187 captures per 100 trap nights. The 95% binomial confidence interval for trap success (probability of each trap capturing at least one snake on a given night) with a live mouse lure is 0.000473 to 0.0528 trap successes per 100 trap nights.

Non-target lizards and crabs were commonly found in brown treesnake traps (Table 2). As with acetaminophen baiting, coconut crabs and Mariana monitors were the most significant non-targets, being large and powerful and capable of damaging traps. Smaller lizards were more common in traps with dead mouse lures, likely drawn by flies and other insects feeding on the carrion lure. Crabs, including coconut crabs, were more often found in the one-piece WS standard traps, possibly because one-piece traps were hung with nylon paracord, per WS standard operations, while two-piece traps were hung by metal tie-wire; it is conceivable that metal wire provides less purchase for crabs attempting to access traps. Crabs were more prevalent in traps with live mouse lures, probably attracted by feed waste and faeces generated by the mouse.



Figure 9. The only brown treesnake captured in over 10,500 trap nights on Cocos Island. This was a 1,249 mm snout-vent length, 360 g, male. This trap contained a live mouse in a protected chamber as the lure.

Common name	Latin name	One-piece	Two-piece	One-piece	Two-piece	Total
		Live	Live	Dead	Dead	
Lizards						
Green anole	Anolis carolinensis	1	0	2	3	6
Blue-tailed skink	Emoia caeruleocauda	0	0	1	0	1
Oceanic gecko	Gehyra oceanica	0	0	2	0	2
Other geckos	Gekkonidae	1	1	1	1	4
Mariana monitor	Varanus tsukamotoi	8	11	14	5	38
Crabs						
Hermit crabs	Coenobita spp.	183	24	14	1	222
Coconut crab	Birgus latro	57	9	22	17	105
Totals		250	45	56	27	378

Table 2. Non-target captures in brown treesnake traps. Results are tabulated by trap type (one-pieces stainless steel versus two-piece galvanised) and lure type (live mouse or dead mouse). Counts are per trap observation; multiple individuals in the trap at the same time are counted as only one observation.

Stainless steel one-piece traps were non-functional for 5.2% of trap nights, while galvanised two-piece traps were non-functional for 7.1% of nights (P < 0.001). Live mouse lure traps were non-functional 5.6% of trap nights and dead mouse lure traps were non-functional 6.8% of nights (P = 0.015). Stainless steel one-piece traps required 21 field repairs for a total of 2.83 hours of labour, while galvanised two-piece traps required 82 repairs totalling 12.4 hours of field labour. Only one trap, a twopiece galvanised trap, was removed for workshop repairs. At the end of the trapping effort, repair and cleaning requirements were recorded by trap type (Table 3). The one-piece stainless steel traps were more durable to crab damage and required less labour to fix, primarily straightening deformations of unbroken wire mesh, while all damaged two-piece galvanised wire traps required more serious repairs, such as hole patches. No traps of either type were damaged beyond repair. Although galvanised wire traps are likely more prone to corrosion than stainless steel over prolonged use in the near-marine environment, there was little noticeable corrosion over the course of this study. The one-piece traps required more time to clean due to the lure chambers being integrated into the trap body.

Table 3. Repair and cleaning requirements by trap type. Minor repairs included straightening wire deformations from coconut crab damage, while major repairs required patching of holes in the mesh from crab damage.

Trap type	No repair (n)	Minor repair	Major repair	Repair labour	Cleaning
		(n)	(n)	(hr)	labour (hr)
One-piece stainless steel	28	47	24	13	20
Two-piece galvanised wire	31.5*	0	67.5	27	7
Totals	59.5*	47	31.5	40	27

* The two-piece traps were recorded by interchangeable trap halves (two halves = one trap), hence the 0.5-trap increments.

During 1,100 trap nights monitored by infrared cameras (20 cameras, 647,733 total photos), we recorded only two trap encounters by brown treesnakes, of 2 and 8 minutes in duration (Fig. 10). In neither of these cases did the snake successfully enter the trap. Both recorded encounters were of a similarly-sized snake on the same trap with a live mouse lure 10 nights apart; we believe it reasonable to consider these to be two observations of the same snake.

Visual detection during rapid response

Throughout the time period of our baiting and trapping efforts (December 2020 to July 2021), USGS personnel performed 163.3 km of linear search effort over 376 hours and sighted 31 snakes, three of which escaped capture, for a visual detection rate of 0.188 snakes per km and 0.083 per hour of search. Prey sighting rates were high, with an average lizard sighting rate of 37.8/km (including 10 species) and 32.8 birds/ km (8 species). Specifically for notable species, sighting rates were 15.5 green anoles, 11.3 oceanic geckos, 22.3 black noddies and 8.1 white terns per km of searching (U.S. Geological Survey 2023). The mean SVL of snakes captured and removed was 1073 \pm 295 (range: 650, 1622) and mean weight was 256 \pm 195 g. The demography of the population encountered was biased towards snakes typically attracted to endothermic prey (60% of 28 captures > 900 mm SVL).



HYPERFIRE 2 COVERT

Figure 10. A large brown treesnake at a two-piece trap with a live mouse lure recorded via time-lapse infrared photograph. The snake probed the body of the trap for 8 minutes, then left without returning that night. A snake of a similar size (likely the same snake) attempted to enter the same trap for 2 minutes 10 days later.

Discussion

The results of this study are clear: two of the primary tools for brown treesnake removal on Guam will not be effective for brown treesnake eradication on Cocos Island. Brown treesnake detection rates, based on visual CPUE, are apparently lower on Cocos Island (0.188 snakes/km) than most other detection efforts on Guam. Within a long-term 5-ha geographically enclosed population representative of disturbed limestone forest and secondary forest on Guam, Nafus et al. (2023) recorded 0.758 brown treesnakes per km of transect searched. In a 55-ha snake enclosure surrounding degraded limestone forest in northern Guam, Boback et al. (2020) documented a CPUE of 0.906 brown treesnakes per searcher hour, compared to 0.083 on Cocos Island. Following aerial baiting at this same site (Dorr et al. 2016; Siers et al. 2018), the CPUE dropped to 0.049 per hour, lower than the Cocos Island CPUE. With all of the caveats that come with using CPUE as an index of relative abundance (e.g. detectability differences amongst different habitat types), these data demonstrate that visual detection rates on Cocos Island are lower than unmanipulated habitat on Guam (24% compared to Nafus et al. (2023) per km and 9.2% compared to Boback et al. (2020) per hour before aerial suppression), but nearly twice as high as within an aerially-suppressed study plot (Boback et al. 2020). Nonetheless, despite 40% lower visual contact rates than on Cocos Island, the suppressed population on Guam continued to take non-toxic dead mouse baits at rates averaging approximately 20% (Siers et al. 2018b), while the Cocos Island bait take rate was 0%.

Live trapping with mouse lures prior to snake suppression on Guam yielded a capture rate of 0.3 snakes per 100 trap days (Nafus et al. 2018). These differences in snake detection rates by location indicate that greater bait or trap captures on Cocos Island would be expected given the level of effort we applied to each tool. Carrion bait take rates on Guam in areas without active brown treesnake control tend to range from approximately 30% to nearly 100% (Savarie et al. 2001; Clark and Savarie 2012; Siers et al. 2018b, 2019b, 2020a), while we failed to observe a single bait take within 2,427 bait monitoring days (upper 95% confidence interval of 0.151%). Trapping captures per 100 trap nights on Guam are commonly higher in areas where they are not being operationally suppressed, (4-9; Nafus et al. (2018)) and ranged as high as 25 to 60 in the 1990s (Rodda et al. 1999b). In areas on Guam in which brown treesnakes have been suppressed to 0.16 snake/ha (as estimated from forest interior visual survey CPUE), trapping CPUE was maintained at the rate of 0.3 captures per 100 trap nights (Nafus et al. 2018; Boback et al. 2020). When using live mouse lures on Cocos Island, we achieved only 0.0187 captures per 100 trap nights which is substantially lower than the anticipated levels based on Guam efforts (Guam CPUE 213 to 481 times higher than Cocos Island when compared to Nafus et al. (2018) results).

Our failure to attract brown treesnakes to baits and traps is most likely due to the extraordinary abundance of preferred live prey on Cocos Island, particularly large geckos, birds and their eggs, compared to the relatively prey-depauperate nature of Guam's forests resulting from prolonged brown treesnake predation (Fritts and Rodda

1998; Wiles et al. 2003; Siers 2015). Cocos Island is populated by abundant lizards, many of which have been extirpated from parts or all of Guam by brown treesnake predation, such as federally and locally endangered Mariana skinks, locally endangered littoral skinks (Emoia atrocostata), fragile Micronesian geckos (Perochirus ateles) and Pacific snake-eyed skinks (Cryptoblepharus poecilopleurus), as well as mutilating geckos (Gehyra mutilata), oceanic geckos (G. oceanica) and green anoles (Anolis carolinensis). Small brown treesnakes, in particular, appear to be specialists on small lizards, which are an almost exclusive prey item in stomach contents (Savidge 1988; Siers 2015). They also exhibit strong preference during laboratory feeding trials (Lardner et al. 2009) and present a venom composition that is more effective for ectotherms as juveniles (Mackessy et al. 2006). Our green anole sighting rates were relatively high (15.5/km), but close to estimates from the snake-free island of Saipan (14.9/km; Lardner et al. (2019a)), while observations of this species in brown treesnake stomach contents from mainland Guam are extremely low in forest habitats, from where they have been essentially extirpated by brown treesnake predation (Rodda and Fritts 1992; Siers 2015). Relatively large oceanic geckos, which are roughly equivalent in mass to a small mouse, may additionally offer a rewarding prey item that reduces the efficacy of baitand lure-based control tools on Cocos Island. We commonly observed oceanic geckos on Cocos Island (11.3/km), while they are no longer documented in brown treesnake stomach contents on mainland Guam (Siers 2015), having been essentially extirpated by brown treesnake predation (Rodda and Fritts 1992). Nafus et al. (2023) recorded no observations of green anoles or oceanic geckos and their total lizard sighting rates were 6.0/km (4 species) compared to 37.8/km (10 species) on Cocos Island, indicating

substantially greater lizard prey availability. Unchecked brown treesnake predation and population growth could put all small lizard species at risk of almost certain local extinction on Cocos Island (Rodda and Fritts 1992; Rodda et al. 1997; Fritts and Rodda 1998; USFWS 2019).

Of all the ecological damage that have occurred since their introduction to Guam, the pervasive impacts of brown treesnake predation on birds have been the most profound (Savidge 1987; Wiles et al. 2003; Pollock et al. 2019; Klug et al. 2021b). Our Cocos Island bird sighting rate of 32.8/km and 11.3/hour is high, given the well-documented collapse and functional extinction of the forest bird avifauna on mainland Guam (Savidge 1987; Wiles et al. 2003). Any experienced brown treesnake searcher can attest that bird sightings in forest habitats on mainland Guam are quite rare, earning Guam its reputation for "silent forests" and the cascading ecological effects of bird loss (Savidge 1987; Rodda et al. 1997; Rogers 2011, 2020). Nafus et al. (2023) documented only four bird sightings of indeterminate species in 816.2 km of transect searching in the northern Guam 5-ha enclosure (0.005 birds/km). It is unclear how severely brown treesnake predation on Cocos Island has affected the abundance of bird and lizard prey species, but prey resources clearly remain much more abundant on Cocos Island.

Brown treesnakes on Guam that forage in areas of increased prey availability, including birds, such as urban areas and swiftlet caves, tend to be in better body condition (Siers 2015; Siers et al. 2017b; Yackel Adams et al. 2019; Klug et al. 2021b), a characteristic that is evident in the extremely heavy snakes that have been found during visual surveys on Cocos Island. The mean weight of snakes reported in other contemporary studies on Guam (Siers et al. 2017b) suggests that the average weight of snakes removed from Cocos Island during the period of this study is much greater. There is also emerging evidence that brown treesnakes conditioned to feeding on live birds exhibit less attraction to rodent-based baits and lures (Nafus et al. 2021). During brown treesnake removal from Cocos Island, encounters with seabirds, Guam rails and Micronesian starlings (*Aplonis opaca*) were not uncommon (U.S. Geological Survey 2023), supporting the impression that avian prey currently remain abundant on Cocos Island relative to Guam. For these reasons, it is apparent that, although brown treesnakes on Guam will readily consume carrion, live lures are far more effective than any dead animal baits or other inanimate lures (Shivik et al. 2000; Savarie and Clark 2006; Kimball et al. 2016) and that the availability of abundant preferred prey (birds) on Cocos Island diminishes the attraction to carrion baits.

These findings demonstrate that higher prey availability negatively affects brown treesnake detection and capture rates. On Guam, temporary experimental suppression of rodent prey abundance was demonstrated to increase trap capture rates (Gragg et al. 2007) and increasing movement distances of brown treesnakes (Christy et al. 2017). Free-ranging brown treesnakes that have recently taken large meals have been experimentally demonstrated to significantly reduce movement for 5 to 7 days, with an associated reduction in the ability to visually detect or trap snakes during this time (Siers et al. 2018a). Foraging for carrion may also be a futile strategy for brown treesnakes on Cocos Island where coconut crabs and Mariana monitors rapidly remove all carrion from the ground (Figs 7–9). Moreover, prior studies have indicated that brown treesnake attraction to carrion and especially mouse carrion, may decrease as snakes increase in size (Shivik and Clark 1999; Nafus et al. 2021), while large snakes may be the most important demographic to remove from the perspective of both avian conservation and eradication potential (Savidge 1988; Yackel Adams et al. 2019; Nafus et al. 2022).

Both baiting and trapping appear to be relatively safe for Cocos Island wildlife. In only two instances did a native bird (a Pacific reef heron and a Guam rail) take a bait (both DNM) from the ground. Although acetaminophen may also be toxic to birds, they are not known to have the same genetic basis for sensitivity to acetaminophen toxicosis as snakes (van den Hurk and Kerkkamp 2019) and crows in cage trials picked around acetaminophen tablets rather than ingesting them, with no signs of toxicosis (Avery et al. 2004). Clearly, crabs and Mariana monitors quickly cleanse the forest floor of any carrion baits containing acetaminophen. Crabs tend to eat around acetaminophen tablets when consuming carrion baits and show no signs of toxicosis (Johnston et al. 2002). On the other hand, Mariana monitors were the most problematic scavengers of baits on the ground and in bait stations and other monitor species are susceptible to acetaminophen toxicosis (Mauldin and Savarie 2010). Although recent evidence indicates that Mariana monitors are native to the Mariana Islands (Weijola et al. 2020), Mariana monitors on Cocos Island are being actively depredated

by GDAWR for the protection of endangered Guam rails and their nests. It is likely that several small Mariana monitors succumbed to acetaminophen toxicosis and would continue to do so if acetaminophen were used as part of a brown treesnake eradication programme on Cocos Island. Small Mariana monitors were also frequently caught live in traps; these could be released unharmed, but GDAWR requested that trapped Mariana monitors be removed and euthanised rather than released, in furtherance of their Guam rail protection efforts. Some small coconut crabs and hermit crabs expired in brown treesnake traps, likely due to dehydration, although the numbers would not be expected to have population-level impacts. Factors such as these need to be considered when assessing the potential environmental impacts of baiting or trapping for invasive snakes, although ineffectiveness in our case could make these issues moot.

The practical information on baiting and trapping is of little avail when brown treesnake removal by these techniques is almost completely ineffectual in the context of abundant alternative prey availability. Initial eradication discussions for Cocos Island included a notional plan of a 20×20 -m grid of bait stations and a 40×40 -m grid of traps. At this density, we might have arrayed as many as 825 bait stations and 206 traps with live mouse lures across the entire 33 ha of the Island for the duration of an eradication attempt that is expected to last at least 5 to 10 years (based on ad hoc population estimates and demographic projections; USDA Wildlife Services (2021a)). The pilot studies we report here have forestalled what might have been very costly investments in baiting and trapping with little or no payoff. Financial estimates for the costs of an island-wide baiting and/or trapping effort are beyond the scope of this article, complicated and inflated by considerations, such as boat travel, transect establishment etc., although these issues also affect visual search and removal efforts.

Instead, available funding is being programmed for visual searching and manual removal of snakes, the only tactic that has thus far been effective on Cocos Island. Although night-time visual searching can be logistically challenging, disruptive to work schedules and tedious, it is also the tool that is considered to be the least biased with respect to snake sizes, putting all brown treesnake size classes at risk of detection (Rodda et al. 2007; Christy et al. 2010; Yackel Adams et al. 2018; Lardner et al. 2019b). To date, there have been no documented successful invasive snake eradications beyond a temporary 1-ha experimental exclusion plot (Campbell et al. 2012); however, with judicious and sustained application of the right techniques, the small, isolated island of Cocos Island could potentially be the location of the first successful island-wide eradication.

Further work would be beneficial to validate whether live bird lures would be more effective than mice and carrion on Cocos Island. Field and laboratory experiments have demonstrated that traps with a live bird as the lure show increased capture rates of large, well-conditioned snakes, as well as longer trap investigation times overall compared to those with mouse lures (Yackel Adams et al. 2019; Klug et al. 2021a; Nafus et al. 2021). Additionally, a pattern of feeding success on birds may reduce a brown treesnake's interest in rodents and this may be particularly true on an island where rodents were eradicated in 2009. Verification of the disinterest of Cocos Island snakes to all potential attractant-based lures would be an important next step.

In the event of future invasions, prospects for complete removal of brown treesnakes from larger, prey-rich islands with difficult-to-access terrain would be challenging, particularly when our results indicate that application of the newly developed landscapescale aerial baiting technology would be ineffective (Siers et al. 2020b). Work to differentiate between the context dependency of control tool attraction, based on total prey availability or species compositions of prey, can also be informative in amending current interdiction programmes or developing emergency response protocols if an incipient population of brown treesnakes is located on another island.

Conclusions

Our results indicate that standard invasive brown treesnake control tools, acetaminophen baiting and trapping with mouse lures, are seemingly not effective enough to warrant significant investment of limited project resources where preferred alternative prey are abundant. These results have profound ramifications for the potential of rapid removal and eradication of incipient brown treesnake populations on any other islands at risk of invasion, such as the Hawaiian Islands, the Northern Mariana Islands and throughout Micronesia and the rest of the Pacific where snake-free islands are rich in diversity and density of potential prey (e.g. Lardner et al. (2019a), Table 1). This work underscores the benefits of a continued emphasis on interdiction – prevention of accidental translocation through strong pre- and post-border inspections and reduction of potential stowaways in high-risk areas on Guam – over reliance on early detection and rapid response to resolve any new brown treesnake invasions that might occur. These results are also important for consideration of prevention, early detection, rapid response, suppression and/or eradication of other invasive snakes on islands or elsewhere. Similar issues are likely to be faced during any invasive snake removal programme, particularly where abundant alternative prey is available.

Acknowledgements

We'd like to thank all additional participants in the field work associated with these studies, including Jordan Barcinas, Ahmi Cacapit, Juan-Carlos Mungaray, Ella Norris and Alyssa Taitano. Zach Quiogue, Elizabeth Frasch, Amn Nacpil, Brianna Mont-gomery, Martin Felisan, Shiho Koike, Thomas Fies, Scott Goetz, Levi Gray, Thomas Hinkle, Charlene Hopkins, Peter Xiong, Marijoy Viernes, Dusty Jordan and Karen Watson contributed to the USGS RRT efforts to remove BTS from Cocos Island mentioned in this report. Olympia Terral and Martin Kastner coordinated the volunteer brown treesnake capture efforts. The contracted support from the University of Guam was facilitated by Adrian Ares, Daniel Lindstrom, Aubrey Moore and Cathleen Moore-Linn. We also wish to thank all participants in Cocos Island eradication planning, permitting and environmental compliance consultation including Michelle Bogardus, Dawn Bruns, Jacqueline Flores, Jeffrey Flores, Thomas Hall, Steve Hanser, Martin Kastner, MaryJo Mazurek, Benton Pang, Jeff Quitigua, Haldre Rogers, Jason Suckow, Olympia Terral and Lorena Wada. This research and the USGS rapid response was made possible by funding from the U.S. Department of the Interior Office of Insular Affairs coordinated by Hailey McCoy. Any use of trade, firm or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

References

- Aguon CF, Campbell EW III, Morton JM (2002) Efficacy of electrical barriers used to protect Mariana crow nests. Wildlife Society Bulletin 30(3): 703–708. https://doi. org/10.7591/9781501737688-049
- Amburgey SM, Yackel Adams AA, Gardner B, Hostetter NJ, Siers SR, McClintock BT, Converse SJ (2021) Evaluation of camera trap-based abundance estimators for unmarked populations. Ecological Applications 31(7): e02410. https://doi.org/10.1002/eap.2410
- Avery ML, Tillman EA, Savarie P (2004) Responses of captive fish crows (*Corvus ossifragus*) to acetaminophen baits and bait stations for brown tree snake (*Boiga irregularis*) control on Guam. Bird Behaviour 16(1): 1–6.
- Barnhart PD, Quiogue Z, Frasch ER, Vice D, Hopkins CB, Yackel Adams AA, Reed RN, Nafus MG (2022) Boiga irregularis (Brown Treesnake). Herpetological Review 53(3): 444–445.
- Boback SM, Nafus MG, Yackel Adams AA, Reed RN (2020) Use of visual surveys and radiotelemetry reveals sources of detection bias for a cryptic snake at low densities. Ecosphere 11(1): e03000. https://doi.org/10.1002/ecs2.3000
- BTSTWG (2015) Brown Treesnake strategic plan. Brown Treesnake Technical Working Group. November 2015, Agana, Guam, 86 pp.
- Bushar LM, Reynolds RG, Tucker S, Pace LC, Lutterschmidt WI, Odum RA, Reinert HK (2015) Genetic characterization of an invasive *Boa constrictor* population on the Caribbean island of Aruba. Journal of Herpetology 49(4): 602–610. https://doi.org/10.1670/14-059
- Campbell EW III, Yackel Adams AA, Converse SJ, Fritts TH, Rodda GH (2012) Do predators control prey species abundance? An experimental test with brown treesnakes on Guam. Ecology 93(5): 1194–1203. https://doi.org/10.1890/11-1359.1
- Capinha C, Seebens H, Cassey P, García-Díaz P, Lenzner B, Mang T, Moser D, Pyšek P, Rödder D, Scalera R, Winter M, Dullinger S, Essl F (2017) Diversity, biogeography and the global flows of alien amphibians and reptiles. Diversity & Distributions 23(11): 1313–1322. https://doi.org/10.1111/ddi.12617
- Caves EM, Jennings SB, Hille Ris Lambers J, Tewksbury J, Rogers HS (2013) Natural experiment demonstrates that bird loss leads to cessation of dispersal of native seeds from intact to degraded forests. PLoS ONE 8(5): e65618. https://doi.org/10.1371/journal. pone.0065618
- Chiszar D (1990) The behavior of the brown treesnake: a study in applied comparative psychology. In: Dewsbury DA (Ed.) Contemporary Issues in Comparative Psychology. Sinauer Associates, Inc., Sunderland, 101–123. https://doi.org/10.1037/11525-005

- Christy MT, Yackel Adams AA, Rodda GH, Savidge JA, Tyrrell CL (2010) Modelling detection probabilities to evaluate management and control tools for an invasive species. Journal of Applied Ecology 47(1): 106–113. https://doi.org/10.1111/j.1365-2664.2009.01753.x
- Christy MT, Savidge JA, Yackel Adams AA, Gragg JE, Rodda GH (2017) Experimental landscape reduction of wild rodents increases movements in the invasive brown treesnake (*Boi-ga irregularis*). Management of Biological Invasions: International Journal of Applied Research on Biological Invasions 8(4): 455–467. https://doi.org/10.3391/mbi.2017.8.4.01
- Clark L, Savarie PJ (2012) Efficacy of aerial broadcast baiting in reducing brown treesnake numbers. Human-Wildlife Interactions 6: 212–221. https://www.jstor.org/stable/24874095
- Clark L, Savarie PJ, Shivik JA, Breck SW, Dorr BS (2012) Efficacy, effort, and cost comparisons of trapping and acetaminophen baiting for control of brown treesnakes on Guam. Human-Wildlife Interactions 6: 222–236. https://www.jstor.org/stable/24874096
- Clark L, Clark C, Siers S (2018) Brown Treesnakes: Methods and approaches for control. In: Pitt WC, Beasley JC, Witmer GW (Eds) Ecology and Management of Terrestrial Vertebrate Invasive Species in the United States. CRC Press, Boca Raton, 107–134. https://doi. org/10.1201/9781315157078
- Clout MN, Williams PA (2009) Invasive species management: a handbook of principles and techniques. Oxford University Press, Oxford. https://doi.org/10.1093/ oso/9780199216321.001.0001
- Deso G, Probst JM (2007) *Lycodon aulicus* Linnaeus, 1758 et son impact sur l'herpétofaune insulaire à La Réunion (Ophidia: Colubridae: Lycodontinae). Bulletin Phaeton 25: 29–36.
- DIISE (2023) The Database of Island Invasive Species Eradications: developed by Island Conservation, Coastal Conservation Action Laboratory UCSC, IUCN SSC Invasive Species Specialist Group, University of Auckland and Landcare Research, New Zealand. http:// diise.islandconservation.org
- Dorr BS, Clark CS, Savarie PJ (2016) Aerial application of acetaminophen-treated baits for control of brown tree snakes (RC-200925; NWRC Study Number: QA-1828). Final Report, US-Department of Defense ESTCP.
- Durso AM, Willson JD, Winne CT (2011) Needles in haystacks: Estimating detection probability and occupancy of rare and cryptic snakes. Biological Conservation 144(5): 1508– 1515. https://doi.org/10.1016/j.biocon.2011.01.020
- Engeman RM, Linnell MA (1998) Trapping strategies for deterring spread of brown tree snakes from Guam. Pacific Conservation Biology 4: 348–353. https://doi.org/10.1071/ PC980348
- Engeman RM, Shiels AB, Clark CS (2018) Objectives and integrated approaches for the control of brown tree snakes: An updated overview. Journal of Environmental Management 219: 115–124. https://doi.org/10.1016/j.jenvman.2018.04.092
- Fosberg R (1960) The vegetation of Micronesia 1. General descriptions, the vegetation of the Marianas Islands, and a detailed consideration of the vegetation of Guam. Bulletin of the American Museum of Natural History 119: 1–1.
- Freedman MG, Miller RH, Rogers HS (2018) Landscape-level bird loss increases the prevalence of honeydew-producing insects and non-native ants. Oecologia 188(4): 1263–1272. https://doi.org/10.1007/s00442-018-4273-5

- Fricke EC, Tewksbury JJ, Rogers HS (2014) Multiple natural enemies cause distance-dependent mortality at the seed-to-seedling transition. Ecology Letters 17(5): 593–598. https:// doi.org/10.1111/ele.12261
- Fritts TH (1988) The Brown Tree Snake, *Boiga irregularis*, a Threat to Pacific Islands. US Department of Interior, Fish and Wildlife Service, Research and Development, Honolulu, 36 pp.
- Fritts TH (2002) Economic costs of electrical system instability and power outages caused by snakes on the island of Guam. International Biodeterioration & Biodegradation 49(2–3): 93–100. https://doi.org/10.1016/S0964-8305(01)00108-1
- Fritts TH, McCoid MJ (1991) Predation by the brown tree snake, *Boiga irregularis*, on poultry and other domesticated animals in Guam. Snake 23: 75–80.
- Fritts TH, Rodda GH (1998) The role of introduced species in the degradation of island ecosystems: A case history of Guam. Annual Review of Ecology and Systematics 29(1): 113–140. https://doi.org/10.1146/annurev.ecolsys.29.1.113
- Fritts TH, McCoid MJ, Haddock RL (1990) Risks to infants on Guam from bites of the Brown Tree Snake (*Boiga irregularis*). The American Journal of Tropical Medicine and Hygiene 42(6): 607–611. https://doi.org/10.4269/ajtmh.1990.42.607
- Fritts TH, McCoid MJ, Haddock RJ (1994) Symptoms and circumstances associated with bites by the brown tree snake (Colubridae: *Boiga irregularis*) on Guam. Journal of Herpetology 28(1): 27–33. https://doi.org/10.2307/1564676
- Genovesi P (2001) Guidelines for eradication of terrestrial vertebrates: A European contribution to the invasive alien species issue. Convention on the Conservation of European Wildlife and Natural Habitats 24: 1–28.
- Goetz SM, Yackel Adams AA, Siers SR (2020) Validating deployment of aerially-delivered toxic bait cartridges for control of invasive brown treesnakes. Wildlife Society Bulletin 44(3): 617–622. https://doi.org/10.1002/wsb.1106
- Goetz SM, Hileman ET, Nafus MG, Yackel Adams AA, Bryant A, Reed RN, Siers SR (2021) Brown treesnake mortality after aerial application of toxic baits. The Journal of Wildlife Management 85(7): 1507–1514. https://doi.org/10.1002/jwmg.22108
- Gragg JE, Rodda GH, Savidge JA, White GC, Dean-Bradley K, Ellingson AR (2007) Response of brown treesnakes to reduction of their rodent prey. The Journal of Wildlife Management 71(7): 2311–2317. https://doi.org/10.2193/2006-444
- Guam Department of Agriculture (2020) Invasive brown treesnake present on Cocos Island, agencies working to prevent further spread. Press release. Guam Department of Agriculture, Mangilao, 2 pp.
- Hall TC (1996) Operational control of the brown tree snake on Guam. In: Timm R, Crabb AC (Eds) Proceedings of the 17th Vertebrate Pest Conference, March 1996. University of California, Davis, 234–240.
- Johnston JJ, Savarie PJ, Primus TM, Eisemann JD, Hurley JC, Kohler DJ (2002) Risk assessment of an acetaminophen baiting program for chemical control of brown tree snakes on Guam: Evaluation of baits, snake residues, and potential primary and secondary hazards. Environmental Science & Technology 36(17): 3827–3833. https://doi.org/10.1021/ es015873n

- Kimball BA, Stelting SA, McAuliffe TW, Stahl RS, Garcia RA, Pitt WX (2016) Development of artificial bait for brown treesnake suppression. Biological Invasions 18(2): 359–369. https://doi.org/10.1007/s10530-015-1031-z
- Klug PE, Yackel Adams AA, Reed RN (2021a) Olfactory lures in predator control do not increase predation risk to birds in areas of conservation concern. Wildlife Research 49(2): 183–192. https://doi.org/10.1071/WR21022
- Klug PE, Yackel Adams AA, Siers SR, Brindock KM, Mazurek MJ, Mosher SM, Pitt WC, Reed RN (2021b) Locally abundant, endangered Mariana Swiftlets impact the abundance, behavior, and body condition of an invasive predator. Oecologia 195(4): 1083–1097. https:// doi.org/10.1007/s00442-021-04876-0
- Kraus F (2009) Alien reptiles and amphibians: A scientific compendium and analysis. Invading Nature-Springer Science & Business Media 4: 1–563. https://doi.org/10.1007/978-1-4020-8946-6
- Kraus F (2015) Impacts from invasive reptiles and amphibians. Annual Review of Ecology, Evolution, and Systematics 46(1): 75–97. https://doi.org/10.1146/annurev-ecolsys-112414-054450
- Lardner B, Savidge JA, Rodda GH, Reed RN (2009) Prey preferences and prey acceptance in juvenile Brown Treesnakes (*Boiga irregularis*). Herpetological Conservation and Biology 4(3): 313–323.
- Lardner B, Yackel Adams AA, Savidge JA, Rodda GH, Reed RN, Clark CS (2013) Effectiveness of bait tubes for Brown Treesnake control on Guam. Wildlife Society Bulletin 37(3): 664–673. https://doi.org/10.1002/wsb.297
- Lardner B, Yackel Adams AA, Savidge JA, Reed RN (2019b) Optimizing walking pace to maximize snake detection rate: A visual encounter survey experiment. Herpetologica 75(3): 218–223. https://doi.org/10.1655/D-18-00020
- Lardner B, Yackel Adams AA, Knox AJ, Savidge JA, Reed RN (2019a) Do observer fatigue and taxon bias compromise visual encounter surveys for small vertebrates? Wildlife Research 46(2): 127–135. https://doi.org/10.1071/WR18016
- Lindberg AC, Shivik JA, Clark L (2000) Mechanical mouse lure for brown treesnakes. Copeia 2000(3): 886–888. https://doi.org/10.1643/0045-8511(2000)000[0886:MMLFBT]2.0. CO;2
- Mackessy SP, Sixberry NM, Heyborne WH, Fritts T (2006) Venom of the Brown Treesnake, *Boiga irregularis*: Ontogenetic shifts and taxa-specific toxicity. Toxicon 47(5): 537–548. https://doi.org/10.1016/j.toxicon.2006.01.007
- Mathies T, Scarpino R, Levine BA, Clark C, Savidge JA (2011) Excluding nontarget species from Brown Tree Snake, *Boiga irregularis* (Reptilia: Colubridae), bait stations: Experimental tests of station design and placement. Pacific Science 65(1): 41–57. https://doi. org/10.2984/65.1.041
- Mauldin RE, Savarie PJ (2010) Acetaminophen as an oral toxicant for Nile monitor lizards (*Varanus niloticus*) and Burmese pythons (*Python molurus bivittatus*). Wildlife Research 37(3): 215–222. https://doi.org/10.1071/WR08168
- Medina S, Aguon C (2000) Establishment of an experimental population of Guam rails on Rota or other Northern Mariana Islands. In: Davis GW, Pitlik TJ, Leberer T, Vice D (Eds)

Annual Report Fiscal Year 2000, Division of Aquatic and Wildlife Resources, Guam Department of Agriculture, Mangilao, 174–183.

- Monzón-Argüello C, Patiño-Martínez C, Christiansen F, Gallo-Barneto R, Cabrera-Pérez MÁ, Peña-Estévez MÁ, López-Jurado LF, Lee PLM (2015) Snakes on an island: Independent introductions have different potentials for invasion. Conservation Genetics 16(5): 1225– 1241. https://doi.org/10.1007/s10592-015-0734-0
- Nafus MG, Yackel Adams AA, Klug PE, Rodda GH (2018) Habitat type and structure affect trap capture success of an invasive snake across variable densities. Ecosphere 9(8): e02339. https://doi.org/10.1002/ecs2.2339
- Nafus MG, Yackel Adams AA, Boback SM, Siers SR, Reed RN (2020) Behavior, size, and body condition predict susceptibility to management and reflect post-treatment frequency shifts in an invasive snake. Global Ecology and Conservation 21: e00834. https://doi. org/10.1016/j.gecco.2019.e00834
- Nafus MG, Xiong PX, Paxton EH, Yackel Adams AA, Goetz SM (2021) Foraging behavior in a generalist snake (brown treesnake, *Boiga irregularis*) with implications for avian reintroduction and recovery. Applied Animal Behaviour Science 243: e105450. https://doi. org/10.1016/j.applanim.2021.105450
- Nafus MG, Siers SR, Levine BA, Quiogue ZC, Yackel Adams AA (2022) Demographic response of brown treesnakes to extended population suppression. The Journal of Wildlife Management 86(1): e22136. https://doi.org/10.1002/jwmg.22136
- Nafus MG, Collins AF, Viernes M, Hopkins C, Nacpil A (2023) Guam, USGS Closed Population (NWFN), an experimental eradication of brown treesnakes in a 5-ha study site, 2016 2023: US Geological Survey data release. https://doi.org/10.5066/P9QRWKQB
- Nelson Jr L, Clark FW (1973) Correction for sprung traps in catch/effort calculations of trapping results. Journal of Mammalogy 54(1): 295–298. https://doi.org/10.2307/1378903
- Perry G, Morton JM (1999) Regeneration rates of the woody vegetation of Guam's Northwest Field following major disturbance: Land use patterns, feral ungulates, and cascading effects of the brown treesnake. Micronesica 32: 125–142.
- Perry G, Vice D (2009) Forecasting the risk of brown tree snake dispersal from Guam: A mixed transport-establishment model. Conservation Biology 23(4): 992–1000. https:// doi.org/10.1111/j.1523-1739.2009.01169.x
- Perry G, Pierce J, Griffin D, van Buurt G, Lazell J (2003) Geographic distribution: *Elaphe guttata guttata*. Herpetological Review 34: e264.
- Pollock HS, Savidge JA, Kastner M, Seibert TF, Jones TM (2019) Pervasive impacts of invasive brown treesnakes drive low fledgling survival in endangered Micronesian Starlings (*Aplonis opaca*) on Guam. The Condor 121(2): duz014. https://doi.org/10.1093/condor/duz014
- Pollock HS, Kastner M, Wiles GJ, Thierry H, Dueñas LB, Paxton EH, Suckow NM, Quitugua J, Rogers HS (2021) Recent recovery and expansion of Guam's locally endangered Såli (Micronesian Starling) *Aplonis opaca* population in the presence of the invasive brown treesnake. Bird Conservation International 32(1): 95–110. https://doi.org/10.1017/S0959270920000726
- R Core Team (2021) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna. https://www.R-project.org/

- Reed RN, Kraus F (2010) Invasive reptiles and amphibians: Global perspectives and local solutions. Animal Conservation 13(s1, Supplement 1): 3–4. https://doi.org/10.1111/j.1469-1795.2010.00409.x
- Reed RN, Tucker AD (2012) Determining age, sex, and reproductive condition. In: McDiarmid RW, Foster MS, Guyer C, Gibbons JW, Chernoff N (Eds) Reptile Biodiversity: Standard Methods for Inventory and Monitoring. University of California Press, Berkeley, 51–63.
- Richmond JQ, Wood DA, Stanford JW, Fisher RN (2015) Testing for multiple invasion routes and source populations for the invasive brown treesnake (*Boiga irregularis*) on Guam: implications for pest management. Biological Invasions 17: 337–349. https://doi.org/10.1007/ s10530-014-0733-y
- Rodda GH, Fritts TH (1992) The impact of the introduction of the colubrid snake *Boiga irregularis* on Guam's lizards. Journal of Herpetology 26(2): 166–174. https://doi. org/10.2307/1564858
- Rodda GH, Reed RN (2007) Size-based trends and management implications of microhabitat utilization by Brown Treesnakes, with emphasis on juvenile snakes. In: Witmer G, Pitt WC, Fagerstone KA (Eds) Managing Vertebrate Invasive Species: Proceedings of an International Symposium, Fort Collins (Colorado), August 2007. USDA/APHIS/WS, National Wildlife Research Center, Colorado, 257–267.
- Rodda GH, Savidge JA (2007) Biology and impacts of Pacific island invasive species. 2. *Boiga irregularis*, the brown tree snake (Reptilia: Colubridae). Pacific Science 61(3): 307–324. https://doi.org/10.2984/1534-6188(2007)61[307:BAIOPI]2.0.CO;2
- Rodda GH, Fritts TH, Chiszar D (1997) The disappearance of Guam's wildlife. Bioscience 47(9): 565–574. https://doi.org/10.2307/1313163
- Rodda GH, McCoid MJ, Fritts TH, Campbell EW (1999a) 17. Population trends and limiting factors in *Boiga irregularis*. In: Rodda GH, Sawai Y, Chiszar D, Tanaka H (Eds) Problem Snake Management: The Habu and the Brown Treesnake. Cornell University Press, Ithaca, 236–254. https://doi.org/10.7591/9781501737688-025
- Rodda GH, Fritts TH, Clark CS, Gotte SW, Chiszar D (1999b) A state-of-the-art trap for the Brown Treesnake. In: Rodda GH, Sawai Y, Chiszar D, Tanaka H (Eds) Problem Snake Management: The Habu and the Brown Treesnake. Cornell University Press, Ithaca, 268– 305. https://doi.org/10.7591/9781501737688-029
- Rodda GH, Savidge JA, Tyrrell CL, Christy MY, Ellingson AR (2007) Size bias in visual searches and trapping of brown treesnakes on Guam. The Journal of Wildlife Management 71(2): 656–661. https://doi.org/10.2193/2005-742
- Rogers HS (2011) The fate of a silent forest: the effects of complete bird loss on the forest of Guam. PHD Dissertation, University of Washington, Seattle, Washington.
- Rogers H (2020) Natural history of a silent forest. Journal of Natural History Education and Experience 14: 22–26.
- Rogers HS, Hille Ris Lambers J, Miller R, Tewksbury JJ (2012) 'Natural experiment' demonstrates top-down control of spiders by birds on a landscape level. PLoS ONE 7(9): e43446. https://doi.org/10.1371/journal.pone.0043446
- Savarie PJ, Clark L (2006) Evaluation of bait matrices and chemical lure attractants for brown tree snakes. In: Timm RM, O'Brien JM (Eds) Proceedings of the 22nd Vertebrate

Pest Conference, October 2006. University of California, Davis, 483–488. https://doi. org/10.5070/V422110077

- Savarie PJ, York DL, Hurley JC, Volz S (2000) Testing the dermal and oral toxicity of selected chemicals to brown treesnakes. In: Salmon TP, Crabb AC (Eds) Proceedings of the 19th Vertebrate Pest Conference, March 2009. University of California, Davis, 139–145. https://doi.org/10.5070/V419110219
- Savarie PJ, Shivik JA, White GC, Hurley JC, Clark L (2001) Use of acetaminophen for largescale control of brown treesnakes. The Journal of Wildlife Management 65(2): 356–365. https://doi.org/10.2307/3802916
- Savidge JA (1987) Extinction of an island forest avifauna by an introduced snake. Ecology 68(3): 660–668. https://doi.org/10.2307/1938471
- Savidge JA (1988) Food habits of *Boiga irregularis*, an introduced predator on Guam. Journal of Herpetology 22(3): 275–282. https://doi.org/10.2307/1564150
- Shivik JA (1998) Brown treesnake response to visual and olfactory cues. The Journal of Wildlife Management 62(1): 105–111. https://doi.org/10.2307/3802268
- Shivik JA (1999) Carrion, context and lure development: The relative importance of sensory modalities to foraging brown tree snakes (*Boiga irregularis*). PHD Disseretation. Colorado State University, Fort Collins, Colorado.
- Shivik JA, Clark L (1997) Carrion seeking in brown tree snakes: Importance of olfactory and visual cues. The Journal of Experimental Zoology 279(6): 549–553. https://doi. org/10.1002/(SICI)1097-010X(19971215)279:6<549::AID-JEZ2>3.0.CO;2-N
- Shivik JA, Clark L (1999) Ontogenetic shifts in carrion attractiveness to brown tree snakes (*Boiga irregularis*). Journal of Herpetology 33(2): 334–336. https://doi.org/10.2307/1565737
- Shivik JA, Bourassa J, Donnigan SN (2000) Elicitation of brown treesnake predatory behavior using polymodal stimuli. The Journal of Wildlife Management 64(4): 969–975. https:// doi.org/10.2307/3803206
- Shwiff SA, Gebhardt K, Kirkpatrick KN, Shwiff SS (2010) Potential economic damage from introduction of Brown Tree Snakes, *Boiga irregularis* (Reptilia: Colubridae), to the islands of Hawai'i. Pacific Science 64(1): 1–10. https://doi.org/10.2984/64.1.001
- Siers SR (2015) Microgeographic and ontogenetic variability in the ecology of invasive Brown Treesnakes on Guam, and effects of roads on their landscape-scale movements. PHD Thesis. Colorado State University, Fort Collins, Colorado.
- Siers SR (2021) Allometric regression of snake body length from head image measurements. Wildlife Society Bulletin 45(3): 538–545. https://doi.org/10.1002/wsb.1213
- Siers SR, Savidge JA, Demeulenaere E (2017a) Restoration Plan for the Habitat Management Unit, Naval Support Activity Andersen, Guam. Prepared for Naval Facilities Engineering Command. Colorado State University, Fort Collins, 238 pp.
- Siers SR, Savidge JA, Reed RN (2017b) Quantile regression of microgeographic variation in population characteristics of an invasive vertebrate predator. PLoS ONE 12(6): e0177671. https://doi.org/10.1371/journal.pone.0177671
- Siers SR, Yackel Adams AA, Reed RN (2018a) Behavioral differences following ingestion of large meals and consequences for management of a harmful invasive snake: A field experiment. Ecology and Evolution 8(20): 10075–10093. https://doi.org/10.1002/ece3.4480

- Siers SR, Dorr BS, Shiels AB, Chlarson FM, Macaoay LG, Mundo RM, Rabon JAB, Volsteadt RM, Hall MA, Clark CS, Mosher SM, Savarie PJ (2018b) Assessment of Brown Treesnake activity and bait take following large-scale snake suppression in Guam. USDA, APHIS, WS, National Wildlife Research Center Final Report QA-2438: 1–10.
- Siers SR, Pitt WC, Eisemann JD, Clark L, Shiels AB, Clark CS, Gosnell RJ, Messaros MC (2019a) An automated bait manufacturing and aerial delivery system for landscape-scale control of invasive Brown Treesnakes on Guam. In: Veitch CR, Clout MN, Martin AR, Russell JC, West CJ (Eds) Island invasives: Scaling up to meet the challenge, Occasional Paper SSC 62. Gland, 348–355. https://doi.org/10.2305/IUCN.CH.2019.SSC-OP.62.en
- Siers SR, Shiels AB, Payne CG, Chlarson FM, Clark CS, Mosher SM (2019b) Photographic validation of target versus nontarget take of brown treesnake baits. Wildlife Society Bulletin 43(4): 752–759. https://doi.org/10.1002/wsb.1023
- Siers SR, Shiels AB, Barnhart PD (2020a) Invasive snake activity before and after automated aerial baiting. The Journal of Wildlife Management 84(2): 256–267. https://doi. org/10.1002/jwmg.21794
- Siers SR, Eisemann JD, Pitt WC, Clark L, Goetz SM, Gosnell RJ, Collins AF, Hileman ET, Nafus MG, Yackel Adams AA, Messaros MC, Coon WGN (2020b) Automated aerial baiting for invasive Brown Treesnake control: System overview and program status. In: Woods DM (Ed.) Proceedings of the 29th Vertebrate Pest Conference, Santa Barbara, California, March 2020: 1–7.
- Siers SR, Goetz SM, Volsteadt RM, Nafus MG (2021) Evaluating lethal toxicant doses for the largest individuals of an invasive vertebrate predator with indeterminate growth. Management of Biological Invasions: International Journal of Applied Research on Biological Invasions 12(2): 476–494. https://doi.org/10.3391/mbi.2021.12.2.17
- Silva-Rocha I, Salvi D, Sillero N, Mateo JA, Carretero MA (2015) Snakes on the Balearic Islands: An invasion tale with implications for native biodiversity conservation. PLoS ONE 10(4): e0121026. https://doi.org/10.1371/journal.pone.0121026
- Stanford JW, Rodda GH (2007) The Brown Treesnake Rapid Response Team. In: Witmer G, Pitt WC, Fagerstone KA (Eds) Managing Vertebrate Invasive Species: Proceedings of an International Symposium, Fort Collins (Colorado), August 2007. USDA/APHIS/WS, National Wildlife Research Center, Colorado, 175–217.
- Tonge S (1990) The past, present and future of the herpetofauna of Mauritius. Bulletin of the Chicago Herpetological Society 25: 220–226.
- Tyrrell CL, Christy MT, Rodda GH, Yackel Adams AA, Ellingson AR, Savidge JA, Dean-Bradley K, Bischof R (2009) Evaluation of trap capture in a geographically closed population of brown tree snakes on Guam. Journal of Applied Ecology 46(1): 128–135. hhttps://doi. org/10.1111/j.1365-2664.2008.01591.x
- U.S. Geological Survey (2023) Cocos Island, Guam Brown Treesnake rapid response visual survey and capture data, 10/2020–05/2023. U.S. Geological Survey data release. https:// doi.org/10.5066/P9MT1JNO
- USDA Wildlife Services (2009) Cocos Island biosecurity plan. Prepared by U. S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services. Barrigada, Guam.

- USDA Wildlife Services (2021a) Biological Assessment for invasive Brown Treesnake eradication to protect natural resources on Cocos Island, Guam. U.S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services. Barrigada, Guam.
- USDA Wildlife Services (2021b) Cocos Island, Guam Brown Treesnake eradication. Draft Environmental Assessment. U.S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services. Barrigada, Guam.
- USFWS (2019) Species report for Slevin's Skink (*Emoia slevini*) Final draft. U. S. Fish and Wildlife Service, Pacific Islands Fish and Wildlife Office. Honolulu.
- van den Hurk P, Kerkkamp HM (2019) Phylogenetic origins for severe acetaminophen toxicity in snake species compared to other vertebrate taxa. Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology 215: 18–24. https://doi.org/10.1016/j. cbpc.2018.09.003
- Vice DS, Engeman RM, Vice DL (2005) A comparison of three trap designs for capturing brown treesnakes on Guam. Wildlife Research 32(4): 355–359. https://doi.org/10.1071/ WR04046
- Weijola V, Vahtera V, Koch A, Schmitz A, Kraus F (2020) Taxonomy of Micronesian monitors (Reptilia: Squamata: Varanus): endemic status of new species argues for caution in pursuing eradication plans. Royal Society Open Science 7(5): e200092. https://doi.org/10.1098/ rsos.200092
- Wiles GJ (1987) The status of fruit bats on Guam. Pacific Science 41: 1-4.
- Wiles GJ, Bart J, Beck Jr RE, Aguon CF (2003) Impacts of the brown tree snake: Patterns of decline and species persistence in Guam's avifauna. Conservation Biology 17(5): 1350– 1360. https://doi.org/10.1046/j.1523-1739.2003.01526.x
- Yackel Adams AA, Lardner B, Knox AJ, Reed RN (2018) Inferring the absence of an incipient population during a rapid response for an invasive species. PLoS ONE 13(9): e0204302. https://doi.org/10.1371/journal.pone.0204302
- Yackel Adams AA, Nafus MG, Klug PE, Lardner B, Mazurek MJ, Savidge JA, Reed RN (2019) Contact rates with nesting birds before and after invasive snake removal: Estimating the effects of trap-based control. NeoBiota 49: 1–17. https://doi.org/10.3897/neobiota.49.35592
- Yackel Adams AA, Barnhart PD, Rodda GH, Hileman ET, Nafus MG, Reed RN (2021) Can we prove that an undetected species is absent? Evaluating whether brown treesnakes are established on the island of Saipan using surveillance and expert opinion. Management of Biological Invasions: International Journal of Applied Research on Biological Invasions 12(4): 901–926. https://doi.org/10.3391/mbi.2021.12.4.09

RESEARCH ARTICLE



Asymmetrical insect invasions between three world regions

Rylee Isitt¹, Andrew M. Liebhold^{2,3}, Rebecca M. Turner⁴, Andrea Battisti⁵, Cleo Bertelsmeier⁶, Rachael Blake⁷, Eckehard G. Brockerhoff^{8,9}, Stephen B. Heard¹, Paal Krokene¹⁰, Bjørn Økland¹⁰, Helen F. Nahrung¹¹, Davide Rassati⁵, Alain Roques¹², Takehiko Yamanaka¹³, Deepa S. Pureswaran¹⁴

I Department of Biology, University of New Brunswick, Fredericton, NB, Canada 2 US Forest Service, Northern Research Station, 180 Canfield St., Morgantown, WV, USA 3 Faculty of Forestry and Wood Sciences, Czech University of Life Sciences, Prague, Czech Republic 4 Scion (New Zealand Forest Research Institute), 10 Kyle St., Riccarton, Christchurch, New Zealand 5 Department of Agronomy, Food, Natural Resources, Animals and the Environment, University of Padova, Padova, Legnaro, Italy 6 Department of Ecology and Evolution, University of Lausanne, Lausanne, Switzerland 7 Intertidal Agency, Oakland, CA, USA 8 Swiss Federal Research Institute WSL, Birmensdorf, Zürich, Switzerland 9 School of Biological Sciences, University of Canterbury, Christchurch, New Zealand 10 Division of Biotechnology and Plant Health, Norwegian Institute of Bioeconomy Research, 1431, Ås, Norway 11 Forest Research Institute, University of the Sunshine Coast, Sippy Downs, Queensland, Australia 12 French National Institute for Agriculture, Food, and Environment (INRAE), Unité de Recherches Zoologie Forestière, Orléans, France 13 Research Center for Agricultural Information Technology, National Agriculture and Food Research Organization (NARO), Tsukuba, Ibaraki 305-0856, Japan 14 Natural Resources Canada, Canadian Forest Service, Atlantic Forestry Centre, 1350 Regent St., Fredericton, NB, Canada

Corresponding author: Rylee Isitt (risitt@protonmail.com)

Academic editor: V. Lantschner | Received 11 August 2023 | Accepted 27 November 2023 | Published 8 January 2024

Citation: Isitt R, Liebhold AM, Turner RM, Battisti A, Bertelsmeier C, Blake R, Brockerhoff EG, Heard SB, Krokene P, Økland B, Nahrung HF, Rassati D, Roques A, Yamanaka T, Pureswaran DS (2024) Asymmetrical insect invasions between three world regions. NeoBiota 90: 35–51. https://doi.org/10.3897/neobiota.90.110942

Abstract

The geographical exchange of non-native species can be highly asymmetrical, with some world regions donating or receiving more species than others. Several hypotheses have been proposed to explain such asymmetries, including differences in propagule pressure, source species (invader) pools, environmental features in recipient regions, or biological traits of invaders. We quantified spatiotemporal patterns in the exchange of non-native insects between Europe, North America, and Australasia, and then tested possible explanations for these patterns based on regional trade (import values) and model estimates of invader pool sizes. Europe was the dominant donor of non-native insect species between the three regions, with

most of this asymmetry arising prior to 1950. This could not be explained by differences in import values (1827–2014), nor were there substantial differences in the sizes of modelled invader pools. Based on additional evidence from literature, we propose that patterns of historical plant introductions may explain these asymmetries, but this possibility requires further study.

Keywords

International trade, non-native insects, plants, propagule pressure, species pools

Introduction

Non-native insects have been implicated in displacing native species, altering the composition of ecological communities, damaging economically important trees and food crops, vectoring diseases, and more (Kenis et al. 2009; Bradshaw et al. 2016). An intriguing aspect of insect invasions is that some regions appear to have donated disproportionately more non-native insects during biotic exchange than others. For example, considerably more phytophagous forest insects have invaded North America from Europe than the reverse (Niemelä and Mattson 1996), and Europe has contributed a large fraction of New Zealand's non-native insect fauna (Edney-Browne et al. 2018). Consequently, the question of why such asymmetries may occur has fascinated ecologists for decades, with several mutually compatible hypotheses offered: (1) differences in the magnitude of invasion vectors, such as international trade, may lead to differences in the arrival and establishment rates of non-native species; (2) differences in the size of potential invader pools may drive differences in the numbers of species being donated to other regions; (3) environmental differences (e.g., climate and availability of host plants) in recipient regions may promote or inhibit invasion; and (4) biological traits of insects native to some regions may make them better at invading or competing than insects native to other regions (Vermeij 1991, 1996; Niemelä and Mattson 1996; Visser et al. 2016).

The latter two hypotheses are often tested on a single insect order or guild and at smaller spatial scales (e.g., Rigot et al. 2014; Guyot et al. 2015; Rassati et al. 2016), but less commonly on multiple insect orders and multiple geographical regions. Testing them requires regional knowledge of the nature of recipient environments and their ecological communities, and of the biological traits of the invaders, information that is often available only for certain regions or certain insect groups/guilds. The former two hypotheses are more approachable, given the availability of datasets on international trade, regional insect richness, and modelling approaches that can estimate invader pool sizes.

Our research goals were firstly to test for the existence of asymmetries in the cumulative numbers of insect invaders, across all taxa, exchanged between three world regions of interest: North America, Europe, and Australasia (limited to Australia and New Zealand). These regions were chosen due to their histories of anthropogenic interactions and exchange of species, existing literature suggesting asymmetrical exchange of insects between them (see above), and the availability of data. Secondly, if clear
asymmetries were found, we aimed to determine if they could be explained by differences in propagule pressure (using the value of international trade as a proxy) or by differences in estimates of invader pool sizes. We did not statistically test hypotheses (3) and (4), above, but considered them as possible explanations for asymmetries that could not be explained by hypotheses (1) and (2).

Methods

Datasets and world regions

Insect establishment data were based on the International Non-native Insect Establishment database (Turner et al. 2021), supplemented by several other online datasets (Seebens et al. 2017; Nahrung and Carnegie 2020; Liebhold et al. 2021; GBIF.org 2022; Mally et al. 2022). We used an automated taxonomic cleaning script (Blake and Turner 2021) using the GBIF (GBIF.org 2022) API to standardize species names (merge synonyms and correct misspellings). The resulting dataset contained dated records of non-native insect discoveries, the species identity (order, family, genus, and species), the region/nation in which the species was discovered, the native biogeographic range of the species, and other data such as whether the introduction was deliberate, if the species was found only indoors (e.g., greenhouses), and if the species is herbivorous.

Our choices of world regions and their spatial extents were constrained by the available data. We used a subset of the establishment database that allowed us to compare the reciprocal flows of insects between donor and recipient regions. The only regions that could be compared in this way were North America (NA), Europe (EU), and Australia and New Zealand combined into an Australasian region (AU). Due to spatial gaps in these data, there were minor mismatches in the spatial extents of these regions depending on context. For example, as a donor region, Australasia included Papua New Guinea, but as a recipient region, it only included Australia and New Zealand because we did not have non-native insect discovery records for Papua New Guinea. In this case, correcting for this mismatch would require estimating the number of insects from North America and Europe that have established into Papua New Guinea, and excluding species that also established into Australia or New Zealand. Since the spatial mismatches were relatively minor, and such corrections would themselves be prone to error, we opted not to attempt corrections.

For all analyses, we excluded discovery records where: (1) species had native ranges spanning multiple biogeographic regions (e.g., Holarctic or cosmopolitan species); (2) the native ranges and establishment regions were the same (indicating species that spread within these regions); (3) the establishment was limited to "indoors" (e.g., greenhouses); or (4) the establishment was a result of intentional introduction. This left us with a dataset of 2,324 non-native insect discovery records across the six pairwise flows between North America, Europe, and Australasia, with the dated records spanning 1617–2021.

Regional import value data were obtained from the TradeHist database (Fouquin and Hugot 2016), modified with modern ISO-3 country codes. The TradeHist database describes the annual value of trade goods from 1827–2014 in British pounds sterling (not corrected for inflation) flowing from origin to destination countries. The database does not include details on trade volume/frequency or commodity type. We corrected all trade values for inflation relative to 2020 based on the annual percent change of the UK retail prices index (Office for National Statistics 2021). We grouped the origin and destination countries into the same regions as above (North America, Europe, and Australasia), with some minor unavoidable differences where national borders did not follow biogeographic boundaries. From these groupings of countries, we created a subset of the TradeHist database representing the six pairwise flows between North America, Europe, and Australasia by summing annual trade value across all countries within each region. Records of trade between countries within each of the resulting biogeographic regions were dropped.

Testing for asymmetries and temporal variation in establishment rates

To test for invasional asymmetries, we tallied the number of first discoveries of nonnative insects for each of the six pairwise flows between North America, Europe, and Australasia. We further split these cumulative counts by insect order and (separately) by herbivory (herbivores vs non-herbivores). We used G-tests (log-likelihood ratio goodness-of-fit tests) to compare these counts between each donor/recipient pair, separately for each order and herbivory category (e.g., one test for the counts of Coleoptera exchanged in both directions between Europe and North America, another for Hemiptera, etc.), with the null hypothesis being equal numbers of insects exchanged in each direction. We adjusted the *P*-values for multiple comparisons across orders and herbivory categories using the Holm-Bonferroni procedure. To visualize temporal variation in the establishment rates of insects over each flow, we plotted cumulative discoveries versus cumulative import values following Levine and D'Antonio (2003).

Testing for the effects of differential trade and invader pool sizes

To determine if asymmetries in non-native insect establishments between regions could be explained by unequal trade or invader pools, we adapted Poisson process models from Costello et al. (2007) and Morimoto et al. (2019) in which the number of annually established insects flowing from donor to recipient regions were proportional to annual import values and model-fit invader pool sizes. Because the Poisson-process models required dated annual discovery and import values for each observation, this limited our establishment records to the date range of the TradeHist database. This left us with 1,872 dated establishment records (~80% of the establishment dataset) with corresponding import values from 1827–2014.

Our models estimated the lag between establishments and discoveries, predicting the annual establishments necessary to fit to observed discoveries given the lag estimates. This was done to address concerns over records of first discovery being poor proxies for the actual timing of establishments given the extended lag frequently occurring between establishment and discovery (Costello and Solow 2003). As discovery probability depends in large part on discovery effort, this allows our models to (indirectly) account for overall differences in discovery effort between flows.

To account for the possibility of "saturation" (depletion of invader pools) that might gradually reduce establishment rates, we used AIC-based model selection to choose between models which included or omitted a rate-limiting component based on the observed number of cumulative discoveries compared to a predicted maximum. All models contained an 'annual establishment rate' parameter (r) representing the number of non-native insects per billion pounds sterling of imports prior to any depletion of invader pools. If differences in import values fully explained asymmetries in non-native insect establishments, we would expect no significant differences between reciprocal flows in the value of r.

We omitted an intercept term in our models, forcing them to account for all establishments as a function of imports. We modelled the gradual depletion of invader pools as a non-linear rate-limiting factor based on the idea that early invaders are more likely the best or most numerous invaders, leading to a rapid initial decrease in the probability of establishment per unit of propagule pressure (Liebhold et al. 2017). These modifications were necessary to produce good fits to our data – initial attempts to use the same models as in Morimoto et al. (2019) resulted in nonsensical parameter estimations and poor fits in most cases. Our full model was:

$$\lambda_{t} = rv_{t}s_{t}$$

$$s_{t} = \left(1 - \frac{d_{c,t}}{d_{sat}}\right)^{2}$$

$$N_{t} \sim \text{Poisson}\left(\delta_{t} = \sum_{j=1827}^{t} \lambda_{j}p_{t}\right)$$

$$p_{j,t} = \pi (1 - \pi)^{t-j},$$
(1)

where:

 λ_t is the predicted number of new non-native establishments in year *t*,

r is the number of species established per billion pounds sterling (prior to saturation),

 v_t is the value of imports (2020 billion pounds sterling) in year t,

*s*_{*t*} is a rate-limiting factor of interval [0,1] which approaches 0 as the cumulative number of species discoveries approaches a predicted maximum,

 $d_{c,t}$ is the (observed) cumulative species discovered by year *t*, d_{sat} is the number of discoveries after which new establishments cease (saturate), N_t is the actual number of non-native discoveries in year *t*, δ_t is the predicted number of non-native discoveries in year *t*, $p_{j,t}$ is the probability that a species which established in year *j* will be discovered in year *t*, and π is the annual probability of discovery. The cumulative sum of discoveries $(d_{c,t})$ was calculated by summing the number of annual discoveries from the first year of records (1827) to year *t*, inclusive. We used the sum of discoveries instead of establishments for modelling the saturation of species pools because discovery sums could be easily calculated from the original data. The main drawback to this technique was that it slightly complicated the interpretation of the saturation parameter (d_{sat}) : rather than being a direct prediction of the invader pool size, it was the predicted number of cumulative discoveries at the time of full depletion of the invader pool.

We fit the models to observed annual discoveries (N_p) for each combination of donor and recipient region, minimizing the maximum likelihood as described by Morimoto et al. (2019):

$$L(r, d_{sat}, \pi) \propto \prod_{j=1827+\tau}^{t} \exp(-\delta_j) \delta_j^{N_j}$$
(2)

where $\tau = 20$ as "preservation years" to prevent fitting the model to species that established prior to 1827 (the first year of discovery records in our database) but were discovered after 1827. Without these "preservation years", δ_t (the predicted number of discoveries in year *t*) may be underestimated near the start of the dataset because there will be a lack of prior years of predicted establishments from which to model the lagged discoveries (Morimoto et al. 2019). We also used a reduced model which omitted the depletion of invader pools from Eq. (1), effectively making s_t a constant with a value of 1. We then removed the associated parameter (d_{sat}) from Eq. (2). This "without saturation" model was otherwise identical to the full model.

For parameter estimation, we set lower and (in a few cases) upper bounds on each parameter using the Limited-memory Broyden–Fletcher–Goldfarb–Shanno algorithm (L-BFGS-B) method (Byrd et al. 1995). We bounded the rate of establishments (r)to ≥ 0.005 non-native species per billion pounds sterling of imports. This was done to prevent model optimization from testing ecologically nonsensical parameter estimates (negative or zero species introductions per billion pounds sterling of imports); 0.005 was chosen to be well below the initial slopes of the curves of cumulative non-native species versus cumulative import values. We bounded the annual probability of discovery (π) to between 0.0125 and 0.95 (corresponding to 1.05–80 years of discovery lag), which was chosen as an ecologically reasonable range based on discovery lag estimates from prior publications (Morimoto et al. 2019; MacLachlan et al. 2021). If the lag estimates were left unbounded, the models typically failed to converge. For the saturation term (d_{ext}) , we set the lower bound to the cumulative numbers of observed, dated discoveries of non-native insects (since we know the invader pool must be at least this large). This lower bound varied for each of the six flows (1121 for EU to NA, 205 for NA to EU, 349 for EU to AU, 70 for AU to EU, 74 for NA to AU, and 53 for AU to NA). We did not impose an upper bound on the estimates of d_{sat} . We fit both the full and reduced models (the latter lacking the saturation term) to each flow and selected the one with the lowest Akaike information criterion (AIC) value.

To determine if asymmetries may be explained by differences in the size of invader pools, we compared 95% confidence intervals of the predicted numbers of non-native

insect discoveries after full depletion of the invader pool (d_{sat}) between region pairs resulting from our Poisson process models. This was only done when full models (including terms for finite invader pools) were selected for both directions between region pairs. Additionally, we compiled counts of described native insects in each of the three regions for qualitative comparisons to the magnitude of insect invasions across the six flows.

We used the *R* function *optim* for parameter estimation in the Poisson process models (R Core Team 2021). The confidence intervals were approximately calculated using the inverse of the Hessian matrix evaluated at the last iteration in the optimization process. For parameters with lower or upper bounds, we truncated the confidence intervals to the parameter estimation boundaries. All analyses were performed in R 4.1.0 (R Core Team 2021). Model predictions (cumulative annual establishments and discoveries) were included on the plots of cumulative discoveries versus cumulative trade.

Results

Europe has donated approximately six times more non-native insect species to North America and Australasia than it has received from these regions (Fig. 1).

Asymmetries in the reciprocal flows of non-native insects between Europe and North America and between Europe and Australasia were highly significant in total species, across the five largest insect orders, and among both herbivores and non-herbivores (all p < 0.001; Table 1). There were no significant asymmetries in the numbers of non-native insects exchanged between North America and Australasia (all p > 0.05; Table 1).

Plots of cumulative insect establishments versus cumulative import values over time show that the European asymmetry developed quickly and early (Fig. 2). In

Table 1. Counts of non-native insect species discovered for each of the six pairwise flows between North America (NA), Europe (EU), and Australasia (AU), by taxonomic order, herbivory, and sum totals. Col. = Coleoptera, Hem. = Hemiptera, Hym. = Hymenoptera, Lep. = Lepidoptera, Dip. = Diptera. The G statistic was computed to test the null hypothesis of no difference in the number of species exchanged in each direction between a given pair of world regions, separately for each column. We used the Holm-Bonferroni method to control for multiple comparisons across orders and herbivory.

Flow	Order				Herbivory		Total		
	Col.	Hem.	Hym.	Lep.	Dip.	Other	Yes	No	1
EU to NA	477	368	211	144	138	76	854	560	1414
NA to EU	40	72	54	20	29	15	160	70	230
G (df=1)	435	218	99.4	106	77.3	44.7	522	434	948
$p\ (\geq G)$	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001
EU to AU	137	96	67	31	55	57	226	217	443
AU to EU	34	10	14	7	4	8	41	36	77
G (df=1)	66.5	80.7	37.7	16.4	52.5	41.6	141	144	285
$p\ (\geq G)$	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001
NA to AU	20	26	13	8	10	11	57	31	88
AU to NA	18	22	15	4	6	7	48	24	72
G (df=1)	0.11	0.33	0.14	1.36	1.01	0.90	0.77	0.89	1.60
$p (\geq G)$	~ 1	~ 1	~ 1	~ 1	~ 1	~ 1	0.69	0.69	0.21



Figure 1. Flows of non-native insects between North America (NA), Europe (EU), and Australasia (AU). Numbers indicate the total count of species established from donor to recipient, with flow widths being proportional to these counts. Overlapping flows on the donor side indicate the fraction of species that established in both recipient regions.

many cases (Europe to North America and Australasia, and North America to Australasia), the rates of establishment of non-native insects per billion pounds sterling (hereafter referred to simply as establishment rates) were greatest near the very start of the dataset (circa 1827). These rates decreased over time, particularly between 1940 and 1960, with our models explaining these declines as depletion of the invader pools. Approximately 75% of non-native insect species that established from Europe into North America and Australasia had done so by 1950 (Fig. 2a, c). In contrast, the establishment rates of North American insects into Europe have decreased only very slightly over time (Fig. 2b), and there is no evidence (as per AIC-based model selection) of any decline in the establishment rates of Australasian insects into Europe (Fig. 2d, Table 2).

Discoveries and modelled establishments of non-native insects between North America and Australasia were within the same order of magnitude in both directions



Figure 2. Cumulative discoveries (observed and modelled) and establishments (modelled) of non-native insects exchanged between Europe (EU), North America (NA), and Australasia (AU) versus cumulative import value (inflation-corrected to 2020 British pounds sterling, billions), 1827–2014. Alternating background shading indicates decadal increments, with shading omitted prior to the 1940s for clarity.

(Fig. 2e, f). There was evidence of saturation in the flows of non-native insects between North America and Australasia, though less so from Australasia to North America.

The modelled numbers of non-native insect establishments per billion pounds sterling (annual establishment rate, r) were significantly different for the reciprocal flows between Europe and North America and between Europe to Australasia (Table 2). Between Europe and North America, the predicted sizes of the invader pools (based on the number of discoveries at maximum establishments, d_{sat}) favors Europe, but with overlapping 95% confidence intervals (Table 2). **Table 2.** Parameters and 95% confidence intervals of Poisson-process models of establishments and lagged discoveries of non-native insect species exchanged between Europe (EU), North America (NA), and Australasia (AU). All models included a parameter for imports (r, the number of annual establishments per billion pounds sterling) and lag (π , the annual probability of discovery of established species). Models including an additional term for saturation (a decrease in establishment probability as the cumulative number of discoveries approaches d_{sat} , were selected in most cases, with model selection based on Akaike information criterion (AIC) values.

Flow	Best model	Annual	Discoveries	Annual discovery	Lag years
	($\Delta AIC \ of \ next-best \ model$)	establishment rate,	at maximum	probability,	(95% CI)
		r (95% CI)	establishments,	π (95% CI)	
			d _{sat} (95% CI)		
EU to NA	Imports + saturation + lag (2118)	1.58 (1.40-1.77)	1121	0.0277	36.1 (29.0-48.0)
			(1089–1153)	(0.0345-0.0208)	
NA to EU	Imports + saturation + lag (6.08)	0.0194	701 (290–1114)	0.499 (1-0)	2.0 (1.00-∞)
		(0.0144-0.0245)			
EU to AU	Imports + saturation + lag (251)	1.212 (0.825-1.60)	366 (312-419)	0.0245	40.9 (25.9–96.7)
				(0.0386-0.0103)	
AU to EU	Imports + lag (2.0 [*])	0.0647	n/a	0.0259 (0.0690-0)	38.5 (14.5–∞)
		(0.0173-0.112)			
NA to AU	Imports + saturation + lag (99.6)	0.771 (0.448-1.09)	76 (68–83)	0.0721	13.9 (7.11–283)
				(0.141-0.00354)	
AU to NA	Imports + saturation + lag (8.37)	0.621 (0-2.23)	53 (1.60–104)	0.0153 (0.0598–0)	65.5 (16.7–∞)

* Although this low Δ AIC could be considered "substantial evidence" for both the full and reduced model (Burnham and Anderson 2004), the $d_{_{AP}}$ parameter estimate in the full model greatly exceeded the number of insect species in the donor region, thus the full model effectively lacked saturation and was not ecologically appropriate.

Discussion

Considerably more insect species have invaded North America and Australasia from Europe than in the opposite directions. This concurs with the previously observed overrepresentation of tree-feeding insects from Europe in North America (Niemelä and Mattson 1996), and with non-native insects from the western Palearctic (i.e., Europe) being overrepresented in New Zealand (Edney-Browne et al. 2018). Our results demonstrate that these asymmetries are consistent across all insect orders considered in the analysis, including both herbivorous and non-herbivorous insects.

International trade is considered the single most important pathway for unintentional introductions of insects (Brockerhoff and Liebhold 2017), and greater trade activity generally results in greater propagule pressure of non-native species. Existing literature identifies a positive correlation between import value and the establishment of non-native species (Levine and D'Antonio 2003; Seebens et al. 2017; Lantschner et al. 2020; MacLachlan et al. 2021). Similarly, our models provided excellent fits of inflation-corrected import values to temporal changes in non-native insect establishment rates (after accounting for gradual depletion of source pools). However, the modelled establishment rates (r), which represent the maximum rates of establishments per billion pounds sterling of imports prior to any depletion of source pools, differed significantly between the Europe to North America flow and its converse, and between the Europe to Australasia flow and its converse (Table 2). These significant differences indicate that even after accounting for differences in trade values, large asymmetries between flows remain unexplained by the models. Contrary to our expectations, and despite the important role of trade in facilitating the establishment of non-native species, we must look to other explanations for these asymmetries.

Temporal variation in establishment rates may hold some clues as to the possible causes of the invasional asymmetries. While global establishments of non-native species have not slowed (Seebens et al. 2017; MacLachlan et al. 2021), our results show that establishment rates may be slowing down at regional scales. Establishments of European insects in North America and Australasia per billion pounds sterling of imports have drastically decreased since 1950 (Fig. 2a, c). Several authors have noted or predicted similar declines in the rate of accumulation of exotic species into the United States (Levine and D'Antonio 2003; Liebhold et al. 2017; MacLachlan et al. 2021; Seebens et al. 2021), with two possible explanations offered: depletion of source invader pools, or improved biosecurity measures.

Unequal flows of non-native insects may arise from differences in the numbers of potential invaders present in the donor regions (Vermeij 1991). Our models attempted to predict the size of these invader pools, provided that a decreasing trend in establishment rates could be suitably explained (based on Δ AIC) by the depletion of these pools. Our results suggest that some of the asymmetry in non-native insects exchanged between Europe and North America may be due to a ~60% larger pool of European insect invaders. However, this was not a significant difference, given the large confidence intervals around these estimates. Described insect species richness in Europe is approximately equal to that of North America (de Jong et al. 2014; Arnett 2000). Again, this suggests that the asymmetry between Europe and North America cannot be explained by differences in invader source pool sizes (assuming that the ratio of described to undescribed species is not strikingly greater in North America and Australasia than it is in Europe).

Scientific effort almost certainly varies regionally, and this may impact the interpretation of our results. Over the last few hundred years, Europe has had a consistently greater population density than either North America or Australasia (Goldewijk 2005). If this corresponds to greater scientific effort in Europe (more biologists/naturalists per unit area), the proportion of established species which have been discovered and the proportion of native species which have been described are likely greater in Europe than in North America and Australasia. This has several implications for our analyses. First, it suggests that we may have relatively underestimated the numbers of European insects in North America and Europe and thus the asymmetries may be even more dramatic than our analyses suggest. Second, the published counts of described insect species may be biased in favor of a relatively greater number in Europe, which may predispose us towards suggesting that the asymmetries are due to a greater richness of European invaders. We attempted to account for differential scientific effort in our models, in the form of an annual probability of detection that could vary independently for each of the six flows. Unfortunately, this parameter seldom had a clear optimum, and the resulting confidence intervals are large. These wide confidence intervals may be due in part to using a fixed annual probability of detection (unchanging over

time). Future research may benefit from allowing the discovery probability to change over time, perhaps by incorporating regional time series of proxies for scientific effort.

Despite the lack of statistical significance, the larger estimate for the pool of European insect invaders in North America versus the opposite could be considered a point in favor of the European crucible hypothesis proposed by Niemelä and Mattson (1996). This hypothesis suggests that a history of extensive glaciations may have reduced the niche diversity and 'invasibility' of Europe by leading to extinctions of plant genera, while simultaneously selecting for competition-hardened species that thrive in disturbed habitats, making European species better invaders. However, Europe has been heavily colonized by insects from regions other than North America, particularly the Asian Palearctic (Roques et al. 2020), suggesting that European insects established into North America 81 times more quickly (per billion pounds sterling of imports) than North American insects established into Europe. If this remarkable difference could be explained largely by European insects being better invaders, we would expect model estimates of the invader pool sizes between Europe and North America (using d_{sat} as a proxy) to be considerably more different than they were found to be, and significantly so.

Although we have modelled declining establishment rates as the gradual depletion of source invader pools, it is also likely that biosecurity measures have contributed. International biosecurity regulations, specifically phytosanitary measures, began in earnest in the 20th century (Roques 2010; Allen et al. 2017). With plant-feeding insects making up 58% of all non-native insect species in our dataset, strengthened phytosanitary measures applied to pathways including live plants, wood, and crops have almost certainly led to contemporary reductions in establishment rates. Europe has also had less strict phytosanitary measures than Australia and New Zealand for many decades (Eschen et al. 2015), which may partly explain the relative lack of declines in the rates of discoveries of North American and Australiasia insects (per billion pounds sterling) in Europe. However, strengthening biosecurity efforts are most likely to have influenced the latter half of our time series (1900 and onwards), whereas considerable asymmetry in the numbers of species exchanged between Europe, North America, and Australasia had already accumulated by 1900. Therefore, differential biosecurity is an unlikely driver of these asymmetries.

Though historical invasion discoveries began much earlier, available import data only began in 1827. Given that the greatest establishment rates were seen at the very start of the dataset, it is possible – perhaps even likely – that the main causal agents explaining the dominance of Europe as a source of non-native insects in North America and Australasia were transient phenomena that began prior to 1827. This is complicated further by invasion biology being a relatively new discipline: early records of novel species may be both lacking and underrepresented in scientific literature. After a non-native species establishes, there is typically a time lag until it is discovered (Essl et al. 2011). Although our models attempt to account for lags between establishment and discovery, we used an annual probability of detection that does not change over time, and the lag estimates often have wide confidence intervals. Therefore, we are not confident that our data could be used to extrapolate far into the past. Well before our dataset begins, North America and Australasia were experiencing a period of dramatic change as European colonies were founded. This colonization promoted both deliberate and accidental introductions of European plants (Lenzner et al. 2018). Introductions of exotic plants by colonial powers accelerated in the 19th and early 20th centuries, with a lasting impact on the global composition of floral communities (Lenzner et al. 2022). This is noteworthy, because plant imports and introductions may be a strong predictor of insect invasions (Liebhold et al. 2012; Liebhold et al. 2018; Bonnamour et al. 2023). North America and Australasia each have nearly twice as many extra-continental non-native plant species as Europe, and Europe is second to Asia as a dominant source of non-native plants worldwide (van Kleunen et al. 2015). This history, and the close relationships between insects and plants, suggest a potential explanation for both the existence of the asymmetries we observe and their temporal trends. We suggest that future research focuses on European colonization and coincident plant introductions as possible explanations for why Europe has donated so many more non-native insects into North America and Australasia than it has received from these regions.

We cannot rule out other factors not addressed here, such as differences in establishment probability driven by climate suitability or biotic resistance, the effect of establishments originating from non-native populations ('bridgeheads'), or differences in propagule pressure driven by the specific types of trade goods exchanged between regions. This latter factor is likely the most important to consider for future research, as overall import values may not capture changes over time in the relative contribution of specific commodities (such as plants and plant products) to overall trade. From the discussion above, we know to expect close associations between insects and plant products. Plant products may also have low values per unit of volume, thus being poorly represented in overall import values. Analyses which considered different commodities separately were conducted by Morimoto et al. (2019) but these were limited to Japan, 1951–2016. Data for continent-scale regions (i.e., North America and Europe) going back into the early 1800s do not exist, so far as we are aware, in any cohesive form. To compile such data from historical records would be a major interdisciplinary effort and was beyond the scope of our present research. Regardless, our results are an important step forward in confirming the existence of strong asymmetries in insect establishments between our focal regions and suggesting possible explanations for their cause.

Acknowledgements

This project was funded by the National Socio-Environmental Synthesis Center (National Science Foundation DBI-1639145), Natural Sciences and Engineering Research Council of Canada (NSERC) Discovery Grants (SBH, DSP), USDA Forest Service International Programs 21-IG-11132762-241 (RT), Grant EVA4.0, No. CZ.02.1.0 1/0.0/0.0/16_019/0000803 financed by Czech Operational Programme Science, Research, and Education (AML), European Union project HOMED (HOlistic Management of Emerging Forest Pests and Diseases- grant No. 771271) (AR, EGB), Fondation Sandoz-Monique de Meuron (CB), Swiss National Science Foundation (SNSF) (CB), and the University of Padua under the 2019 STARS Grants program (project: MOPI–Microorganisms as hidden players in insect invasions) (DR).

References

- Allen E, Noseworthy M, Ormsby M (2017) Phytosanitary measures to reduce the movement of forest pests with the international trade of wood products. Biological Invasions 19(11): 3365–3376. https://doi.org/10.1007/s10530-017-1515-0
- ArnettJrRH(2000)AmericanInsects(2nd edn.).CRCPress.https://doi.org/10.1201/9781482273892
- Blake RE, Turner R (2021) reblake/insectcleanr: Initial release. https://doi.org/10.5281/ ZENODO.4555787
- Bonnamour A, Blake RE, Liebhold AM, Nahrung HF, Roques A, Turner RM, Yamanaka T, Bertelsmeier C (2023) Historical plant introductions predict current insect invasions. Proceedings of the National Academy of Sciences of the United States of America 120(24): e2221826120. https://doi.org/10.1073/pnas.2221826120
- Bradshaw CJA, Leroy B, Bellard C, Roiz D, Albert C, Fournier A, Barbet-Massin M, Salles J-M, Simard F, Courchamp F (2016) Massive yet grossly underestimated global costs of invasive insects. Nature Communications 7(1): e12986. https://doi.org/10.1038/ncomms12986
- Brockerhoff EG, Liebhold AM (2017) Ecology of forest insect invasions. Biological Invasions 19(11): 3141–3159. https://doi.org/10.1007/s10530-017-1514-1
- Burnham KP, Anderson DR (2004) Multimodel Inference: Understanding AIC and BIC in Model Selection. Sociological Methods & Research 33(2): 261–304. https://doi. org/10.1177/0049124104268644
- Byrd RH, Lu P, Nocedal J, Zhu C (1995) A limited memory algorithm for bound constrained optimization. SIAM Journal on Scientific Computing 16(5): 1190–1208. https://doi. org/10.1137/0916069
- Costello CJ, Solow AR (2003) On the pattern of discovery of introduced species. Proceedings of the National Academy of Sciences of the United States of America 100(6): 3321–3323. https://doi.org/10.1073/pnas.0636536100
- Costello C, Springborn M, McAusland C, Solow A (2007) Unintended biological invasions: Does risk vary by trading partner? Journal of Environmental Economics and Management 54(3): 262–276. https://doi.org/10.1016/j.jeem.2007.06.001
- de Jong Y, Verbeek M, Michelsen V, Bjørn PDP, Los W, Steeman F, Bailly N, Basire C, Chylarecki P, Stloukal E, Hagedorn G, Wetzel F, Glöckler F, Kroupa A, Korb G, Hoffmann A, Häuser C, Kohlbecker A, Müller A, Güntsch A, Stoev P, Penev L (2014) Fauna Europaea – all European animal species on the web. Biodiversity Data Journal 2: e4034. https://doi. org/10.3897/BDJ.2.e4034
- Edney-Browne E, Brockerhoff EG, Ward D (2018) Establishment patterns of non-native insects in New Zealand. Biological Invasions 20(7): 1657–1669. https://doi.org/10.1007/ s10530-017-1652-5
- Eschen R, Britton K, Brockerhoff E, Burgess T, Dalley V, Epanchin-Niell RS, Gupta K, Hardy G, Huang Y, Kenis M, Kimani E, Li H-M, Olsen S, Ormrod R, Otieno W, Sadof C,

Tadeu E, Theyse M (2015) International variation in phytosanitary legislation and regulations governing importation of plants for planting. Environmental Science & Policy 51: 228–237. https://doi.org/10.1016/j.envsci.2015.04.021

- Essl F, Dullinger S, Rabitsch W, Hulme PE, Hülber K, Jarošík V, Kleinbauer I, Krausmann F, Kühn I, Nentwig W, Vilà M, Genovesi P, Gherardi F, Desprez-Loustau M-L, Roques A, Pyšek P (2011) Socioeconomic legacy yields an invasion debt. Proceedings of the National Academy of Sciences of the United States of America 108(1): 203–207. https://doi.org/10.1073/pnas.1011728108
- Fouquin M, Hugot J (2016) Two centuries of bilateral trade and gravity data: 1827–2014. CEPII Working Paper 2016–14. http://www.cepii.fr/CEPII/en/publications/wp/abstract. asp?NoDoc=9134
- GBIF.org (2022) GBIF Home Page. https://www.gbif.org/
- Goldewijk KK (2005) Three centuries of global population growth: A Spatial referenced population (density) database for 1700–2000. Population and Environment 26(4): 343–367. https://doi.org/10.1007/s11111-005-3346-7
- Guyot V, Castagneyrol B, Vialatte A, Deconchat M, Selvi F, Bussotti F, Jactel H (2015) Tree diversity limits the impact of an invasive forest pest. PLoS ONE 10(9): e0136469. https://doi.org/10.1371/journal.pone.0136469
- Kenis M, Auger-Rozenberg M-A, Roques A, Timms L, Péré C, Cock MJW, Settele J, Augustin S, Lopez-Vaamonde C (2009) Ecological effects of invasive alien insects. Biological Invasions 11(1): 21–45. https://doi.org/10.1007/s10530-008-9318-y
- Lantschner MV, Corley JC, Liebhold AM (2020) Drivers of global Scolytinae invasion patterns. Ecological Applications 30(5): e02103. https://doi.org/10.1002/eap.2103
- Lenzner B, Essl F, Seebens H (2018) The Changing Role of Europe in Past and Future Alien Species Displacement. In: Rozzi R, May RH, Chapin Iii FS, Massardo F, Gavin MC, Klaver IJ, Pauchard A, Nuñez MA, Simberloff D (Eds) From Biocultural Homogenization to Biocultural Conservation. Ecology and Ethics. Springer International Publishing, Cham, 125–135. https://doi.org/10.1007/978-3-319-99513-7_8
- Lenzner B, Latombe G, Schertler A, Seebens H, Yang Q, Winter M, Weigelt P, Van Kleunen M, Pyšek P, Pergl J, Kreft H, Dawson W, Dullinger S, Essl F (2022) Naturalized alien floras still carry the legacy of European colonialism. Nature Ecology & Evolution 6(11): 1723–1732. https://doi.org/10.1038/s41559-022-01865-1
- Levine JM, D'Antonio CM (2003) Forecasting Biological Invasions with Increasing International Trade. Conservation Biology 17(1): 322–326. https://doi.org/10.1046/j.1523-1739.2003.02038.x
- Liebhold AM, Brockerhoff EG, Garrett LJ, Parke JL, Britton KO (2012) Live plant imports: The major pathway for forest insect and pathogen invasions of the US. Frontiers in Ecology and the Environment 10(3): 135–143. https://doi.org/10.1890/110198
- Liebhold AM, Brockerhoff EG, Kimberley M (2017) Depletion of heterogeneous source species pools predicts future invasion rates. [Beggs J (Ed.)] Journal of Applied Ecology 54: 1968–1977. https://doi.org/10.1111/1365-2664.12895
- Liebhold AM, Yamanaka T, Roques A, Augustin S, Chown SL, Brockerhoff EG, Pyšek P (2018) Plant diversity drives global patterns of insect invasions. Scientific Reports 8(1): 12095. https://doi.org/10.1038/s41598-018-30605-4

- Liebhold AM, Turner RM, Blake RE, Bertelsmeier C, Brockerhoff EG, Nahrung HF, Pureswaran DS, Roques A, Seebens H, Yamanaka T (2021) Invasion disharmony in the global biogeography of native and non-native beetle species. Diversity and Distributions 27: 2050–2062. https://doi.org/10.1111/ddi.13381
- MacLachlan MJ, Liebhold AM, Yamanaka T, Springborn MR (2021) Hidden patterns of insect establishment risk revealed from two centuries of alien species discoveries. Science Advances 7(44): eabj1012. https://doi.org/10.1126/sciadv.abj1012
- Mally R, Turner RM, Blake RE, Fenn-Moltu G, Bertelsmeier C, Brockerhoff EG, Hoare RJB, Nahrung HF, Roques A, Pureswaran DS, Yamanaka T, Liebhold AM (2022) Moths and butterflies on alien shores: Global biogeography of non-native Lepidoptera. Journal of Biogeography 49(8): 1455–1468. https://doi.org/10.1111/jbi.14393
- Niemalä P, Mattson WJ (1996) Invasion of north american forests by european phytophagous insects. Bioscience 46(10): 741–753. https://doi.org/10.2307/1312850
- Morimoto N, Kiritani K, Yamamura K, Yamanaka T (2019) Finding indications of lag time, saturation and trading inflow in the emergence record of exotic agricultural insect pests in Japan. Applied Entomology and Zoology 54(4): 437–450. https://doi.org/10.1007/ s13355-019-00640-2
- Nahrung HF, Carnegie AJ (2020) Non-native Forest Insects and Pathogens in Australia: Establishment, Spread, and Impact. Frontiers in Forests and Global Change 3: 1–37. https:// doi.org/10.3389/ffgc.2020.00037
- Office for National Statistics (2021) Office for National Statistics, consumer price inflation time series. https://www.ons.gov.uk/economy/inflationandpriceindices/datasets/consumerpriceindices
- R Core Team (2021) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna. https://www.R-project.org/
- Rassati D, Faccoli M, Haack RA, Rabaglia RJ, Petrucco Toffolo E, Battisti A, Marini L (2016) Bark and ambrosia beetles show different invasion patterns in the USA. PLoS ONE 11(7): e0158519. https://doi.org/10.1371/journal.pone.0158519
- Rigot T, Van Halder I, Jactel H (2014) Landscape diversity slows the spread of an invasive forest pest species. Ecography 37(7): 648–658. https://doi.org/10.1111/j.1600-0587.2013.00447.x
- Roques A (2010) Alien forest insects in a warmer world and a globalised economy: Impacts of changes in trade, tourism and climate on forest biosecurity. New Zealand Journal of Forestry Science 40: S77–S94.
- Roques A, Shi J, Auger-Rozenberg M-A, Ren L, Augustin S, Luo Y (2020) Are invasive patterns of non-native insects related to woody plants differing between Europe and China? Frontiers in Forests and Global Change 2: 1–91. https://doi.org/10.3389/ffgc.2019.00091
- Seebens H, Blackburn TM, Dyer EE, Genovesi P, Hulme PE, Jeschke JM, Pagad S, Pyšek P, Winter M, Arianoutsou M, Bacher S, Blasius B, Brundu G, Capinha C, Celesti-Grapow L, Dawson W, Dullinger S, Fuentes N, Jäger H, Kartesz J, Kenis M, Kreft H, Kühn I, Lenzner B, Liebhold A, Mosena A, Moser D, Nishino M, Pearman D, Pergl J, Rabitsch W, Rojas-Sandoval J, Roques A, Rorke S, Rossinelli S, Roy HE, Scalera R, Schindler S, Štajerová K, Tokarska-Guzik B, Van Kleunen M, Walker K, Weigelt P, Yamanaka T, Essl F (2017) No

saturation in the accumulation of alien species worldwide. Nature Communications 8(1): e14435. https://doi.org/10.1038/ncomms14435

- Seebens H, Bacher S, Blackburn TM, Capinha C, Dawson W, Dullinger S, Genovesi P, Hulme PE, Kleunen M, Kühn I, Jeschke JM, Lenzner B, Liebhold AM, Pattison Z, Pergl J, Pyšek P, Winter M, Essl F (2021) Projecting the continental accumulation of alien species through to 2050. Global Change Biology 27(5): 970–982. https://doi.org/10.1111/gcb.15333
- Turner R, Blake R, Liebhold AM (2021) International Non-native Insect Establishment Data. https://doi.org/10.5281/ZENODO.5245302
- van Kleunen M, Dawson W, Essl F, Pergl J, Winter M, Weber E, Kreft H, Weigelt P, Kartesz J, Nishino M, Antonova LA, Barcelona JF, Cabezas FJ, Cárdenas D, Cárdenas-Toro J, Castaño N, Chacón E, Chatelain C, Ebel AL, Figueiredo E, Fuentes N, Groom QJ, Henderson L, Inderjit, Kupriyanov A, Masciadri S, Meerman J, Morozova O, Moser D, Nickrent DL, Patzelt A, Pelser PB, Baptiste MP, Poopath M, Schulze M, Seebens H, Shu W, Thomas J, Velayos M, Wieringa JJ, Pyšek P (2015) Global exchange and accumulation of non-native plants. Nature 525(7567): 100–103. https://doi.org/10.1038/nature14910
- Vermeij GJ (1991) Anatomy of an invasion: The trans-Arctic interchange. Paleobiology 17(3): 281–307. https://doi.org/10.1017/S0094837300010617
- Vermeij GJ (1996) An agenda for invasion biology. Biological Conservation 78(1–2): 3–9. https://doi.org/10.1016/0006-3207(96)00013-4
- Visser V, Wilson JRU, Fish L, Brown C, Cook GD, Richardson DM (2016) Much more give than take: South Africa as a major donor but infrequent recipient of invasive non-native grasses: South Africa, major donor, but minor recipient of invasive grasses. Global Ecology and Biogeography 25(6): 679–692. https://doi.org/10.1111/geb.12445

Supplementary material I

Data for Asymmetrical insect invasions between three world regions

Authors: Rylee Isitt, Andrew M. Liebhold, Rebecca M. Turner, Andrea Battisti, Cleo Bertelsmeier, Rachael Blake, Eckehard G. Brockerhoff, Stephen B. Heard, Paal Krokene, Bjørn Økland, Helen F. Nahrung, Davide Rassati, Alain Roques, Takehiko Yamanaka, Deepa S. Pureswaran

Data type: docx

- Explanation note: Annual (1827–2014) and undated discoveries of non-native insects and annual inflation-corrected import values (in 2020 British pounds sterling, billions) exchanged between Europe (EU), North America (NA, north of Mexico) and Australasia (AU, Australia and New Zealand only).
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.110942.suppl1

RESEARCH ARTICLE



Worldwide distribution and phylogeography of the agave weevil Scyphophorus acupunctatus (Coleoptera, Dryophthoridae): the rise of an overlooked invasion

Andrea Viviano^{1,2}, Arturo Cocco³, Paolo Colangelo^{4,5}, Giuseppe Marco Delitala⁶, Roberto Antonio Pantaleoni^{1,3}, Laura Loru¹

National Research Council (CNR), Institute of Research on Terrestrial Ecosystems (IRET), trav. La Crucca 3, Baldinca, 07100 Sassari, Italy 2 National Research Council (CNR), Institute of Research on Terrestrial Ecosystems (IRET), via Madonna del Piano 10, 50019 Sesto Fiorentino (Florence), Italy 3 Department of Agriculture, University of Sassari, Viale Italia 39, 07100, Sassari, Italy 4 National Research Council (CNR), Institute of Research on Terrestrial Ecosystems (IRET), Strada Provinciale 35d 9, 00010 Montelibretti (Rome), Italy 5 National Biodiversity Future Center, 90133, Palermo, Italy 6 via Claudio Fermi 24, 07100, Sassari, Italy

Corresponding author: Arturo Cocco (acocco@uniss.it)

Academic editor: Katelyn Faulkner | Received 14 February 2023 | Accepted 6 November 2023 | Published 9 January 2024

Citation: Viviano A, Cocco A, Colangelo P, Delitala GM, Pantaleoni RA, Loru L (2024) Worldwide distribution and phylogeography of the agave weevil *Scyphophorus acupunctatus* (Coleoptera, Dryophthoridae): the rise of an overlooked invasion. NeoBiota 90: 53–78. https://doi.org/10.3897/neobiota.90.101797

Abstract

Global plant trade represents one of the main pathways of introduction for invertebrates, including insects, throughout the world. Non-native insects include some of the most important pests affecting cultivated and ornamental plants worldwide. Defining the origins and updating the distribution of non-native invasive species is pivotal to develop effective strategies to limit their spread. The agave weevil, *Scyphophorus acupunctatus* (Coleoptera, Dryophthoridae), is a curculionid beetle native to Central and North America, although it also occurs in Eurasia, Africa, Oceania and South America as a non-native species. Despite being widespread, the extent of occurrence and origins of European populations of the agave weevil have been overlooked. In the present study, the current and potential worldwide distribution of *S. acupunctatus* was assessed and an analysis of its genetic diversity in the native and non-native ranges was performed. By analysing occurrences from local phytosanitary bulletins and citizen-science platforms, the agave weevil was confirmed to be widely distributed and to occur on all continents, except Antarctica. Additionally, there is potential for expansion throughout the world, as estimated by species distribution models. Nucleotide and haplotype diversity of the COXI mitochondrial gene (about 650 bp) was lower in the non-native (n = 39 samples) than native populations (n = 26 samples). The majority of introduced

Copyright Andrea Viviano et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

individuals belonged to the same haplotype, suggesting that most introductions in Europe might have occurred from a small geographical area in Central America. Constant transboundary monitoring and national laws must be considered to reduce the spread of the agave weevil, given that a bridgehead effect may occur from non-native populations to new suitable areas.

Keywords

Agave, mitochondrial COXI gene, non-native invasive insects, population genetics, species distribution model

Introduction

Non-native invasive species are taxa that have been introduced and/or spread into regions outside their native ranges and have subsequently established and spread, affecting local ecosystem dynamics (CBD 2010). Since the Holocene and the earliest explorers, human migration has been essential to the movement of species from their native ranges to areas where they were not present (Foster et al. 2002; Banks et al. 2015). Globalisation has intensified the human-assisted spread of living species in non-native areas, following international trade and human journeys (Meyerson and Mooney 2007). In addition, the creation of ecological corridors has facilitated the range expansion of many taxa in non-native countries (Mattson et al. 2007; Horsák et al. 2019).

Crop pests are widely distributed worldwide due to accidental introductions through the intensive trade of goods, including plants of ornamental and agronomic interest (Deutsch et al. 2018). Amongst crop pests, many invertebrate species have been thoroughly studied, particularly in biocontrol and pest management research (Geier 1966; Parsons et al. 2020). Well-studied crop pest species include Halyomorpha halys (Stål) and Tuta absoluta (Meyrick) (Biondi et al. 2018; Cianferoni et al. 2018). However, most non-native insect pests have been poorly investigated and their impact and distribution are currently still under assessment (e.g. Corythauma ayyari (Drake) and Stator limbatus (Horn): Mazza et al. (2020); Cocco et al. (2021)). For instance, palms in Mediterranean countries are threatened by both the well-known red palm weevil, Rhynchophorus ferrugineus (Olivier) (Soroker and Colazza 2017) and the still mostly unknown and overlooked palm borer moth, Paysandisia archon (Burmeister) (Mori et al. 2023). Curculionid coleopterans (e.g. weevils) are an important threat to many cultivated species including corn, figs, palms and other ornamental plants (Guzmàn et al. 2012; Inghilesi et al. 2015; Farina et al. 2020). Amongst those, the agave weevil, Scyphophorus acupunctatus Gyllenhal (Coleoptera, Dryophthoridae), is one of the least-studied species. This weevil is native to southern North America, Mexico and other countries in Central America (Vaurie 1971), although it has been introduced to several parts of the world including American islands and South America (US Virgin Islands and Hawaii, Cayman Islands, Puerto Rico, Cuba, Haiti, Jamaica, Dominican Republic, Curaçao, Colombia, Venezuela and Brazil), Africa (Kenya, Tanzania and South Africa), Asia (Indonesia and Saudi Arabia), Oceania (South Australia and Fiji Islands) and Europe (Portugal including Madeira, Spain including Canary and Balearic

Islands, France, Italy, Croatia, Greece and Cyprus: Setliff and Anderson (2011); CABI/ EPPO (2014); Vassiliou and Kitsis (2015); Andrade (2022); Pernek and Cvetković (2022)). Populations of *S. acupunctatus* in Central America (Honduras, Belize, Guatemala, Costa Rica, El Salvador and Nicaragua) have an uncertain origin, as it is unclear whether they are native or not. These populations may represent an undocumented natural range expansion from northern countries, i.e. Mexico, in recent times or they might have been introduced through plant and horticultural trade (Vassiliou and Kitsis 2015; EPPO 2022a). Occurrences of the agave weevil in Israel, New Zealand, Queensland (Australia), Argentina and the United Kingdom that have been reported by some authors (CABI 2023), have never been confirmed in the scientific literature, nor in citizen-science platforms or social networks. In general, the distribution of this weevil is mainly known at the country level, with little known about its actual distribution within each country (Martín-Taboada et al. 2019).

The agave weevil is a major pest of agave. Agaves (Asparagaceae, Agavoideae/Agavaceae) include several genera and species that have been introduced worldwide for ornamental purposes (Thiede et al. 2019). Most agave species are susceptible to this weevil, particularly those belonging to the genus *Agave* (Vaurie 1971; Bolaños et al. 2014; Palemòn-Alberto et al. 2022). Plants are directly damaged by the agave weevil whose larvae feed on agave heads by boring galleries (Figueroa-Castro et al. 2016). The consumption of plant parts by the agave weevil larva may cause plant mortality (Aquino-Bolaños et al. 2013). Adults cause little damage in comparison to larvae.

The taxonomy of the Scyphophorus genus is still unresolved (Chamorro et al. 2016). Although two species are traditionally recognised, S. acupunctatus and S. yuccae Horn, no reliable information on the phylogeography of this genus is available. Genetic analyses of Scyphophorus spp. have been carried out on a limited sample size or in limited geographical areas of Central America (Azuara-Domínguez et al. 2013; Chamorro et al. 2016). Furthermore, no molecular data are available to disentangle the two Scyphophorus species, given that the only deposited sequence of S. yuccae is actually belonging to S. acupunctatus, questioning the actual validity of the former species (Chamorro et al. 2016). Assessing the geographic origin of non-native populations of S. acupunctatus may help to provide information for plant trade controls and assist with preventing new invasions. Although S. acupunctatus is also recorded in Africa, SE Asia and Australia, most non-native populations of this weevil occur in Europe, where S. acupunctatus has been introduced through the plant trade (e.g. Beaucarnea recurvata Lem., Agave americana L. and Yucca spp.), most likely from different countries of Central and North America (e.g. from Nicaragua to Italy: EPPO (2022b)). The agave weevil was reported for the first time in the Netherlands in 1980 (van Rossem et al. 1981) and, subsequently, in Italy, France, Spain and Greece (Colombo 2000; Flinch and Alonso-Zarazaga 2007; EPPO 2008; Kontodimas and Kallinikou 2010). Therefore, we focused mostly on European and Mediterranean countries, as these countries include most of the non-native range of this weevil species.

The aims of our work were to: (*i*) update the distribution of the agave weevil in nonnative areas with special regard to Mediterranean countries; (*ii*) determine the climatic suitability throughout the world, with special regard to Europe, where most non-native populations occur and predict its potential distribution; and (*iii*) assess the phylogeographic pattern of *S. acupunctatus* and trace the origin of European populations.

Materials and methods

Updating the distribution of the agave weevil

The distribution of the agave weevil in its non-native range was updated by searching for published and unpublished records in the grey and scientific literature and online databases, including records collected through citizen-science and validated by experts (i.e. iNaturalist: www.inaturalist.org; GBIF: www.gbif.org DOI: https://doi.org/10.15468/ dl.pd22mh; Forum Natura Mediterraneo: www.naturamediterraneo.com; Forum Entomologi Italiani: www.entomologiitaliani.net. All accessed on 15.05.2023). The search for occurrence records was conducted from October 2022 to May 2023. Further searches were performed on free posts with photos on Social Networks (e.g. Facebook) and on video-sharing websites (e.g. YouTube). The literature search was carried out by assessing studies in online databases (i.e. ISI Web of Science, Scopus, Zoological Records and Google Scholar). Search terms included all possible combinations of the words: 'agave weevil', 'Scyphophorus acupunctatus', 'distribution' and 'non-native species'. The same words were searched in English, French, Spanish, Portuguese and Italian. Maps representing the agave weevil distribution using geographical coordinates were downloaded from the ESRI (https://server.arcgisonline.com) and Eurostat (Countries - GISCO -Eurostat, europa.eu) websites. The distribution of the weevil was mapped using QGIS software version 3.28 Firenze (QGIS Development Team 2019).

The suitability of current and future climates for the agave weevil: preliminary analyses

The potential worldwide distribution of *S. acupunctatus* was modelled to identify areas throughout the Globe that are climatically suitable for this weevil. To the best of our knowledge, no previous studies have focused on the climatic preferences of this weevil, despite its high impact on agro-economy and urban parks.

Occurrence records from both the native and non-native ranges were collected, representing the whole realised ecological niche (Srivastava et al. 2021). This approach resulted in a total of 1525 high-accuracy occurrence records (uncertainty < 1 km). The raw dataset underwent a meticulous analysis to identify and eliminate duplicate entries. This process was carried out in two steps: an initial manual inspection employing the duplicate search function in Microsoft Excel (Microsoft Office 365), followed by subsequent verification using the "duplicated" function of "spocc" package (Chamberlain et al. 2017) in the R software version 4.1.2 (R Core Team 2019). By implementing these measures, overlapping data points from various sources were successfully identified and removed. A final new dataset of 1135 occurrences without duplicates was obtained.

Moran's correlograms were employed to test for the presence of significant spatial autocorrelation (De Marco et al. 2008), using spatial analysis tools available in ArcGIS Pro (ESRI 2011). The spatial autocorrelation analysis was conducted using the final dataset as the input file. In detail, we assessed the spatial autocorrelation between 1 and 10 km at 1 km intervals (De Marco et al. 2008; Crase et al. 2014).

The Moran's correlogram is a graphical representation of the spatial autocorrelation coefficient (Moran's I) at different distance intervals, which helps to identify patterns of spatial dependence and assess whether neighbouring observations are more similar or dissimilar from each other than expected by chance (Crase et al. 2014). The Moran's I coefficient ranges from -1 to 1, where positive values indicate positive spatial autocorrelation (similar values tend to cluster together), negative values indicate negative spatial autocorrelation (dissimilar values tend to be clustered) and values close to zero indicate no spatial autocorrelation (values are randomly distributed across space: Crase et al. (2014), Suppl. material 1: fig. S1).

In this work, the computed Moran's Index was 0.03, indicating a slight positive spatial autocorrelation in the dataset. The Z-score, which measures the standard deviation from the expected mean under the assumption of spatial randomness, was 0.18. The associated P-value was 0.86, suggesting that the observed spatial pattern was not significantly different from what would be expected by chance. Overall, these findings suggested the absence of significant spatial clustering or dispersion in the analysed spatial context. The final dataset used in the model consisted of 718 occurrences.

A distance threshold of 10 km was set to define spatial relationships between observations. This threshold represents the maximum distance at which observations are spatially related. The analysis was performed without any specific selection set, meaning that all observations within the study area were included in the analysis. No weight matrix file was used, suggesting that all observations were assumed to have equal influence in the analysis.

Dispersal abilities of *Scyphophorus* weevils are limited (< 50 metres), as reported by the scientific literature (Huxman et al. 1997; Figueroa-Castro et al. 2016). In line with the spatial autocorrelation analysis, a 10 km distance was selected to filter the occurrences (Di Cola et al. 2017; Montalva et al. 2017; Atauchi et al. 2018; Guevara et al. 2018).

In the final analysis, occurrences were filtered by selecting the minimum distance of 10 km between different occurrence points using the "spThin" R package (Aiello-Lammens et al. 2015). This distance threshold allows for the consideration of occurrences as independent from one another and aligns with the resolution of climate data (Ancillotto et al. 2023).

Selection of variables

The modelling process was started by obtaining 19 climatic variable layers from the Worldclim (version 2.1) website, with a resolution of 2.5 minutes of a degree (Fick and Hijmans 2017). Subsequently, a Principal Component Analysis (PCA: Suppl. material 1: figs S2, S3) was performed using the "ade4" package in R to identify variables with a high collinearity and explore their correlation structure (Fourcade et al. 2014).

Variables were carefully chosen for modelling *S. acupunctatus* by excluding those showing strong intercorrelation. As a result, six highly-significant variables were selected to model the distribution of *S. acupunctatus* (Suppl. material 1: table S1). These variables included BIO1 (Annual Mean Temperature), BIO4 (Temperature Seasonality), BIO6 (Minimum Temperature of the Coldest Month), BIO7 (Temperature Annual Range), BIO9 (Mean Temperature of the Driest Quarter) and BIO11 (Mean Temperature of the Coldest Quarter).

Additionally, the Variance Inflation Factor (VIF) for all selected variables was computed using the "usdm" package in R (Naimi et al. 2014). The VIF values were examined to ensure that all values were below 3, indicating a very low level of multicollinearity (Prakash 2019). Specifically, variables with a Pearson's correlation coefficient of below 0.70 or above -0.70 were retained (Alin 2010; Kock and Lynn 2012; Regos et al. 2020) (Suppl. material 1: table S2). The six bioclimatic variables that were selected to model the distribution of *S. acupunctatus* under current climatic conditions were also chosen to model the distribution of the species under future climates, spanning from 2041 to 2070. Future climate data were downloaded under the Representative Concentration Pathways (RCP 2.6) scenario. The RCP 2.6 future bioclimatic raster is widely acknowledged in literature as a representative case for mitigation strategies aimed at constraining the rise of global mean temperature to 2 °C (van Vuuren et al. 2011).

Algorithm selection

A first comprehensive evaluation was conducted to estimate the performance of nine algorithms through a combination of R packages such as "ENMeval" and "sdm" (Kass et al. 2021; Montoya-Jiménez et al. 2022).

The evaluation encompassed a range of algorithms, namely the Generalised Linear Model (GLM, with a logit-link function), Boosted Regression Trees (BRT, with 15% holdout validation point and bagging fraction set to 0.5: Mui (2015)), Random Forest (RF, with max. tree depth = 2-4: Valavi et al. (2021)), Maximum Entropy (MaxEnt), Generalised Additive Model (GAM), Multivariate Adaptive Regression Splines (MARS), Geometric Brownian Motion (GBM), BIOCLIM and Functional Data Analysis (FDA: Pecchi et al. 2019; Steen et al. 2021). The goal was to identify the most suitable models for the study and reduce computational efforts. To achieve this, along with presence records, 6000 random background points (1000 background points per continental area where the occurrence of S. acupunctatus is reported, i.e. North America, South America, Europe, Asia, Africa and Oceania) were generated (Barber et al. 2022; Buonincontri et al. 2023). In particular, background points were selected in a buffer of 10,000 metres around occurrences, in line with previous literature (Iturbide et al. 2015; Rotllan-Puig and Traveset 2021). Evaluation metrics, such as the Area Under the Curve (AUC) and True Skill Statistics (TSS), were employed to assess the model performance (Suppl. material 1: table S3: Steen et al. (2021)). Unsuitable models (AUC < 0.90; TSS < 0.75) were discarded.

Modelling

Species Distribution Models (SDM) were performed using the R packages "biomod2" and "sdm" (Thuiller 2014; Naimi and Araújo 2016). Following the previous evaluation, only the most suitable model algorithms were selected for the inclusion in the ensemble model. The selection process aimed at choosing models with the highest performance to promote accurate forecasts and ensure reliable results (Thuiller 2014). An ensemble species distribution model was fitted using four algorithms: MaxEnt, RF, GLM and GAM (Araujo and New 2007). By incorporating both statistical and machine-learning approaches, the ensemble approach enables a comprehensive analysis and assessment of the species' potential distribution, which cannot be reached with a single-model approach when the performance of the individual models is low (Araújo and New 2007; Buisson et al. 2010; Hao et al. 2019). This integration of different modelling techniques enhances the robustness of the analysis and improves the overall understanding of the studied phenomenon. Amongst the obtained models, the RF performed the best, with an AUC = 0.99 and TSS = 0.90 (Suppl. material 1: table S3).

The results of the models were assembled with a weighted average of all predictions from all fitted models (Buisson et al. 2010; Smith et al. 2017). The variables for future projections (2041–2070) were then downloaded. Future projections of these variables were obtained for the emission-conservative scenario known as RPC 2.6. Built models were then projected under future climatic conditions. The bioclimatic rasters for future climates at a 2.5-minute degree resolution were evaluated following the same procedures described earlier (Ancillotto et al. 2016, 2020; Cancellario et al. 2023). This approach provided valuable insights into the possible impacts of climate change on the climatic suitability of the world for the agave weevil. For the RCP 2.6 scenario and for each variable, the median of five Global Circulation Models (GCMs) was used: GFDL-ESM4, UKESM1-0LL, MPIESM1-2-HR, IPSLCM6A-LR and MRI-ESM2-0 (Mori et al. 2023). Models were validated using spatial cross validation with the R package "blockCV" (Valavi et al. 2019). The K-fold cross validation was performed, with K = 5 as determined through the "buffer evaluation", i.e. by using the function "cv_buffer" (Pohjankukka et al. 2017).

Model performance was measured using TSS and AUC. For present and future projections, an occurrence probability raster was obtained for each statistical model by calculating the mean of all the projections with a TSS > 0.75 and an AUC > 0.90 (Mori et al. 2023).

Then, differences between predictions under future and current climates were obtained using consensus models, by subtracting the average predictions under current climates from those under future climate. Raster cells with positive values indicated a predicted improvement in climatic conditions for *S. acupunctatus*, whereas raster cells with negative values indicated a decreased climatic suitability for the future. To estimate the uncertainty in the predictions due to disagreements amongst four different algorithms, subtraction per model was performed and the following values were assigned: value -1 was assigned to all cells with negative values of the average singlemodel predictions; similarly, the value +1 was assigned to all cells with positive values and 0 otherwise (Mori et al. 2023).

The consensus of model predictions was obtained by summing the four three-value maps (-1, 0, 1). A raster map was obtained with values ranging between -4 and +4, with extreme values suggesting that all the four statistical models predicted a decrease (-4) or an increase (+4) in the probability of occurrence, whereas intermediate values indicated a partial (± 2 ; ± 3) or high disagreement (-1 to +1) amongst the predictions of the algorithms (Suppl. material 1: fig. S4).

The potential non-analogue climate was checked using a Multivariate Environmental Similarity Surface (MESS) analysis (Elith et al. 2011; Fischer et al. 2011). The MESS analysis estimates the similarity between environments used to train the model and the new projected areas for every grid cell (Elith et al. 2011). The analysis was used to detect regions with environments that are outside the range of environments in the training area (Fischer et al. 2011). Climatic similarities between regions and periods were determined by MESS values. Negative values represent non-analogue climatic conditions.

Phylogenetic and genetic diversity analysis

A total of 32 individual samples of *S. acupunctatus* were collected in Europe and preserved in 95% ethanol at -20 °C, before genetic analyses. Four other samples from Liguria (Pallanca and Hanbury Botanical Gardens, located in Bordighera and Ventimiglia, respectively, Imperia Province, NW Italy) were previously collected by the CNR-IRET researchers and stored in absolute ethanol at the laboratory of CNR-IRET in Sesto Fiorentino (Florence, Italy) (Table 1).

Genomic DNA from all samples was extracted using QIAGEN Blood and Tissue kit (Qiagen Inc., USA), following the manufacturer's protocol. A fragment of the mitochondrial DNA Cytochrome Oxidase I (COXI) was amplified and compared with sequences deposited in the GenBank. COXI was amplified using the primers LCO1490: 5'-GGTCAACAAATCATAAAGATATTGG-3' and HCO2198: '5-TAAACTTCAGGGTGACCAAAAAATCA-3' (Folmer et al. 1994). These primers were previously used to amplify the same gene in *S. acupunctatus* from Central America for species-identification purposes (Azuara-Domínguez et al. 2013; Chamorro et al. 2016: Table 2) using the amplification protocol reported by Baratti et al. (2005) and Chamorro et al. (2016).

PCR products were run on a 1.5% agarose gel, then purified (ExoSAP-IT, Amersham Biosciences) and finally sent to BMR Genomics (Padua, Italy) for Sanger sequencing. Electropherograms were visualised with the software Chromas 1.45 (http://www. technelysium.com.au. Accessed on 17.12.2022). The sequences were visually corrected and aligned using ClustalX 2.1 (Thompson et al. 1997), together with all the available COXI sequences of *S. acupunctatus* retrieved from GenBank and BOLD System, for a total of 65 sequences (627–903 bp: Table 2).

The phylogenetic reconstruction was conducted by applying Neighbour Joining (NJ), Bayesian Inference (BI) and Maximum Likelihood (ML) methods.

Sample ID	Location of origin	Country	Latitude (°N) / Longitude (°E)
S1	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.04893°N, 8.93734°E
S2	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.04588°N, 8.93496°E
S3	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.04454°N, 8.93399°E
S4	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.04150°N, 8.92494°E
S5	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.03500°N, 8.92161°E
S6	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.03612°N, 8.92197°E
S7	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.03348°N, 8.91776°E
S8	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02657°N, 8.89292°E
S9	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02547°N, 8.89186°E
S10	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02617°N, 8.89052°E
S22	Villamaniscicle coll del Quirc, Girona	Spain	42.38092°N, 3.07618°E
S23	Villamaniscicle coll del Quirc, Girona	Spain	42.38092°N, 3.07618°E
S29	Tamaracciu, Corsica	France	41.55294°N, 9.31810°E
S30	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02632°N, 8.88836°E
S31	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02580°N, 8.88484°E
S32	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02669°N, 8.88217°E
S33	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.02668°N, 8.88250°E
S34	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.01517°N, 8.88777°E
S35	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.01586°N, 8.88914°E
S36	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.01103°N, 8.88029°E
S37	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.01449°N, 8.87612°E
S38	Isola Rossa - Costa Paradiso, Sardinia	Italy	41.05372°N, 8.94518°E
S44	La Crau, Var	France	43.16317°N, 6.09292°E
S47	Sperlonga, Latium	Italy	41.25847°N, 13.43976°E
S57	Cittadella Universitaria, Catania, Sicily	Italy	37.52546°N, 15.07199°E
S59	Cittadella Universitaria, Catania, Sicily	Italy	37.52546°N, 15.07199°E
S61	Località Balzi Rossi, Ventimiglia, Liguria	Italy	43.78361°N, 7.53638°E
Spal1	Pallanca Garden, Bordighera, Liguria	Italy	43.78835°N, 7.68749°E
Spal2	Pallanca Garden, Bordighera, Liguria	Italy	43.78839°N, 7.68736°E
Shan1	Hanbury Garden, Ventimiglia, Liguria	Italy	43.78408°N, 7.55429°E
Shan2	Hanbury Garden, Ventimiglia, Liguria	Italy	43.78445°N, 7.55415°E
Españ 1	Passeig Maritim de la Barceloneta, Barcelona	Spain	41.38474°N, 2.19592°E

Table 1. Location of the 32 sampling sites for *Scyphophorus acupunctatus* in Europe. Coordinates are expressed in UTM WGS84.

The Kimura-2-parameters nucleotide substitution model was selected by jModelTest 2 (Darriba et al. 2012) with the Akaike Information Criterion (AIC) and corrected for rate heterogeneity amongst sites with a Gamma distribution. The NJ was performed by MEGA 11 software with 10,000 bootstrap replicates (Tamura et al. 2021). The BI analysis was performed with MrBayes v.3.12 (Ronquist and Huelsenbeck 2003), using the best model selected. Four chains of Markov Chain Monte Carlo were simultaneously run and sampled every 1000 generations for 4 million generations. The first 1000 sampled trees from each run were discarded as burn-in. The ML phylogenetic analysis was conducted with SeaView software (Gouy et al. 2010). Outgroups (*Dryophthorus corticalis* (Paykull), Stromboscerini sp. and *Aclees taiwanensis* Kôno) were selected in line with their close phylogenetic placement within the family to the study taxon. Nucleotide diversity, haplotype diversity, number of parsimony-informative and variable sites were

						· /		
Table	2. Accession	numbers	of sequences	used for t	the phylogenetic	reconstructio	ons of <i>Scypho</i>	phorus

Accession	Sampling location	Sampling country	Population	Reference
number	1 0	18 2	status	
AY131110	Not available	Continental USA	Native	Direct submission to GenBank
AY131122	Massachusetts	Continental USA	Native	Direct submission to GenBank
GBCL49633-19	California	Continental USA	Native	Direct submission to BOLD Systems
HM433616	Colorado	Continental USA	Native	Direct submission to GenBank
KU896920	Arizona	Continental USA	Native	Chamorro et al. (2016)
KU896921	Arizona	Continental USA	Native	Chamorro et al. (2016)
KU896922	Arizona	Continental USA	Native	Chamorro et al. (2016)
KU896923	Arizona	Continental USA	Native	Chamorro et al. (2016)
KU896924	Arizona	Continental USA	Native	Chamorro et al. (2016)
JX134898	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
JX134899	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
JX134900	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134901	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134902	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134903	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134904	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134905	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134906	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134907	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134908	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134909	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
IX134910	Not available	Mexico	Native	Azuara-Dominguez et al. (2013)
ASSCR6360-12	Not available	Costa Rica	Most likely native	Direct submission to BOLD Systems
ASSCR6362-12	Not available	Costa Rica	Most likely native	Direct submission to BOLD Systems
KU896927	Not available	Guatemala	Most likely native	Chamorro et al. (2016)
KU896929	Not available	Guatemala	Most likely native	Chamorro et al. (2016)
00198464	La Crau	Continental	Non native	Present work
UQ198404	La Clau	France	INOII-Hative	Flesent work
OQ198455	Corsica	France	Non-native	Present work
OQ193159	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ193160	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ193161	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ193162	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ193165	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ193176	Isola Rossa – Costa Paradico, Sardinia	Italy	Non-native	Present work
OQ193177	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ194007	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work
OQ194008	Isola Rossa – Costa Paradiso, Sardinia	Italy	Non-native	Present work

acupunctatus.

Accession Sampling location Sampling country Population Ret	ference
number status	
OQ194015 Isola Rossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	
OQ194016 Isola Rossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	
OQ198466 Isola Rossa – Costa Italy Non-native Press	ent work
Paradiso, Sardinia	
OQ194025 Isola Rossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	
OQ194031 Isola Rossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	
OQ194033 Isola Rossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	
OQ198456 Isola Rossa – Costa Italy Non-native Press	ent work
Paradiso, Sardinia	
OQ198458 Isola Rossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	1
OQ198459 Isola Kossa – Costa Italy Non-native Prese	ent work
Paradiso, Sardinia	
Danadico Sandinia	ent work
CO10/017 Polzi Dossi Italy Non nativo Dros	one moule
Ventimiolia Liguria	ent work
OO198461 Pallanca Cardens Italy Non-native Press	ent work
Liouria	cht work
OO198457 Pallanca Gardens. Italy Non-native Press	ent work
Liguria	
OO193174 Hanbury Gardens, Italy Non-native Press	ent work
Liguria	
OQ198462 Hanbury Gardens, Italy Non-native Press	ent work
Liguria	
OQ194018 Catania, Sicily Italy Non-native Prese	ent work
OQ194019 Catania, Sicily Italy Non-native Prese	ent work
OQ198463 Sperlonga, Latium Italy Non-native Prese	ent work
OQ193157 Villamaniscicle Spain Non-native Press	ent work
OO193158 Villamaniscicle Spain Non-native Press	ent work
OO193175 Passeig Maritim Spain Non-native Press	ent work
de la Barceloneta.	
Barcelona	
MW520550 Porto Santo Portugal Non-native Stüben	et al. (2021)
HM433615 Not available Virgin Islands Non-native Direct submit	ssion to GenBank
KU896925 Not available Virgin Islands Non-native Chamore	o et al. (2016)
KU896926 Not available Virgin Islands Non-native Chamore	o et al. (2016)
KU896928 Not available Virgin Islands Non-native Chamorr	o et al. (2016)
KU896931 Not available Virgin Islands Non-native Chamore	o et al. (2016)
KU896932 Not available Virgin Islands Non-native Chamorr	o et al. (2016)

computed both for the native and the invaded ranges of *S. acupunctatus* through Mega XI (Tamura et al. 2021). A Templeton, Crandall and Sing (TCS) parsimony Network (Clement et al. 2000) connecting haplotypes was obtained with popART (http://popart. otago.ac.nz, Accessed on 20.12.2022) with the aim to visualise the relationship amongst the new and previously-described mitochondrial haplotypes (see Sciandra et al. (2022)).

Results

Species distribution

Overall, the agave weevil was reported on all continents, except for Antarctica. Based on genetic analyses and literature, the native range of this species includes the USA, Mexico and, most likely, the rest of continental Central America (Vaurie 1971). The invasive range of this species (Fig. 1a) includes four South American countries (Brazil, Colombia, Ecuador and Venezuela), the insular USA (including Hawaii and Virgin Islands), Caribbean islands, southern European countries (Portugal including Madeira, Spain including Canary and Balearic Islands, Italy including Sardinia, Sicily and several small islands, Greece including Aegean Islands, Croatian islands and Cyprus), South Africa, Kenya, Tanzania, Saudi Arabia, Java, Sumatra, Borneo and southern Australia (Fig. 1b). Occurrences from other countries (i.e. Israel, the Netherlands, UK and Argentina) were not confirmed and may represent single interceptions.

Species distribution models

Projections of each statistical model (Suppl. material 1: fig. S5) produced slightly different results that were averaged in the ensemble model. The ensemble model for current climate showed a high climatic suitability in the native range and in some parts of the non-native range, i.e. the eastern areas of South Africa, the northern Rift Valley (i.e. from Eritrea and Ethiopia to Tanzania), parts of South America and the central and western Mediterranean countries (Fig. 2a). Highly suitable areas were also predicted in southern Australia, where *S. acupunctatus* has been scarcely recorded so far, the area around the Caspian Sea and the Middle East, where the weevil has not yet been recorded (Fig. 2a).

Considering future climate scenarios forecast for 2070, the areas suitable for *S. acupunctatus* would increase especially towards temperate-cold latitudes, both in Europe and worldwide (Fig. 2b).

Values representing the degree of climatic similarity between future and present conditions are shown in Fig. 2c, with an increase in suitability of 72.62% and a decrease of 27.43%, based on the number of cells around the Globe. There was agreement between the different algorithms used to predict the species' distribution under future climates (Fig. 2d; Suppl. material 1: fig. S5). The climate suitability of most temperate areas of both Hemispheres will increase for *S. acupunctatus* in the future.

The MESS analysis showed that the projection area shared a medium to high environmental similarity with many countries in the training area, except for a few northern Eurasian areas (Suppl. material 1: fig. S6).

Genetic analyses

The COXI sequences were obtained from all analysed samples. All sequences generated in the present study were deposited in GenBank (Table 2). The alignment of COXI gene



Figure 1. a Worldwide distribution of *Scyphophorus acupunctatus* in both native (central and southern North America) and non-native ranges (n = 1135 occurrences) **b** distribution of *S. acupunctatus* in southern European Countries (orange dots refer to occurrence sites of agave weevil). The white dotted line includes occurrences from the native range, whereas the solid red line includes occurrences of uncertain origin. Occurrences outside dotted lines are non-native populations. Sources: Data SIO, NOAA, US Navy, NGA, GEBCO 2016 TerraMetrics 2016 Google; Wikimedia Commons, user Norman Einstein, CC-BY-SA-3.0.

consists of 627–903 nucleotides for 65 individuals, including 32 from the Mediterranean area. All individuals belonged to *S. acupunctatus*, as no record of *S. yuccae* was confirmed in the analysed samples nor in any sequence deposited in the GenBank. Nucleotide and haplotype diversity was lower in the alien than in the non-native range (Table 3).

An ML tree is presented in Fig. 3 and supports the monophyly of *S. acupunctatus* (Fig. 3). Samples from the native range (Mexico and Continental USA) clustered to-



Figure 2. a Current potential distribution of *Scyphophorus acupunctatus* worldwide (suitability increasing from pink to black) **b** future potential distribution of *S. acupunctatus* under climate projections using the global climate model for 2070 (suitability increasing from pink to black) **c** differences between future and present conditions [future-current] for the RCP 2.6 scenario obtained by subtracting, for each cell, the predicted suitability in the future d consensus change for RCP 2.6 scenario. Dark blue (+4) indicates that all models predicted an increase in suitability, whereas dark orange (-4) indicates a full agreement in predicting a decrease in suitability; white indicates disagreement across models (0 value).

Table 3. Indices of genetic divers	ity for native and most-likely	v native (n = 26 sample	es) and non-native
(n = 39 samples) populations of Sa	yphophorus acupunctatus (cf. T	Гable 2).	

	Total	Native and most-likely	Alien
		native populations	populations
π (nucleotide diversity index \pm standard deviation)	0.22 ± 0.05	0.59 ± 0.05	0.03 ± 0.01
h (haplotype diversity index \pm standard deviation)	0.42 ± 0.15	0.61 ± 0.19	0.09 ± 0.01
Number of segregating sites	170	161	115
Number of Parsimony Informative sites	154	148	71

gether and represented the sister group of the clade that included samples from southern countries of Central America (Costa Rica and Guatemala) and all the non-native range (Fig. 3).

The TCS network highlighted that the majority of introduced individuals in Sardinia, Sicily, Corsica, continental Italy (Latium and Liguria), continental France, Spain and Portugal belonged to the same haplotype, as in Costa Rica and Guatemala (Fig. 4).

Discussion

This study showed for the first time the actual and potential global distribution of the agave weevil, both in the native and non-native ranges and assessed the phylogenetic relationships between native and non-native populations at the global scale.



Figure 3. Maximum Likelihood (ML) phylogenetic tree obtained from the analysis of COXI for 65 individuals of *Scyphophorus acupunctatus* (n = 39 from non-native range, n = 22 from native range, n = 4 from most-likely native range, cf. Table 2). The statistical support of major clades is shown at their nodes (NJ Bootstrap support/Bayesian probabilities/ML Bootstrap support).



Figure 4. *Scyphophorus acupunctatus* TCS Network showing the relationship amongst mitochondrial COXI haplotypes. Circles represent different haplotypes (n = 10). Circle size is proportional to the number of samples for each haplotype. Mutations are shown as hatch marks.

The presence of this species was confirmed in several countries, whereas some of those listed in CABI's overview of invasive species (the Netherlands, UK, Israel, New Zealand and Argentina: CABI (2023)) were not confirmed. In contrast, the occurrence of non-native *S. acupunctatus* was reported for the first time in Ecuador, through the iNaturalist repository, as well as in many Italian regions where this weevil was previously not reported (Calabria, Sardinia, Piedmont and Campania: Suppl. material 1: fig. S7). In particular, the first record of this weevil in Sardinia, in the north-western part of the island, was due to detailed and addressed research by the authors of this work.

Despite being reported as the most important pest for agave species (Waring and Smith 1986), the agave weevil is not commonly identified by the public; thus, it is unsuitable for citizen-science surveys (cf. Mazza et al. (2020) for *C. ayyari*). Accordingly, most data on the distribution of this species were obtained from scientific research and insect monitoring projects conducted by specialists (Kontodimas and Kallinikou 2010; Vassiliou and Kitsis 2015). The occurrence of the agave weevil was also confirmed in all the other regions where it was previously reported, i.e. Sicily, Basilicata, Apulia, Latium, Tuscany and Liguria, as well as some small Thyrrenian islands (i.e. Giglio, Elba, Giannutri and Ponza).

The presence of the agave weevil in other Italian peninsular regions along the coastline (e.g. Molise, Abruzzo, Marche, Emilia Romagna and Veneto) cannot be ruled out. Thus, a focused monitoring programme is required, particularly in late spring and during the daytime, when most observations occur (López-Martínez et al. 2011; Figueroa Castro et al. 2013).

Species distribution modelling showed a high climatic suitability for this species throughout the Mediterranean Basin, potentially increasing with increasing temperature and decreasing precipitation, i.e. with the ongoing climatic change. Accordingly, the native range of *S. acupunctatus* currently includes mostly dry areas of Central

America, also suggesting the adaptation of this insect to hot desert areas (including mountainous ones), where most Agavaceae, i.e. succulent plants representing the staple of its diet and reproductive sites, grow. The distribution of *S. acupunctatus* in Europe and Africa is linked to the distribution of Agavaceae and Dracaenaceae as ornamental plants. Particularly, in the Mediterranean countries, these plants mostly occur in botanical gardens and along the coastline, i.e. where most records of *S. acupunctatus* have been reported (Smith and Figueiredo 2007; Celesti Grapow et al. 2016; Cascone et al. 2021).

Genetic analyses showed a strong genetic uniformity for the non-native populations. A lower nucleotide and haplotype diversity was observed in the non-native range compared to the native range, possibly due to a founder effect. The presence of a single widespread haplotype in Europe suggested that most of the introductions may have originated from a small geographical area in Central America or a small number of introduction events occurred. This contrasts with other species, which were introduced through multiple unintentional introductions in Europe. These include *C. ayyari, H. halys* and *Megachile (Callomegachile) sculpturalis* Smith, which show a high genetic diversity linked to several introduction events (Cesari et al. 2018; Mazza et al. 2020; Lanner et al. 2021). *Scyphophorus acupunctatus* in Europe may have originated from one or a few introduction events from Central America (most likely from Guatemala or Costa Rica) through the ornamental plant trade (Global Invasive Species Database 2023). This is in line with EPPO's report (EPPO 2022b), which traces the source of the first introduction of agave weevil to Italy to countries of southern Central America, based on interception data.

Driving definite conclusions from single-gene analyses may be misleading. However, the largest genetic library for *S. acupunctatus* built in the present study may serve as a comparison for future studies and for species identification (Azuara-Domínguez et al. 2013; Chamorro et al. 2016). All analysed samples belonged to *S. acupunctatus* and the only deposited sequence of the sister species *S. yuccae* on GenBank suggests that this species could be a synonym to *S. acupunctatus*, as already hypothesised by Chamorro et al. (2016).

In general, our data showed a high climatic suitability for *S. acupunctatus* in Eurasia and Africa (particularly in the Mediterranean Basin coastline), including areas where this weevil is not yet present. This suggests that if no management actions are taken to limit its spread, there is potential for range expansion towards continental and temperate Europe in the upcoming years. Given the impacts on cultivated agave plants, early detection of this species in new areas should be promoted to prevent further invasions, by means of free online citizen-science platforms and coordination of phytosanitary services and national institutions for the prevention of biological invasions.

Acknowledgements

This work was supported by CNR: Research project FOE – Capitale Naturale e Risorse per il Futuro dell'Italia and Progetto di Ricerca@CNR – USEit Utilizzo di sinergie operative per lo studio e la gestione integrata di specie aliene invasive in Italia.

AC gratefully acknowledges Project ALIEM APOSTROPHE "Action pour Limiter les risques de diffusion des espèces Introduites Envahissantes en Méditerranée" PC IFM 2014–2020 for financial support. Authors would like to thank E. Colonnelli, M. Depratis, L. Forbicioni, E. Giroux, S. Longo, L. Nuccitelli, E. Vandel, C. Berquier and J. Ventura who collected, provided or identified samples of Scyphophorus acupunctatus. We wish also to thank G. Mazza and E. Mori who provided us with four unpublished genetic sequences from Liguria. Moreover, we also thank M. Baratti and E. Paoletti, who allowed us to conduct genetic analyses at the CNR-IRET laboratories in Sesto Fiorentino. The Italian Legislative Decree 19/2021 ("Rules for the protection of plants from harmful organisms") imposes that any previously unrecorded species in any Italian region must be immediately reported to the National Phytosanitary Service before any publication (both scientific and newspaper articles). Therefore, updated information on the distribution of this species in Italy has been sent to all Directors of Regional Phytosanitary Service before this publication. We are indebted with L. Pasquali, L. Ancillotto, M. Di Febbraro, L. Bosso, M. Falaschi and D. Strubbe, who provided us with deep help in species distribution model analyses. To conclude, we would like to thank the Subject Editor, Dr. Katelyn Faulkner and two reviewers for the insightful comments they provided on our early manuscript.

References

- Aiello-Lammens ME, Boria RA, Radosavljevic A, Vilela B, Anderson RP (2015) spThin: An R package for spatial thinning of species occurrence records for use in ecological niche models. Ecography 38(5): 541–545. https://doi.org/10.1111/ecog.01132
- Alin A (2010) Multicollinearity. Wiley Interdisciplinary Reviews: Computational Statistics 2(3): 370–374. https://doi.org/10.1002/wics.84
- Ancillotto L, Strubbe D, Menchetti M, Mori E (2016) An overlooked invader? Ecological niche, invasion success and range dynamics of the Alexandrine parakeet in the invaded range. Biological Invasions 18(2): 583–595. https://doi.org/10.1007/s10530-015-1032-y
- Ancillotto L, Bosso L, Smeraldo S, Mori E, Mazza G, Herkt M, Galimberti A, Ramazzotti F, Russo D (2020) An African bat in Europe, *Plecotus gaisleri*: Biogeographic and ecological insights from molecular taxonomy and Species Distribution Models. Ecology and Evolution 10(12): 5785–5800. https://doi.org/10.1002/ece3.6317
- Ancillotto L, Viviano A, Baratti M, Sogliani D, Ladurner E, Mori E (2023) Every branch in its niche: Intraspecific variation in habitat suitability of a widely distributed small mammal, the harvest mouse *Micromys minutus*. Mammal Research 68(4): 575–585. https://doi. org/10.1007/s13364-023-00693-3
- Andrade MM (2022) The presence of the Agave weevil Scyphophorus acupunctatus Gyllenhal, 1838 (Coleoptera: Dryophthoridae) in Madeira Archipelago. A new biological control opportunity or a new invasive species? Weevil News 98: 1–2.
- Aquino-Bolaños T, Ortiz-Hernández YD, Martiínez-Gutiérrez GA (2013) Relationship between *Scyphophorus acupunctatus* Gyllenhal damage and nutrient and sugar content of

Agave angustifolia Haw. The Southwestern Entomologist 38(3): 477–486. https://doi.org/10.3958/059.038.0310

- Araújo MB, New M (2007) Ensemble forecasting of species distributions. Trends in Ecology & Evolution 22(1): 42–47. https://doi.org/10.1016/j.tree.2006.09.010
- Atauchi PJ, Peterson AT, Flanagan J (2018) Species distribution models for Peruvian plantcutter improve with consideration of biotic interactions. Journal of Avian Biology 49(3): jav-01617. https://doi.org/10.1111/jav.01617
- Azuara-Domínguez A, Cibrián-Tovar J, Terán-Vargas AP, Segura-León OL, Cibrián-Jaramillo A (2013) Factors in the response of Agave Weevil, *Scyphophorus acupunctatus* (Coleoptera: Curculionidae), to the major compound in its aggregation pheromone. The Southwestern Entomologist 38(2): 209–220. https://doi.org/10.3958/059.038.0206
- Banks NC, Paini DR, Bayliss KL, Hodda M (2015) The role of global trade and transport network topology in the human-mediated dispersal of alien species. Ecology Letters 18(2): 188–199. https://doi.org/10.1111/ele.12397
- Baratti M, Goti E, Messana G (2005) High level of genetic differentiation in the marine isopod *Sphaeroma terebrans* (Crustacea Isopoda Sphaeromatidae) as inferred by mitochondrial DNA analysis. Journal of Experimental Marine Biology and Ecology 315(2): 225–234. https://doi.org/10.1016/j.jembe.2004.09.020
- Barber RA, Ball SG, Morris RK, Gilbert F (2022) Target-group backgrounds prove effective at correcting sampling bias in Maxent models. Diversity & Distributions 28(1): 128–141. https://doi.org/10.1111/ddi.13442
- Biondi A, Guedes RNC, Wan FH, Desneux N (2018) Ecology, worldwide spread, and management of the invasive South American tomato pinworm, *Tuta absoluta*: Past, present, and future. Annual Review of Entomology 63(1): 239–258. https://doi.org/10.1146/annurevento-031616-034933
- Bolaños TA, Velázquez EP, Hernández UÁ, Gamboa JRD (2014) Host plants of the agave weevil Scyphophorus acupunctatus (Gyllenhal) (Coleoptera: Curculionidae) in Oaxaca, Mexico. The Southwestern Entomologist 39(1): 163–169. https://doi.org/10.3958/059.039.0115
- Buisson L, Thuiller W, Casajus N, Lek S, Grenouillet G (2010) Uncertainty in ensemble forecasting of species distribution. Global Change Biology 16(4): 1145–1157. https://doi. org/10.1111/j.1365-2486.2009.02000.x
- Buonincontri MP, Bosso L, Smeraldo S, Chiusano ML, Pasta S, Di Pasquale G (2023) Shedding light on the effects of climate and anthropogenic pressures on the disappearance of *Fagus sylvatica* in the Italian lowlands: Evidence from archaeo-anthracology and spatial analyses. The Science of the Total Environment 877: 162893. https://doi.org/10.1016/j. scitotenv.2023.162893
- CABI (2023) *Scyphophorus acupunctatus* (agave weevil). https://www.cabidigitallibrary.org/ doi/10.1079/cabicompendium.49421 [Accessed on 10.02.2023]
- CABI/EPPO (2014) *Scyphophorus acupunctatus* [Distribution map]. Distribution Maps of Plant Pests. CABI, Wallingford, Map 66 (2nd revision). https://doi.org/10.1079/DMPP/20153006587
- Cancellario T, Laini A, Wood PJ, Guareschi S (2023) Among demons and killers: Current and future potential distribution of two hyper successful invasive gammarids. Biological Invasions 25(5): 1627–1642. https://doi.org/10.1007/s10530-023-03000-y

- Cascone S, Sperandii MG, Cao Pinna L, Marzialetti F, Carranza ML, Acosta ATR (2021) Exploring temporal trends of plant invasion in Mediterranean coastal dunes. Sustainability (Basel) 13(24): 13946. https://doi.org/10.3390/su132413946
- CBD [Convention for Natural Biodiversity] (2010) What are Invasive Alien Species? https://www.cbd.int/invasive/WhatareIAS.shtml [Accessed on 09.01.2023]
- Celesti-Grapow L, Bassi L, Brundu G, Camarda I, Carli E, D'Auria G, Del Guacchio E, Domina G, Ferretti G, Foggi B, Lazzaro L, Mazzola P, Peccenini S, Pretto F, Stinca A, Blasi C (2016) Plant invasions on small Mediterranean islands: An overview. Plant Biosystems 150(5): 1119–1133. https://doi.org/10.1080/11263504.2016.1218974
- Cesari M, Maistrello L, Piemontese L, Bonini R, Dioli P, Lee W, Chang-Gyu P, Partsinevelos GK, Rebecchi L, Guidetti R (2018) Genetic diversity of the brown marmorated stink bug *Halyomorpha halys* in the invaded territories of Europe and its patterns of diffusion in Italy. Biological Invasions 20(4): 1073–1092. https://doi.org/10.1007/s10530-017-1611-1
- Chamberlain S, Ram K, Hart T (2017) Spocc: interface to species occurrence data sources (version 0.7.0). https://cran.r-project.org/web/packages/spocc/index.html [Accessed on 10.05.2023]
- Chamorro ML, Persson J, Torres-Santana CW, Keularts J, Scheffer SJ, Lewis ML (2016) Molecular and morphological tools to distinguish *Scyphophorus acupunctatus* Gyllenhal, 1838 (Curculionidae: Dryophthorinae): a new weevil pest of the endangered century plant, *Agave eggersiana* from St. Croix, US Virgin Islands. Proceedings of the Entomological Society of Washington 118(2): 218–243. https://doi.org/10.4289/0013-8797.118.2.218
- Cianferoni F, Graziani F, Dioli P, Ceccolini F (2018) Review of the occurrence of *Halyomorpha halys* (Hemiptera: Heteroptera: Pentatomidae) in Italy, with an update of its European and World distribution. Biologia 73(6): 599–607. https://doi.org/10.2478/s11756-018-0067-9
- Clement M, Posada D, Crandall KA (2000) TCS: A computer program to estimate gene genealogies. Molecular Ecology 9(10): 1657–1660. https://doi.org/10.1046/j.1365-294x.2000.01020.x
- Cocco A, Brundu G, Berquier C, Andreï-Ruiz MC, Pusceddu M, Porceddu M, Podda L, Satta A, Petit Y, Floris I (2021) Establishment and new hosts of the non-native seed beetle *Stator limbatus* (Coleoptera, Chrysomelidae, Bruchinae) on acacias in Europe. NeoBiota 70: 167–192. https://doi.org/10.3897/neobiota.70.70441
- Colombo M (2000) First record of *Scyphophorus acupunctatus* (Coleoptera Curculionidae) in Italy. Bollettino di Zoologia Agraria e di Bachicoltura 32: 165–170.
- Crase B, Liedloff A, Vesk PA, Fukuda Y, Wintle BA (2014) Incorporating spatial autocorrelation into species distribution models alters forecasts of climate-mediated range shifts. Global Change Biology 20(8): 2566–2579. https://doi.org/10.1111/gcb.12598
- Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: More models, new heuristics and parallel computing. Nature Methods 9(8): 772. https://doi.org/10.1038/nmeth.2109
- De Marco Jr P, Diniz-Filho JAF, Bini LM (2008) Spatial analysis improves species distribution modelling during range expansion. Biology Letters 4(5): 577–580. https://doi. org/10.1098/rsbl.2008.0210
- Deutsch CA, Tewksbury JJ, Tigchelaar M, Battisti DS, Merrill SC, Huey RB, Naylor RL (2018) Increase in crop losses to insect pests in a warming climate. Science 361(6405): 916–919. https://doi.org/10.1126/science.aat3466
- Di Cola V, Broennimann O, Petitpierre B, Breiner FT, D'Amen M, Randin C, Engler R, Pottier J, Pio D, Dubuis A, Pellissier L, Mateo RG, Hordijk W, Salamin N, Guisan A (2017) ecospat: An R package to support spatial analyses and modeling of species niches and distributions. Ecography 40(6): 774–787. https://doi.org/10.1111/ecog.02671
- Elith J, Phillips SJ, Hastie T, Dudík M, Chee YE, Yates CJ (2011) A statistical explanation of MaxEnt for ecologists. Diversity & Distributions 17(1): 43–57. https://doi.org/10.1111/ j.1472-4642.2010.00725.x
- EPPO (2008) First record of *Scyphophorus acupunctatus* in France. EPPO Reporting Service 11: 2008/220. https://gd.eppo.int/reporting/article-843 [Accessed on 23.12.2022]
- EPPO (2022a) EPPO Global database. EPPO Global database, Paris, France: EPPO, 1 pp. https://gd.eppo.int/ [Accessed on 10.12.2022]
- EPPO (2022b) Update on the situation of *Scyphophorus acupunctatus* in Italy. EPPO Reporting Service 7: 2022/149. https://gd.eppo.int/reporting/article-7380 [Accessed on 23.12.2022]
- ESRI (2011) ArcGIS Desktop: Release 10. Environmental Systems Research Institute, Redlands, California, USA.
- Farina P, Mazza G, Benvenuti C, Cutino I, Giannotti P, Conti B, Bedini G, Gargani E (2020) Biological notes and distribution in Southern Europe of *Aclees taiwanensis* Kôno, 1933 (Coleoptera: Curculionidae): a new pest of the fig tree. Insects 12(1): 5. https://doi. org/10.3390/insects12010005
- Fick SE, Hijmans RJ (2017) WorldClim 2: New 1 km spatial resolution climate surfaces for global land areas. International Journal of Climatology 37(12): 4302–4315. https://doi. org/10.1002/joc.5086
- Figueroa-Castro P, Solís-Aguilar JF, González-Hernández H, Rubio-Cortés R, Herrera-Navarro EG, Castillo-Márquez LE, Rojas JC (2013) Population dynamics of *Scyphophorus acupunctatus* (Coleoptera: Curculionidae) on blue agave. The Florida Entomologist 96(4): 1454–1462. https://doi.org/10.1653/024.096.0425
- Figueroa-Castro P, Rodríguez-Rebollar H, González-Hernández H, Solís-Aguilar JF, del Real-Laborde JI, Carrillo-Sánchez JL, Rojas JC (2016) Attraction range and inter-trap distance of pheromone-baited traps for monitoring *Scyphophorus acupunctatus* (Coleoptera: Dryophthoridae) on blue agave. The Florida Entomologist 99(1): 94–99. https://doi. org/10.1653/024.099.0117
- Fischer D, Thomas SM, Niemitz F, Reineking B, Beierkuhnlein C (2011) Projection of climatic suitability for *Aedes albopictus* Skuse (Culicidae) in Europe under climate change conditions. Global and Planetary Change 78(1–2): 54–64. https://doi.org/10.1016/j.gloplacha.2011.05.008
- Flinch JM, Alonso-Zarazaga MA (2007) El picudo negro de la pita o agave, o max del henequén, *Scyphophorus acupunctatus* Gyllenhal, 1838 (Coleoptera: Dryophthoridae): primera cita para la Península Ibérica. Boletin de la SEA 41: 419–422.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294–299.
- Foster DR, Motzkin G, Bernardos D, Cardoza J (2002) Wildlife dynamics in the changing New England landscape. Journal of Biogeography 29(10–11): 1337–1357. https://doi. org/10.1046/j.1365-2699.2002.00759.x

- Fourcade Y, Engler J, Rödder D, Secondi J (2014) Mapping species distributions with MAXENT using a geographically biased sample of presence data: A performance assessment of methods for correcting sampling bias. PLoS ONE 9(5): e97122. https://doi.org/10.1371/journal.pone.0097122
- Geier PW (1966) Management of insect pests. Annual Review of Entomology 11(1): 471–490. https://doi.org/10.1146/annurev.en.11.010166.002351
- Global Invasive Species Database (2023) Species profile: *Scyphophorus acupunctatus*. http://www. iucngisd.org/gisd/speciesname/Scyphophorus+acupunctatus [Accessed on 15-01-2023]
- Gouy M, Guindon S, Gascuel O (2010) SeaView version 4: A multiplatform graphical user interface for sequence alignment and phylogenetic tree building. Molecular Biology and Evolution 27(2): 221–224. https://doi.org/10.1093/molbev/msp259
- Guevara L, Gerstner BE, Kass JM, Anderson RP (2018) Toward ecologically realistic predictions of species distributions: A cross-time example from tropical montane cloud forests. Global Change Biology 24(4): 1511–1522. https://doi.org/10.1111/gcb.13992
- Guzmán NV, Lanteri AA, Confalonieri VA (2012) Colonization ability of two invasive weevils with different reproductive modes. Evolutionary Ecology 26(6): 1371–1390. https://doi. org/10.1007/s10682-012-9564-4
- Hao T, Elith J, Guillera-Arroita G, Lahoz-Monfort JJ (2019) A review of evidence about use and performance of species distribution modelling ensembles like BIOMOD. Diversity & Distributions 25(5): 1–14. https://doi.org/10.1111/ddi.12892
- Horsák M, Limondin-Lozouet N, Juřičková L, Granai S, Horáčková J, Legentil C, Ložek V (2019) Holocene succession patterns of land snails across temperate Europe: East to west variation related to glacial refugia, climate and human impact. Palaeogeography, Palaeoclimatology, Palaeoecology 524: 13–24. https://doi.org/10.1016/j.palaeo.2019.03.028
- Huxman TE, Huxman KA, Stamer MR (1997) Dispersal characteristics of the yucca weevil (*Scyphophorus yuccae*) in a flowering field of *Yucca whipplei*. The Great Basin Naturalist 1: 38–43. https://www.jstor.org/stable/41712972
- Inghilesi AF, Mazza G, Cervo R, Cini A (2015) A network of sex and competition: The promiscuous mating system of an invasive weevil. Current Zoology 61(1): 85–97. https://doi. org/10.1093/czoolo/61.1.85
- Iturbide M, Bedia J, Herrera S, del Hierro O, Pinto M, Gutiérrez JM (2015) A framework for species distribution modelling with improved pseudo-absence generation. Ecological Modelling 312: 166–174. https://doi.org/10.1016/j.ecolmodel.2015.05.018
- Kass JM, Muscarella R, Galante PJ, Bohl CL, Pinilla-Buitrago GE, Boria RA, Soley-Guardia M, Anderson RP (2021) ENMeval2.0: Redesigned for customizable and reproducible modeling of species' niches and distributions. Methods in Ecology and Evolution 12(9): 1602–1608. https://doi.org/10.1111/2041-210X.13628
- Kock N, Lynn G (2012) Lateral collinearity and misleading results in variance-based SEM: An illustration and recommendations. Journal of the Association for Information Systems 13(7): 1–40. https://doi.org/10.17705/1jais.00302
- Kontodimas DC, Kallinikou E (2010) First record of the sisal weevil Scyphophorus acupunctatus (Coleoptera: Curculionidae) in Greece. Entomologia Hellenica 19(1): 39–41. https://doi. org/10.12681/eh.11594

- Lanner J, Gstöttenmayer F, Curto M, Geslin B, Huchler K, Orr MC, Pachinger B, Sedivy C, Meimberg H (2021) Evidence for multiple introductions of an invasive wild bee species currently under rapid range expansion in Europe. BMC Ecology and Evolution 21(1): 1–15. https://doi.org/10.1186/s12862-020-01729-x
- López-Martínez V, Alia-Tejacal I, Andrade-Rodríguez M, De Jesús García-Ramírez M, Rojas JC (2011) Daily activity of *Scyphophorus acupunctatus* (Coleoptera: Curculionidae) monitored with pheromone-baited traps in a field of Mexican tuberose. The Florida Entomologist 94(4): 1091–1093. https://doi.org/10.1653/024.094.0458
- Martín-Taboada A, Muñoz AR, Díaz-Ruiz F (2019) Updating the distribution of the exotic agave weevil *Scyphophorus acupunctatus* Gyllenhal, 1838 (Coleoptera: Curculionidae) in peninsular Spain. Anales de Biología 41(41): 49–53. https://doi.org/10.6018/analesbio.41.07
- Mattson W, Vanhanen H, Veteli T, Sivonen S, Niemelä P (2007) Few immigrant phytophagous insects on woody plants in Europe: Legacy of the European crucible? Biological Invasions 9(8): 957–974. https://doi.org/10.1007/s10530-007-9096-y
- Mazza G, Nerva L, Strangi A, Mori E, Chitarra W, Carapezza A, Mei M, Marianelli L, Roversi PR, Campanaro A, Cianferoni F (2020) Scent of jasmine attracts alien invaders and records on citizen science platforms: multiple introductions of the invasive lacebug *Corythauma ayyari* (Drake, 1933) (Heteroptera: Tingidae) in Italy and the Mediterranean basin. Insects 11(9): 620. https://doi.org/10.3390/insects11090620
- Meyerson LA, Mooney HA (2007) Invasive alien species in an era of globalization. Frontiers in Ecology and the Environment 5(4): 199–208. https://doi.org/10.1890/1540-9295(2007)5[199:IASIAE]2.0.CO;2
- Montalva J, Sepulveda V, Vivallo F, Silva DP (2017) New records of an invasive bumble bee in northern Chile: Expansion of its range or new introduction events? Journal of Insect Conservation 21(4): 657–666. https://doi.org/10.1007/s10841-017-0008-x
- Montoya-Jiménez JC, Valdez-Lazalde JR, Ángeles-Perez G, De Los Santos-Posadas HM, Cruz-Cárdenas G (2022) Predictive capacity of nine algorithms and an ensemble model to determine the geographic distribution of tree species. iForest-Biogeosciences and Forestry 15: 363. https://doi.org/10.3832/ifor4084-015
- Mori E, Rustici P, Dapporto L, Pasquali L, Petrucci F, Mazza G (2023) Invasions by the palm borer moth *Paysandisia archon* in Italy and assessment of its trophic spectrum. Biological Invasions 25(5): 1373–1386. https://doi.org/10.1007/s10530-022-02981-6
- Mui AB (2015) A multi-temporal remote sensing approach to freshwater turtle conservation. PhD Thesis, University of Toronto, Toronto, 140 pp.
- Naimi B, Araújo MB (2016) sdm: A reproducible and extensible R platform for species distribution modelling. Ecography 39(4): 368–375. https://doi.org/10.1111/ecog.01881
- Naimi B, Hamm NAS, Groen TA, Skidmore AK, Toxopeus AG (2014) Where is positional uncertainty a problem for species distribution modelling? Ecography 37(2): 191–203. https://doi.org/10.1111/j.1600-0587.2013.00205.x
- Palemón-Alberto F, Castañeda-Vildozola Á, Reyes-García G, Domínguez-Monge S, Ramírez YR, Toledo-Hernández E, Toribio-Jiménez J, Terrones-Salgado J, Ortega-Acosta C, Cruz-Lagunas B, Vargas-Ambrosio LF (2022) Damage by *Scyphophorus acupunctatus* Gyllenhal

in Species of Agave. The Southwestern Entomologist 47(2): 437–442. https://doi.org/10.3958/059.047.0219

- Parsons SE, Kerner LM, Frank SD (2020) Effects of native and exotic congeners on diversity of invertebrate natural enemies, available spider biomass, and pest control services in residential landscapes. Biodiversity and Conservation 29(4): 1241–1262. https://doi. org/10.1007/s10531-020-01932-8
- Pecchi M, Marchi M, Burton V, Giannetti F, Moriondo M, Bernetti I, Bindi M, Chirici G (2019) Species distribution modelling to support forest management. A literature review. Ecological Modelling 411: 108817. https://doi.org/10.1016/j.ecolmodel.2019.108817
- Pernek M, Cvetković S (2022) First record of the agave weevil Scyphophorus acupunctatus Gyllenhal (Coleoptera, Curculionidae) in Croatia. Entomologia Croatica 21(1): 25–32. https:// doi.org/10.17971/ec.21.1.4
- Pohjankukka J, Pahikkala T, Nevalainen P, Heikkonen J (2017) Estimating the prediction performance of spatial models via spatial k-fold cross validation. International Journal of Geographical Information Science 31(10): 2001–2019. https://doi.org/10.1080/13658816.2 017.1346255
- Prakash P (2019) Testing equivalency of interpolation derived bioclimatic variables with actual precipitation: A step towards selecting more realistic explanatory variables for Species Distribution Modelling. Research Journal of Chemistry and Environment 23: 38–41.
- QGIS Development Team (2019) QGIS Geographic Information System. Open-Source Geospatial Foundation Project. http://qgis.osgeo.org. [Accessed on 1 April 2023]
- R Core Team (2019) R: a Language and Environment for Statistical Computing. R Foundation for Statistical Computing. https://www.R-project.org [Accessed on 10.05.2023]
- Regos A, Gómez-Rodríguez P, Arenas-Castro S, Tapia L, Vidal M, Domínguez J (2020) Modelassisted bird monitoring based on remotely sensed ecosystem functioning and atlas data. Remote Sensing (Basel) 12(16): 2549. https://doi.org/10.3390/rs12162549
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics (Oxford, England) 19(12): 1572–1574. https://doi.org/10.1093/ bioinformatics/btg180
- Rotllan-Puig X, Traveset A (2021) Determining the minimal background area for species distribution models: MinBAR package. Ecological Modelling 439: 109353. https://doi. org/10.1016/j.ecolmodel.2020.109353
- Sciandra C, Mori E, Solano E, Mazza G, Viviano A, Scarfò M, Bona F, Annesi F, Castiglia R (2022) Mice on the borders: Genetic determinations of rat and house mouse species in Lampedusa and Pantelleria islands (Southern Italy). Biogeographia – The Journal of Integrative Biogeography 37(1): a013. https://doi.org/10.21426/B637155716
- Setliff GP, Anderson JA (2011) First record of the agave snout weevil, Scyphophorus acupunctatus Gyllenhal (Coleoptera: Curculionidae: Dryophthorinae), in Puerto Rico. Insecta Mundi 152: 1–3.
- Smith GF, Figueiredo E (2007) Naturalized species of Agave L. (Agavaceae) on the southeastern coast of Portugal. Haseltonia 13: 52–60. https://doi.org/10.2985/1070-0048(2007)13[52:NSOALA]2.0.CO;2

- Smith AB, Alsdurf J, Knapp M, Baer SG, Johnson LC (2017) Phenotypic distribution models corroborate species distribution models: A shift in the role and prevalence of a dominant prairie grass in response to climate change. Global Change Biology 23(10): 4365–4375. https://doi.org/10.1111/gcb.13666
- Soroker V, Colazza S (2017). Handbook of Major Palm Pests: Biology and Management. John Wiley & Sons, New York, 316 pp. https://doi.org/10.1002/9781119057468
- Srivastava V, Roe AD, Keena MA, Hamelin RC, Griess VC (2021) Oh the places they'll go: Improving species distribution modelling for invasive forest pests in an uncertain world. Biological Invasions 23(1): 297–349. https://doi.org/10.1007/s10530-020-02372-9
- Steen VA, Tingley MW, Paton PW, Elphick CS (2021) Spatial thinning and class balancing: Key choices lead to variation in the performance of species distribution models with citizen science data. Methods in Ecology and Evolution 12(2): 216–226. https://doi. org/10.1111/2041-210X.13525
- Stüben PE, Schütte A, Astrin JJ (2021) Barcoding and interspecific relationships of Macaronesian Weevils (Coleoptera: Curculionoidea). Beiträge zur Entomologie, Contributions to Entomology 71: 127–135. https://doi.org/10.21248/contrib.entomol.71.1.127-135
- Tamura K, Stecher G, Kumar S (2021) MEGA11: Molecular Evolutionary Genetics Analysis version 11. Molecular Biology and Evolution 38(7): 3022–3027. https://doi.org/10.1093/ molbev/msab120
- Thiede J, Smith GF, Eggli U (2019) Infrageneric classification of Agave L. (Asparagaceae: Agavoideae/Agavaceae): a nomenclatural assessment and updated classification at the rank of section, with new combinations. Bradleya 37(37): 240–264. https://doi.org/10.25223/brad.n37.2019.a22
- Thompson AM, Singh HB, Stewart RW, Kucsera TL, Kondo Y (1997) A Monte Carlo study of upper tropospheric reactive nitrogen during the Pacific Exploratory Mission in the Western Pacific Ocean (PEM-West B). Journal of Geophysical Research 102(D23): 28437–28446. https://doi.org/10.1029/97JD02555
- Thuiller W (2014) Editorial commentary on "BIOMOD optimizing predictions of species distributions and projecting potential future shifts under global change. Global Change Biology 20(12): 3591–3592. https://doi.org/10.1111/gcb.12728
- Valavi R, Elith J, Lahoz-Monfort JJ, Guillera-Arroita G (2019) blockCV: An R package for generating spatially or environmentally separated folds for k-fold cross-validation of species distribution models. Methods in Ecology and Evolution 10(2): 225–232. https://doi. org/10.1111/2041-210X.13107
- Valavi R, Elith J, Lahoz-Monfort JJ, Guillera-Arroita G (2021) Modelling species presenceonly data with random forests. Ecography 44(12): 1731–1742. https://doi.org/10.1111/ ecog.05615
- van Rossem G, van de Bund CF, Burger HC, de Goffau LJW (1981) Bijzondere aantastingen door insekten in 1980. Entomologische Berichten 41: 84–87.
- van Vuuren D, Stehfest E, Den Elzen M, Kram T, van Vliet J, Beltran AM, Oostenrijk R, van Ruijven B (2011) RCP2.6: Exploring the possibility to keep global mean temperature increase below 2 °C. Climatic Change 109(1–2): 95–116. https://doi.org/10.1007/s10584-011-0152-3

- Vassiliou V, Kitsis P (2015) First record of the sisal weevil, *Scyphophorus acupunctatus*, in Cyprus. Entomologia Hellenica 24(1): 22–26. https://doi.org/10.12681/eh.11542
- Vaurie P (1971) Review of Scyphophorus (Curculionidae: Rhynchophorinae). Coleopterists Bulletin 25: 1–8.
- Waring GL, Smith RL (1986) Natural history and ecology of *Scyphophorus acupunctatus* (Coleoptera: Curculionidae) and its associated microbes in cultivated and native agaves. Annals of the Entomological Society of America 79(2): 334–340. https://doi.org/10.1093/ aesa/79.2.334

Supplementary material I

Supplementary data

Authors: Andrea Viviano, Arturo Cocco, Paolo Colangelo, Giuseppe Marco Delitala, Roberto Antonio Pantaleoni1, Laura Loru

Data type: docx

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.101797.suppl1

RESEARCH ARTICLE



Assessing the invasion potential of five common exotic vine species in temperate Australian rainforests

Adam Bernich^{1,2}, Kris French¹, Michael Bedward¹

I Centre for Sustainable Ecosystem Solutions, School of Earth Atmospheric and Life Sciences, University of Wollongong, NSW 2508, Wollongong, Australia 2 Science, Economics and Insights Division, Department of Planning and Environment, Sydney, Australia

Corresponding author: Adam Bernich (Adam.Bernich@environment.nsw.gov.au)

Academic editor: Brad Murray | Received 7 August 2023 | Accepted 19 October 2023 | Published 10 January 2024

Citation: Bernich A, French K, Bedward M (2024) Assessing the invasion potential of five common exotic vine species in temperate Australian rainforests. NeoBiota 90: 79–96. https://doi.org/10.3897/neobiota.90.110659

Abstract

To compare the capacity of native and exotic vine species established under a rainforest canopy, a comparison of growth rates and resource allocation was made amongst five exotic vine species that are serious and common invaders and two common native vine species under two light conditions reflective of edge and interior canopy conditions. All species experienced heavy reductions in growth parameters in the lowlight treatment, but three exotic species showed stronger growth under the low light. All exotic species had higher plasticity in leaf morphology showing a significant increase in SLA under low light. Native vines may have a lower capacity to change leaf morphology in shade, as a result of local adaptation to edge habitats. Higher SLA under both low and high light conditions suggests that exotic vines species are able to exploit a range of forest conditions better than the native species. Three species, *Anredera cordifolia, Araujia sericifera* and *Cardiospermum grandiflorum*, appear particularly capable of invading rainforest interiors. Individuals produced few leaves, focusing resources on roots and stems suggesting a response to reach the canopy quickly. With their long-distance seed dispersal, plasticity in leaf SLA and high RGR, these species appear most likely to invade undisturbed rainforest.

Keywords

Anredera cordifolia, Araujia sericifera, Cardiospermum grandiflorum, Cissus antarctica, Delairea odorata, Ipomoea cairica, low light, Pandorea pandorana, SLA

Copyright Adam Bernich et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

In forests, vines can cause structural damage to the canopy, reduce light availability and increase competition for underground resources, which results in reduced growth and survival of host and neighbouring trees (Estrada-Villegas and Schnitzer 2018). This fast growth strategy is achievable as vines do not invest biomass into self-supporting structures and therefore, they invest more in growth and reproductive structures. Compared to other woody plants, vines tend to be situated on the "faster end" of the life-history spectrum (Westoby et al. 2004), through having higher specific leaf area (SLA) (Llorens and Leishman 2008; Mello et al. 2020), higher photosynthetic capacity (Mello et al. 2020) and more investment into stem growth (Ichihashi and Tateno 2015). The difference in traits between vines and trees, as well as the negative impacts of vines on trees is well studied, especially in tropical systems (Ingwell et al. 2010; Estrada-Villegas and Schnitzer 2018; Mello et al. 2020); however, there is also overlap in trait values between some vines and trees, as well as amongst species of invasive and native vine species (Llorens and Leishman 2008; Osunkoya et al. 2010, 2014; French et al. 2017; Mello et al. 2020).

In a meta-analysis of 117 studies, Van Kleunen et al. (2010) compared traits across studies comparing invasive and native plants species that co-occurred, many co-familial. Invasive plant species had higher SLA, higher photosynthetic capability and faster growth rates than native species (Van Kleunen et al. 2010). There is some evidence that the pattern is also true when vines are considered separately. Greenhouse studies in tropical areas (Osunkoya et al. 2010, 2014) showed that four common invasive vine species in eastern Australia have higher photosynthetic capability, SLA and phenotypic plasticity than for common functionally similar native species, although relative growth rates were not different. This suggests that invasiveness is associated with a set of traits that provide a competitive advantage. However, there are other factors that influence invasiveness, such as release from herbivores (Keane and Crawley 2002) and whether the invasive environment is adequate to successfully reproduce and disperse (Van Kleunen et al. 2015). We predict that, if native vines show similar growth and plasticity to invading exotic invasive vines, then other factors, such as release from herbivores and pathogens may be particularly important to allow them to invade and become abundant. Furthermore, if a particular set of traits are critical in invasiveness, then we would predict that native species which can dominate sites and be extremely abundant would show similar invasive traits. A study in a temperate, moist sclerophyll forest found little difference in above-ground biomass allocation between invasive and native vine species (Llorens and Leishman 2008), which could indicate that other factors give exotic invasive vines a competitive advantage. Furthermore, there was no difference found in the height reached or percentage of host height reached between exotic invasive species and native species, suggesting that there was little difference in impact on the community between exotic invasive and native vine species.

Increased investment in growth can be allocated to roots, stems or leaves. Higher investment into stem elongation can be particularly beneficial as it allows the vine to reach higher into the canopy quickly and, thus, gain more light for photosynthesis (Falster and Westoby 2003; Llorens and Leishman 2008). However, investment into stem elongation must be balanced adequately with the other needs of the vine to survive and reproduce. If exotic invasive vines are released from herbivores, then stem elongation to get their leaves into the light would be the most beneficial strategy. Therefore, we predict that exotic invasive vine species are likely to invest more into stem growth than leaf growth compared to native vine species.

While vine invasion is particularly problematic at the light-filled edges of rainforest patches, the ability to grow and establish in the forest interior would be an invasive characteristic that increases the risk and impacts of that species in closed forests. The plasticity to change growth parameters in low-light situations is, thus, an important part of identifying exotic invasive species that pose the greatest risk. Some invasive vine species are considered more problematic in forests than others due to apparent high growth rates or high propagule pressure. In Australia, there are at least 179 species of exotic vines (Harris et al. 2007), though not all are considered equal threats to biodiversity. It is likely that the species that should be considered to be greater threats should have higher photosynthetic ability, invest more into stem elongation and, consequently, have higher growth rates as well as show high rates of growth even in interior light conditions. Furthermore, knowledge of how invasive and native vine species differ in these traits with light availability can help to predict the species that can exploit disturbances better or can invade undisturbed interiors. At present, information on growth rates and resource allocation under differing light levels is poorly understood for all exotic vine species in Australia, preventing good risk assessments and prioritisation in management. Collectively, invasion and establishment of exotic vines and scramblers are listed as a key threatening process in New South Wales (NSW) where 38 entities are listed as affected by vines and scramblers. However the list of vines and scramblers includes species that are much less abundant and problematic and some that occur frequently (A. Bernich and K. French, unpublished data).

In order to help evaluate the invasiveness of key exotic invasive vine species, we assessed growth rates of five common invasive exotic vine species in eastern Australia and two common native species on host trees in a shade house with two shade treatments. We measured relative growth rates (RGR), stem lengths, proportion of biomass allocated to leaves, stems and roots and specific leaf area. We predicted that individuals of each species grown in less shade will have higher RGR and longer stem lengths, though lower SLA. We predicted that invasive species will also have higher RGR and stem lengths and higher SLA indicating higher photosynthetic efficiency, which then leads to relatively less investment in leaves and more relative investment into stems and roots compared to native species. We also predicted that differences in trait values would occur amongst species and show that some species are able to exploit establishment opportunities under the rainforest canopy. One of the native species, *Cissus antarctica*, is especially abundant at the edge of disturbed rainforests in eastern Australia where it can dominate and smother canopy causing significant harm to native host species. There is concern that such dominance may cause forest interiors to become degraded.

Methods

Study species

We grew seven vine species, five exotic invasive species (Anredera cordifolia, Araujia sericifera, Cardiospermum grandiflorum, Delairea odorata and Ipomoea cairica) and two native species (Cissus antarctica and Pandorea pandorana). All species are commonly found in rainforests, wet sclerophyll forests and disturbed sites on the east coast of Australia. Anredera cordifolia (family Basellaceae) is a semi-succulent twiner from South America, that is listed as a Weed of National Significance in Australia. It was introduced in the early 1900s (Vivian-Smith et al. 2007) and quickly establishes at sites through vegetative reproduction via aerial tubers. Araujia sericifera (family Apocynaceae) is a South American twiner also introduced in the early 1900s, which disperses by producing masses of windblown seeds. Cardiospermum grandiflorum (family Sapindaceae) is a tendril climber from South America, introduced in the 1920s, with multiple seeds in a papery "balloon" that can be distributed by wind and water. Delairea odorata (family Asteraceae) is an herbaceous twiner or scrambler from Africa that was introduced in the early 1900s, that spreads locally through vegetative reproduction (stolons and stem fragments), although it also produces viable seeds in Australia. Ipomoea cairica (family Convolvulaceae) is a twiner from tropical Africa and Asia and was introduced in the 1840s, it is common in coastal eastern Australia and can withstand a wide range of environmental conditions (Liu et al. 2016a). The native Cissus antarctica (family Vitaceae) is a tendril climber that can grow thick stems (up to 15 cm in diameter) and is abundant along the east coast of Australia, producing a fleshy fruit. Pandorea pandorana (family Bignoniaceae) is a large woody twiner that is also abundant in eastern Australia, producing winged seeds.

Vine species were all collected from forests near Wollongong, with some species being grown as ~ 30 cm cuttings (*D. odorata, I. cairica, P. pandorana* and *C. antarctica*), from seeds (*C. grandiflorum*), tubers (*A. cordifolia*) or harvested seedlings (*A. sericifera*) which had the first two true leaves, around 5–15 cm in height. We attempted to grow both *C. grandiflorum* and *A. sericifera* as cuttings, though no *C. grandiflorum* cuttings were successful and there was only a 10% success rate for *A. sericifera*. Propagules for all species were collected in September 2021 and were grown until sufficient individuals were established to be used in the experiment. All propagules were collected along forest edges (i.e. tracks or clearings) which were more representative of the medium light treatment (see below). As establishment times varied amongst species, the date that species were potted and placed next to a host tree occurred over two months at the beginning of the Austral Summer (late October – mid December).

All vine individuals were grown on *Acmena smithii* (cultivar 'Speedy Screener', family Myrtaceae) host trees that were potted in 300 mm pots. The host trees ranged in height from 1 m to 1.8 m tall. *Acmena smithii* is a common tree in eastern Australia that grows in rainforests and wet sclerophyll forests.

Experimental design

All vines and host trees were grown in a shade house at the University of Wollongong, NSW 34.4054°S, 150.8784°E. The shade house had two sections, one with low light penetration to mimic the understorey under a rainforest canopy and one with medium light to mimic a gap in a rainforest or rainforest edge. The roof of the low light section was covered in two layers of shade cloth, which allowed 2% of light to reach the floor (similar to 85–95% canopy cover), whereas the medium light section had one layer of medium shade cloth, which allowed 30% of light to reach the floor (similar to 50–60% canopy cover).

Six individuals of each species were randomly selected for the medium light and low light treatments and were transplanted into 300 mm pots filled with commercial potting mix (Osmocote Premium) and given 25 g of slow-release fertiliser (Power-Feed 500 g All Purpose Controlled Release). They were then placed adjacent (on the southern side) to an *A. smithii* individual in their allocated shaded areas. Two to four extra individuals were harvested and dried in an oven at 65 °C for five days to measure dry biomass of roots, stems and leaves at the start of the experiment (the difference in number of individuals for each species was due to the death of some individuals before they could be dried out). Vines and trees were watered by an automatic dripper system attached to a tap timer, with each plant having a dripper spike in the soil of the pot. Plants were drip-watered for 10 minutes at 6 am and 6 pm every day.

The experiment for each species began when plants were placed next to the host plant. Initial plant sizes are shown in Suppl. material 3. Vines were grown for 24 weeks, after which time, vines were removed from host trees and laid horizontally on the ground to measure the longest stem (from the junction of the roots and the stem to the tip of the longest stem, to give a measurement of the potential height each individual could reach). Ten leaves for each vine individual were removed and measured using a Li-Cor leaf area meter (Model Li-3000A, Lincoln, Nebraska, USA), then put into separate labelled bags for drying (65 °C for 5 days), before being weighed and specific leaf area calculated (SLA = leaf area/dry weight). Then, each individual was sorted into leaves, stems, aerial tubers (for *A. cordifolia*) and roots, with roots being washed to remove soil, before being placed in a drying oven at 65 °C for 5 days and then weighed. Relative growth rate (RGR) was calculated using the formula:

$$RGR = \frac{InDW_f - InDW_i}{no.of \, days}$$

where DW_f is the total dry weight at the end of the experiment for an individual and DW_i is the average dry weights of the plants sacrificed at the beginning of the experiment for the species being tested.

The dry weights of each plant part (roots, stems and leaves) were divided by the total dry weight to give percentages of biomass allocation; these parameters are referred

to as root mass fraction (RMF), stem mass fraction (SMF) and leaf mass fraction (LMF). For *A. cordifolia*, aerial tuber weight was added to RMF as a measure of investment into energy storage; however, the proportion of biomass invested into aerial tubers by *A. cordifolia* was also recorded separately. Traits were only measured on individuals that did not die in the experimental period.

Data analysis

We used a Bayesian modelling approach to estimate the distribution of values for each of the measured plant variables for each combination of species and shade treatment. The fitted distributions were then used to estimate the magnitude and direction of differences in response between species within each treatment and between treatments for each species. Stem height and SLA values, which could only be positive, were modelled as gamma-distributed variables with the shape parameter of the distribution being allowed to vary between shade treatments. RGR values were modelled as being drawn from a Student-t distribution since values could be negative and some outliers were evident in the observed data. The shape (degrees of freedom) parameter of the distribution was treated as an unknown quantity to be estimated by the model, while the scale parameter (standard deviation) was allowed to vary between treatments. The proportion of biomass allocated to each of leaf, stem and root fractions was modelled using Dirichlet regression.

Models were fitted by Hamiltonian Monte Carlo sampling via the "brms" package version 2.18 (Bürkner 2017) in R version 4.2.1 (R Core Team 2022). For all models, we set weakly informative prior distributions for parameters, as recommended by Gelman (2009) and van de Schoot (2021), to ensure that fitted distributions reflected the observed data while constraining the model fitting process from exploring unrealistic ranges of parameter values. For the Dirichlet regression of biomass allocation fractions, we explored alternative choices for prior distributions on the intercept and regression coefficients using prior predictive simulation. This involved fitting the model, based only on candidate prior distributions, i.e. with no observed data. The simulations highlighted the potential sensitivity of model predictions to the choice of standard deviation for the Normal priors on the intercept and regression coefficients. Given this, we chose to treat the prior standard deviation as an additional parameter to be learned from the data and set an exponential hyper-prior distribution on it.

For each model, we ran four Markov chains with 5000 iterations and 1000 warmup iterations. Model convergence was assessed using the Gelman-Rubin statistic, which showed convergence for all models and by checking for an adequate number of effectively independent samples to ensure reliable estimates of the tails of the fitted distributions. In addition, we graphed posterior model predictions together with observed data values for each measured variable to check for any disagreement that might indicate a problem with model structure or convergence.

For all models other than stem growth rate, the distribution of differences in response between each pair of species within each shade treatment was estimated by subtracting posterior predictions of mean response for one species from those for the other species. For the stem growth rate model, difference calculations were based on posterior predictions of median rather than mean response as some observed values were close to zero, which resulted in a strongly right-tailed posterior distribution for which the median is a more representative summary statistic.

Results

At the end of the six months, two *A. sericifera* individuals had died in the medium light treatment. In the low light treatment 10 deaths occurred: three *I. cairica* individuals, two *D. odorata*, three *A. sericifera*, one *C. antarctica* and one *P. pandorana*.

Relative growth rate

For all species, mean RGR was consistently higher when grown under medium light compared to low light, with no overlap in the 95% range of predicted mean RGR values (Fig. 1, Table 1). Two exotic vines did poorly in low light, *D. odorata* (93% reduction in mean predicted RGR) and *I. cairica* (75% reduction) and the two native spe-



Figure 1. Mean relative growth rate (RGR) with 95% bounds on the mean predicted values from each model (black lines) and observed RGR value for individual plants (blue dots) for both light treatments. (E) are exotic vines, (N) are native vines.

Table 1. The predicted mean trait values and the 95% bounds on the mean predicted values from each model (smaller font). RGR = relative growth rate (g g⁻¹ day⁻¹), stem growth = increase in stem length per day (cm/day), LMF = leaf mass fraction (%), RMF = root mass fraction (%), SMF = stem mass fraction (%), SLA = specific leaf area (cm²/g), (E) denotes exotic species, (N) denotes native species. * note that RMF for Anredera cordifolia includes the weight of aerial tubers.

Species	Light	Predicted mean trait values								
	treatment	RGR	Stem	LMF	RMF	SMF	SLA			
growth										
Anredera cordifolia	Medium	0.0274	1.55	16.5%	65.8%	17.7%	323.0			
(E)		0.0248-0.0302	1.29-2.31	14.9–22.5	44.3–56	26.3-36.2	272.7-379.4			
	Low	0.0084	0.47	32.9%	42.5%	24.7%	988.1			
		0.0037-0.0129	0.42 - 0.97	22.9-39.5	32.8-51.2	19.8–35.7	814.2-1205.3			
Araujia sericifera (E)	Medium	0.0353	1.14	18.8%	37.0%	44.3%	255.0			
		0.0313-0.0387	0.82-2.05	13.3-23.7	30.8-44.1	37.6–51.4	209.83-302.27			
	Low	0.0141	0.24	11.3%	52.7%	36.0%	625.8			
		0.0085-0.0208	0.14-0.81	6.4-21.1	36.6-67.4	20.8-50.1	463.8-824.1			
Cardiospermum	Medium	0.0257	1.62	24.4%	24.1%	51.5%	329.5			
grandiflorum (E)		0.0227-0.0290	1.25-2.71	19.7–29.3	19.4–29.1	45.7–57.2	281.2-383.9			
	Low	0.0088	0.48	42.2%	12.6%	45.2%	745.7			
		0.0056-0.0121	0.36-1.24	31.3–52.3	7.3-20.5	34.2–55.7	643.9-864.0			
Delairea odorata (E)	Medium	0.0229	1.92	15.3%	12.5%	72.3%	570.4			
		0.0193-0.0258	1.46-3.20	11.6–19.4	9.2–16.6	66.7–76.9	480.4-663.3			
	Low	0.0017	0.62	25.6%	12.1%	62.3%	1607.5			
		-0.0024 - 0.0059	0.43-1.75	15.6–36.7	5.9-21.3	48.7–73.8	1279.6-2013.8			
Ipomoea cairica (E)	Medium	0.0289	2.62	11.9%	45.7%	42.5%	379.3			
		0.0256-0.0320	2.02-4.25	9.1–16.1	39.6–51.0	36.5-48.0	321.5-443.6			
	Low	0.0071	0.74	29.6%	33.5%	36.9%	1219.6			
		0.0013-0.0153	0.48 - 2.24	16.4-41.8	21.4-49.7	22.7-51.3	957.4-1540.2			
Cissus antarctica	Medium	0.0158	0.80	45.6%	13.4%	41.0%	189.8			
(N)		0.0127-0.0186	0.61–1.32	38.5-50.3	10.5-18.2	36.0-47.1	161.6-219.9			
	Low	-0.0013	0.11	45.2%	25.3%	29.6%	362.7			
		-0.0046 - 0.0023	0.07-0.36	34.8-57.8	15.0-35.5	19.0-40.0	285.2-462.7			
Pandorea pandorana	Medium	0.0134	0.91	30.8%	24.6%	44.6%	281.7			
(N)		0.0100-0.0174	0.69 - 1.54	25.2-35.6	20.2-30.0	38.9–50.5	242.1-327.8			
	Low	0.0022	0.20	42.3%	28.5%	29.2%	482.6			
		-0.0014 - 0.0059	0.14-0.58	31.1–53.9	18.5–39.6	19.0-39.9	389.1-585.4			

cies, *P. pandorana* (84% reduction) and *C. antarctica* (~ 98% reduction) showed large reductions in RGR under low light. *C. antarctica* barely grew in the low light treatment making estimates of percentage reductions difficult to calculate. The three other exotic species had reductions of between 60 and 69%. Higher variability in RGR was seen amongst individuals of exotic invasive species in the low light (see Suppl. material 1).

All the exotic invasive species grown in the medium light treatment had higher predicted mean RGR values than the two native species (Fig. 1, Table 1). *A. sericifera* had the highest predicted mean RGR with only *I. cairica* having a slight overlap in the 95% range of predicted mean values. All other exotic invasive vine species had relatively similar predicted mean RGR values with a high percentage of overlap in the 95% bounds on the predicted mean. In the low light treatment, *A. sericifera* still tended to have the highest RGR, though the 95% bounds on the predicted mean substantially overlapped with three of the other exotic invasive species (Fig. 1, Table 1, Suppl. material 2).

Length of main stem

Similar to RGR, the predicted median stem length grown per day for all species in the medium light treatment was greater than the low light treatment, with no overlap in the 95% bounds on the predicted median values from the model (Fig. 2). In the medium light treatment, *I. cairica* had the highest predicted median stem growth per day at 3.2 cm/day and had final raw stem lengths across individuals of 3.6–8.5 m. *D. odorata* also had high stem growth rates, with a slight overlap on the 95% bounds on the predicted median stem lengths of 3.9–6.4 m. *A. sericifera* had the lowest predicted median stem growths for the exotic invasive species. The two native species clearly had the lowest predicted median stem growth (Fig. 2), with there being a slight overlap in the 95% bounds on the predicted median with *A. sericifera* only (Suppl. material 1).

In the low light treatment, *I. cairica* and *D. odorata* had the highest predicted median stem growth per day with high variability amongst individuals. Other species had closer predicted median values. The gap between exotic invasive and native species in the low light treatment was actually higher than the medium light treatment (Fig. 2), though *A. sericifera* only had slightly higher predicted stem growth than *P. pandorana* at 4 mm per day (Fig. 2). Mean predicted values were very similar to median values (Fig. 2, Table 1).



Figure 2. Median (blue lines) and mean (red line) stem growth rate with 95% bounds on the mean predicted values from each model, for both light treatments. Blue dots show the observed value for individual plants. (E) are exotic vines, (N) are native vines.

Biomass allocation

The percentage of biomass invested into leaves, stems and roots differed amongst species and light treatments (Fig. 3, Table 1). In the medium light treatment, exotic invasive species invested very little into leaves, with all species having a predicted mean proportion of leaf biomass less than 25%, while the two natives, *C. antarctica* and *P. pandorana* had predicted mean leaf biomass proportions of 47% and 31%, respectively. *Anredera cordifolia* and *I. cairica* invested relatively highly into root biomass with the predicted mean being 66% and 46%, respectively. For *A. cordifolia* in the medium light, the mean number of aerial tubers produced was 45.3 (range 29–61). Aerial tubers accounted for between 5 and 30% of the total biomass (mean = 15%). The roots, excluding aerial tubers, accounted for, on average, 55% of total biomass. No aerial tubers were produced by individuals grown in the low light treatment. Two species, *D. odorata* and *C. antarctica* invested very little into roots (Fig. 3). *Delairea odorata* invested the greatest amount into stems (predicted mean of 72%) (see Suppl. material 1).

Biomass allocation for all species in the low light treatment was more varied, seen by wider 95% bounds on the predicted mean values (Fig. 3). Natives did invest more biomass in leaves compared to moderate light treatments, but overall, the allocation to



Figure 3. Mean proportion with 95% bounds (black lines) on the mean predicted values from each model, of biomass invested into leaves, roots and stems for all species in both light treatments. Blue dots are observed proportions for individual plants.

leaves across all vines was similar. *A. sericifera* tended to allocate resources into stems and roots rather than leaves. *Delairea odorata* and *C. grandiflorum* invested very little in roots, focusing on stem growth by comparison (Fig. 3, Table 1, Suppl. material 2).

Specific leaf area

The 95% bounds on the mean predicted range for SLA was substantially higher in the low light treatment than the medium light for all species (Fig. 4, Table 1), with there being no overlap in the 95% bounds on predicted means between treatments for each species (Suppl. material 2). The greatest difference in predicted mean SLA between the treatments was seen in *I. cairica* (222% difference) and *A. cordifolia* (206%). *D. odora-ta* (182%), *A. sericifera* (145%) and *C. grandiflorum* (126%) had a moderate predicted increase in SLA, while the two natives had the lowest difference in SLA amongst treatments; *C. antarctica* (91%), then *P. pandorana* (71%).

In general, exotic invasive species had higher SLA than native species, except for *A. sericifera* in the medium light which had a considerable overlap in the 95% bounds on predicted mean values with *P. pandorana*, though only a slight overlap in the low



Figure 4. Observed specific leaf area (SLA) with 95% bounds (black line) on the mean predicted values from each model for all species in both light treatments. Blue dots are observed values for individual plants. (E) are exotic vines, (N) are native vines.

light treatment. *Delairea odorata* and *I. cairica* had the two highest predicted mean SLA in both treatments (Fig. 4, Table 1). *Cardiospermum grandiflorum* and *A. cordifolia* had similar SLA in the medium light, though *A. cordifolia* had much higher SLA in the low light (Fig. 4, Table 1).

Discussion

All species grew at faster rates under the higher light conditions that are reflective of rainforest edges, suggesting that quick invasion was most likely from disturbed edges or light gaps for all species. Low light conditions slowed growth, but the reduction in growth varied amongst species reflecting a differential risk of invasion and establishment into the rainforest interior. The two common native species had very low growth rates in all light levels, but particularly in the low light. They also showed a limited capacity to vary SLA and improve light capture relative to the invasive species. This suggests they would most likely establish in edges and better lit areas. This is despite one of these species being considered problematic; *Cissus antartica* can significantly smother vegetation along rainforest edges. Our results suggest that this issue will not occur under the canopy.

All exotic invasive species had higher relative growth rates than native species. Relative growth rate and high SLA are correlated with invasiveness (Poorter and Bongers 2006; Leishman et al. 2007; Van Kleunen et al. 2010; Dawson et al. 2011). However, some of the same species were used in Osunkoya et al. (2010) which measured no difference in RGR amongst native and exotic species in the tropics. Our work suggests that, at least in temperate areas, high relative growth rates were a characteristic of our invasive species.

Compared to self-supporting woody plants, the native vines in this study still sit on the "faster" side of the life history spectrum. For example, in a comparison of coexisting tropical trees and vines, Mello et al. (2020) found that the mean SLA for trees was 141 cm²/g (\pm 62.41 SD) and vines was 177 cm²/g (\pm 80.77 SD). In our study, the mean SLA for *C. antarctica* (190 cm²/g) and *P. pandorana* (282 cm²/g) grown in medium light was higher than that of the vines measured by Mello et al. (2020). Nevertheless, the difference in SLA between the exotic invasive and native species in this study may be due to natives investing more into leaf defence from herbivores leading to lower SLA, leaving the exotic invasives at an advantage with fewer herbivores in their introduced range (Keane and Crawley 2002).

Coupled with the higher growth rates measured, all exotic invasive vine species showed flexibility in leaf SLA when grown in low light conditions. All had higher SLA values under low light and were higher than both native species in all light conditions. Interestingly, the means in low light were associated with a great deal of variability amongst individuals (large 95% confidence intervals). Invasive exotic species, therefore, show plasticity in their responses to low light conditions, increasing the size of their leaves relative to the leaf biomass to increase light capture capacity. As a result, in comparison to the native species, all exotic invasive species will be capable of better

light harvesting under rainforest canopies, increasing photosynthesis and growth rates. Furthermore, increased SLA, even under medium light conditions, may also help them in forest gaps as they grow leaves better suited to the light environment they are in and, therefore, may be able to respond to canopy disturbances better (Liu et al. 2016b). Phenotypic plasticity of leaves is a trait often observed in invasives (Daehler 2003; Richards et al. 2006; Davidson et al. 2011) and Osunkoya et al. (2014) also found that leaf plasticity of exotic invasive vine species (including *A. cordifolia* and *A. sericifera*) was higher than phylogenetically similar native vine species. Therefore, while more native and exotic vine species need to be compared, it is likely that higher plasticity in leaves correlates with invasiveness and explains why exotic invasive vines often dominate in disturbed forests with patchy canopy cover, but may also increase their capability to invade low light less-disturbed closed forests.

We identified three growth strategies amongst the exotic invasive vines that we investigated, with regards to their risk to rainforest communities. These strategies may be a more general approach for other species, but further species would need to be considered to establish such strategies. Thus our descriptions of a strategy highlight some of the differences in growth responses of the exotic species we tested which may increase risk of invasion. The first strategy was associated with fast growth, exemplified by D. odorata and I. cairica which showed high SLAs and high mean stem growth rates under both canopy and edge conditions. Having fast stem growth rates and high stem biomass allocation is beneficial for vines as it allows individuals to compete with others through early access to canopy light (Falster and Westoby 2003; Llorens and Leishman 2008). High SLA provides leaves with more efficient energy capture to invest into growth (Poorter and Bongers 2006), although it also indicates that these species are likely to be short-lived (Westoby et al. 2004). D. odorata also invested very little into roots and did not produce woody stems suggesting a quick strategy with low competitive capacity. However, both of these species grow horizontal running stems which can form roots and support new stems, perhaps advantageous in capturing more area quickly under higher light conditions. This allows them to spread quickly and persist despite leaves (and perhaps stems) being replaced regularly. These species will be quick to establish both under canopy and at edges, but are likely to be more successful in forest gaps and edges and may do poorly through time under the canopy.

A second, more long-term invasion strategy was evident in the three other exotic species. These three had the highest relative growth rates under low light conditions, providing opportunities for invasion even within undisturbed rainforest patches; *Anredera cordifolia, Araujia sericifera* and *Cardiospermum grandiflorum*. While some *A. sericifera* individuals may be particularly effective at growing under the canopy, we also recorded some mortality in low light conditions. Surviving individuals produced few leaves, focusing resources on roots and stems suggesting a response to reach the canopy as quickly as possible. High dispersal capability using large numbers of windblown seeds (Vivian-Smith and Panetta 2005), coupled with high RGR in higher light, explains why *A. sericifera* is listed as a major weed in multiple countries (Vivian-Smith and Panetta 2005), where it damages vegetation and restricts regeneration, as well being poisonous to humans and livestock (NSW DPI 2020). Germination has been shown to be significantly reduced in cool (< 20 °C) and dark conditions (Vivian-Smith and Panetta 2005), suggesting that the maintenance of canopy cover can decrease establishment. Our results suggest that if it can establish under canopy, then it may well persist increasing its risk in rainforest interiors. If given the right conditions, *A. seric-ifera* is likely the species to build biomass fastest out of the species grown in this study and presents one of the highest invasion risks under rainforest canopies.

One other invasive species in this group of potential understorey invaders was Anredera cordifolia. This species was also able to have quite high RGRs under low light increasing its allocation to leaves and stems in this environment, compared to the location at the edges of rainforests. It also had much greater flexibility in changing leaf light capture under the canopy, compared to A. sericifera. At edges in ideal conditions, it can maximise growth rates through having amphistomatous leaves and high numbers of stomata (Boyne et al. 2013), increasing photosynthetic ability and efficiency. A. cordifolia, however, utilises an additional strategy. Under better growing conditions, resources are used to develop canopy-held tubers which are a robust dispersal strategy, providing new vegetative recruits which receive greater resources maternally than germinants. The strategy enables the species to persist through inclement conditions (Vivian-Smith et al. 2007). In the six months of this experiment, individuals in the medium light produced an average of 45 aerial tubers, highlighting the high propagule pressure of this species, as well as the speed in which it can reach reproductive maturity, which is a trait often used to explain invasiveness and feasibility of control (Panetta and Grigg 2021). Therefore, A. cordifolia can persist under canopies through reallocation of resources to leaves and stems and can spread throughout a site, particularly from the edges, making it difficult to eradicate and contain (Panetta and Grigg 2021). Tubers (included within the root component) resulted in lower allocation to stem growth, but provided a capacity to persist at the edges and create new individuals increasing invasion and risk. This places this species as one of the highest risk species under rainforest canopies.

Conclusion

Our work measured important growth parameters for a range of exotic vines and we were able to identify different strategies that influence how invasive exotic vine species may invade rainforests. While all species perform better in higher light conditions reflective of rainforest edges and gaps, the capacity to maintain higher levels of growth under rainforest canopies showed that some species may well establish and persist causing host tree damage within the rainforest. Coupled with vegetative growth strategies, we identified differences in the capacity of species to establish and persist under canopies. We suggest that maintaining canopy health and controlling edges, still remains the key tool for reducing vine invasion, although at least two species, *Araujia sericifera* and *Anredera cordifolia* have characteristics that suggest that directed control within rainforests is needed.

References

- Boyne RL, Osunkoya OO, Scharaschkin T (2013) Variation in leaf structure of the invasive Madeira vine (*Anredera cordifolia*, Basellaceae) at different light levels. Australian Journal of Botany 61(5): 412–417. https://doi.org/10.1071/BT13083
- Bürkner PC (2017) brms: An R package for Bayesian multilevel models using Stan. Journal of Statistical Software 80(1): 1–28. https://doi.org/10.18637/jss.v080.i01
- Daehler CC (2003) Performance Comparisons of Co-Occurring Native and Alien Invasive Plants: Implications for Conservation and Restoration. Annual Review of Ecology, Evolution, and Systematics 34(1): 183–211. https://doi.org/10.1146/annurev.ecolsys.34.011802.132403
- Davidson AM, Jennions M, Nicotra AB (2011) Do invasive species show higher phenotypic plasticity than native species and, if so, is it adaptive? A meta-analysis. Ecology Letters 14(4): 419–431. https://doi.org/10.1111/j.1461-0248.2011.01596.x
- Dawson W, Fischer M, Van Kleunen M (2011) The maximum relative growth rate of common UK plant species is positively associated with their global invasiveness. Global Ecology and Biogeography 20(2): 299–306. https://doi.org/10.1111/j.1466-8238.2010.00599.x
- Estrada-Villegas S, Schnitzer SA (2018) A comprehensive synthesis of liana removal experiments in tropical forests. Biotropica 50(5): 729–739. https://doi.org/10.1111/btp.12571
- Falster DS, Westoby M (2003) Plant height and evolutionary games. Trends in Ecology & Evolution 18(7): 337–343. https://doi.org/10.1016/S0169-5347(03)00061-2
- French K, Robinson S, Smith L, Watts E (2017) Facilitation, competition and parasitic facilitation amongst invasive and native liana seedlings and a native tree seedling. NeoBiota 36: 17–38. https://doi.org/10.3897/neobiota.36.13842
- Gelman A (2009) Bayes, Jeffreys, Prior Distributions and the Philosophy of Statistics. Statistical Science 24(2): 176–178. https://doi.org/10.1214/09-STS284D
- Harris CJ, Murray BR, Hose GC, Hamilton MA (2007) Introduction history and invasion success in exotic vines introduced to Australia. Diversity & Distributions 13(4): 467–475. https://doi.org/10.1111/j.1472-4642.2007.00375.x
- Ichihashi R, Tateno M (2015) Biomass allocation and long-term growth patterns of temperate lianas in comparison with trees. The New Phytologist 207(3): 604–612. https://doi. org/10.1111/nph.13391
- Ingwell LL, Joseph Wright S, Becklund KK, Hubbell SP, Schnitzer SA (2010) The impact of lianas on 10 years of tree growth and mortality on Barro Colorado Island, Panama. Journal of Ecology 98(4): 879–887. https://doi.org/10.1111/j.1365-2745.2010.01676.x
- Keane RM, Crawley MJ (2002) Exotic plant invasions and the enemy release hypothesis. Trends in Ecology & Evolution 17(4): 164–170. https://doi.org/10.1016/S0169-5347(02)02499-0
- Leishman MR, Haslehurst T, Ares A, Baruch Z (2007) Leaf trait relationships of native and invasive plants: Community- and global-scale comparisons. The New Phytologist 176(3): 635–643. https://doi.org/10.1111/j.1469-8137.2007.02189.x
- Liu G, Gao Y, Huang FF, Yuan MY, Peng SL (2016a) The invasion of coastal areas in South China by *Ipomoea cairica* may be accelerated by the ecotype being more locally adapted to salt stress. PLoS ONE 11(2): e0149262. https://doi.org/10.1371/journal.pone.0149262

- Liu Y, Dawson W, Prati D, Haeuser E, Feng Y, Van Kleunen M (2016b) Does greater specific leaf area plasticity help plants to maintain a high performance when shaded? Annals of Botany 118(7): 1329–1336. https://doi.org/10.1093/aob/mcw180
- Llorens AM, Leishman MR (2008) Climbing strategies determine light availability for both vines and associated structural hosts. Australian Journal of Botany 56(6): 527–534. https://doi.org/10.1071/BT07019
- Mello FNA, Estrada-Villegas S, Defilippis DM, Schnitzer SA (2020) Can functional traits explain plant coexistence? A case study with tropical lianas and trees. Diversity 12(10): 1–15. https://doi.org/10.3390/d12100397

NSW DPI (2020) Moth vine (Araujia sericifera). https://weeds.dpi.nsw.gov.au/Weeds/MothVine

- Osunkoya OO, Bayliss D, Panetta FD, Vivian-Smith G (2010) Leaf trait co-ordination in relation to construction cost, carbon gain and resource-use efficiency in exotic invasive and native woody vine species. Annals of Botany 106(2): 371–380. https://doi.org/10.1093/aob/mcq119
- Osunkoya OO, Boyne R, Boyne R, Scharaschkin T (2014) Coordination and plasticity in leaf anatomical traits of invasive and native vine species. American Journal of Botany 101(9): 1423–1436. https://doi.org/10.3732/ajb.1400125
- Panetta FD, Grigg A (2021) A weed risk analytical screen to assist in the prioritisation of an invasive flora for containment. NeoBiota 66: 95–116. https://doi.org/10.3897/ neobiota.66.67769
- Poorter L, Bongers F (2006) Leaf traits are good predictors of plant performance across 53 rain forest species. Ecological Society of America 87: 1733–1743. https://doi.org/10.1890/0012-9658(2006)87[1733:LTAGPO]2.0.CO;2
- R Core Team (2022) R: A language and environment for statistical computing. R Foundation for Statistical Computing. https://www.r-project.org/
- Richards CL, Bossdorf O, Muth NZ, Gurevitch J, Pigliucci M (2006) Jack of all trades, master of some? On the role of phenotypic plasticity in plant invasions. Ecology Letters 9(8): 981–993. https://doi.org/10.1111/j.1461-0248.2006.00950.x
- van de Schoot R, Depaoli S, King R, Kramer B, Märtens K, Tadesse MG, Vannucci M, Gelman A, Veen D, Willemsen J, Yau C (2021) Bayesian statistics and modelling. Nature Reviews. Methods Primers 1(1): 1–1. https://doi.org/10.1038/s43586-020-00001-2
- Van Kleunen M, Weber E, Fischer M (2010) A meta-analysis of trait differences between invasive and non-invasive plant species. Ecology Letters 13(2): 235–245. https://doi. org/10.1111/j.1461-0248.2009.01418.x
- Van Kleunen M, Dawson W, Maurel N (2015) Characteristics of successful alien plants. Molecular Ecology 24(9): 1954–1968. https://doi.org/10.1111/mec.13013
- Vivian-Smith G, Panetta FD (2005) Seedling recruitment, seed persistence and aspects of dispersal ecology of the invasive moth vine, *Araujia sericifera* (Asclepiadaceae). Australian Journal of Botany 53(3): 225–230. https://doi.org/10.1071/BT04118
- Vivian-Smith G, Lawson BE, Turnbull I, Downey PO (2007) The biology of Australian weeds.46. Anredera cordifolia (Ten.) Steenis. Plant Protection Quarterly 22: 1–10.
- Wright IJ, Reich PB, Westoby M, Ackerly DD, Baruch Z, Bongers F, Cavender-Bares J, Chapin T, Cornelissen JHC, Diemer M, Flexas J, Garnier E, Groom PK, Gulias J, Hikosaka K,

Lamont BB, Lee T, Lee W, Lusk C, Midgley JJ, Navas M-L, Niinemets Ü, Oleksyn J, Osada N, Poorter H, Poot P, Prior L, Pyankov VI, Roumet C, Thomas SC, Tjoelker MG, Veneklaas EJ, Villar R (2004) The worldwide leaf economics spectrum. Nature 428: 821–827. https://idp.nature.com/authorize/casa?redirect_uri=https://www.nature.com/articles/nature02403&casa_token=CtYdn6u63BcAAAAA:cNAkoHi669d547DoKaM3keUixX15 dhKLnmyUD_oX2GBEB1VTe9mtWcMdFoXauJPFpb4iQItw5D0oAvb3

Supplementary material I

Difference in mean and 95% bounds, as well as the percentage overlap of mean values reported as a probability (Probability higher) between species for the physiological measurements taken

Authors: Adam Bernich, Kris French, Michael Bedward

Data type: xlsx

- Explanation note: Species are compared within light treatments only. Probability higher reflects the values of the Species 1 being higher than Species 2. Asterisks in Species 1 column denotes exotic species.
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.110659.suppl1

Supplementary material 2

Difference in mean and 95% bounds, as well as the percentage overlap of mean values reported as a probability (Probability higher than LL) for the seven species grown in the study

Authors: Adam Bernich, Kris French, Michael Bedward

Data type: xlsx

Explanation note: Asterisks denote exotic species.

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.110659.suppl2

Supplementary material 3

The dry weight of the individuals used as the initial values to calculate relative growth rates for each species

Authors: Adam Bernich, Kris French, Michael Bedward

Data type: xlsx

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.110659.suppl3

RESEARCH ARTICLE



Root hemiparasites suppress invasive alien clonal plants: evidence from a cultivation experiment

Tamara Těšitelová¹, Kateřina Knotková¹, Adam Knotek¹, Hana Cempírková², Jakub Těšitel¹

I Department of Botany and Zoology, Faculty of Science, Masaryk University, Kotlářská 2, 611 37 Brno, Czech Republic 2 Department of Experimental Biology, Faculty of Science, Masaryk University, Kotlářská 2, 611 37 Brno, Czech Republic

Corresponding author: Tamara Těšitelová (tamara.malinova@centrum.cz)

Academic editor: R. Bustamante | Received 22 September 2023 | Accepted 29 November 2023 | Published 15 January 2024

Citation: Těšitelová T, Knotková K, Knotek A, Cempírková H, Těšitel J (2024) Root hemiparasites suppress invasive alien clonal plants: evidence from a cultivation experiment. NeoBiota 90: 97–121. https://doi.org/10.3897/ neobiota.90.113069

Abstract

Alien invasive plants threaten biodiversity by rapid spread and competitive exclusion of native plant species. Especially, tall clonal invasives can rapidly attain strong dominance in vegetation. Root-hemiparasitic plants are known to suppress the growth of clonal plants by the uptake of resources from their belowground organs and reduce their abundance. However, root-hemiparasites' ability to interact with alien clonal plants has not yet been tested.

We explored the interactions between native root-hemiparasitic species, *Melampyrum arvense* and *Rhinanthus alectorolophus* and invasive aliens, *Solidago gigantea* and *Symphyotrichum lanceolatum*. We investigated the haustorial connections and conducted a pot experiment. We used seeds from wild hemiparasite populations and those cultivated in monostands of the invasive plants to identify a possible selection of lineages with increased compatibility with these alien hosts. The hemiparasitic species significantly suppressed the growth of the invasive plants. *Melampyrum* inflicted the most substantial growth reduction on *Solidago* (78%), followed by *Rhinanthus* (49%). Both hemiparasitic species reduced *Symphyotrichum* biomass by one-third. Additionally, *Melampyrum* reduced the shoot density of both host species. We also observed some transgenerational effects possibly facilitating the growth of hemiparasites sourced from subpopulations experienced with the host.

Native root hemiparasites can effectively decrease alien clonal plants' biomass production and shoot density. The outcomes of these interactions are species-specific and may be associated with the level of clonal integration of the hosts. The putative selection of lineages with higher performance when attached to the invasive novel hosts may increase hemiparasites' efficiency in future biocontrol applications.

Keywords

Asteraceae, biological invasion, biotic resistance, Orobanchaceae, physiological integration, pot experiment, restoration

Introduction

Alien plant invasions represent a component of global change with profound effects on diversity, ecosystem functioning and services. Invasive species broadly vary in their specific impacts on the habitats they invade due to different abilities to spread and achieve dominance or mechanisms of interaction with native biota (Blackburn et al. 2014). Of particular concern are the so-called transformer invaders (Richardson et al. 2000), which can invade indigenous natural communities over large areas, attain high dominance and change ecosystem functioning. Alien tall clonal herbs with below-ground rhizomes are frequent examples of transformer invaders in grasslands due to their increased competitiveness leading to the exclusion of native plants from infested vegetation throughout temperate regions (Divíšek et al. 2018; Wang et al. 2019; Wan et al. 2021; Lanta et al. 2022). Although conventional control management, represented by mowing or grazing, can reduce the density of the invasive clonal herbs to some extent (e.g. Nagy et al. (2020); Szymura et al. (2022)), they are usually not eliminated and may spread rapidly from the rhizomes if the management measures are ceased. More drastic restoration measures (e.g. use of herbicides, long-term shading) may eradicate the invaders, but can also adversely affect native species, making their use problematic in areas with conservation value (e.g. Weber and Jakobs (2005); Szymura et al. (2022)).

Native parasitic plants have recently been suggested as potential biocontrol agents for a wide range of invasive plants globally (Těšitel et al. 2020). Following the biotic resistance hypothesis (Maron and Vilà 2001), generalist native adversaries, such as parasitic plants, may impede the success of invaders due to the lack of defence or tolerance mechanisms of the host plants against parasitism (Cameron and Seel 2007). Clonal hosts could be especially harmed by parasitism, as the parasitic uptake of resources targets the cornerstone of their growth strategy, that is, the spatial spread of vegetative ramets and clonal integration (e.g. Song et al. (2013); Roiloa (2019)), i.e. the transfer of resources amongst interconnected ramets via rhizome network, which facilitates efficient resource acquisition and sharing (Kavanová and Gloser 2005; Gao et al. 2021). However, parasitic plants may turn this advantage into a liability. A parasite that attaches to one ramet may access nutrients within the network, leading to its vigorous growth and potentially marked biomass decline of the clonal host, including the non-infected ramets (Lepš and Těšitel 2015; Gao et al. 2021). This could explain a substantial decrease in the clonal hosts' abundance in the communities with parasitic plants observed in several studies (Decleer et al. 2013; Demey et al. 2015; Somodi et al. 2018). Moreover, field experiments have demonstrated the ability of root-hemiparasitic Rhinanthus species to significantly reduce harmful expansive clonal grass Calamagrostis epigejos from semi-natural grasslands (Těšitel et al. 2017, 2018),

which was consequently introduced to ecological restoration practice (Lukavský 2020; SPPK D02 002 2021).

Amongst parasitic plants, species of root hemiparasites (or, more precisely, Euphytoid parasites in the new parasitic plant classification of Teixeira-Costa and Davis (2021)) appear to be particularly suitable candidates for suppressing clonal invasive species due to their low host specificity (e.g. Matthies (2017, 2021)), capacity to substantially suppress host growth (e.g. Press et al. (2005); Těšitel et al. (2015b); Matthies (2021)) and ability to form dense populations (van Hulst et al. 1987; Mudrák and Lepš 2010; Heer et al. 2018). Despite available evidence on the negative effects of root hemiparasites on expansive species (reviewed by Těšitel et al. (2020)), only one study has investigated the effect of a root-hemiparasitic species on an alien invader (Walder et al. 2019), which, however, did not show any adverse impact of the parasite on the host species. Two reasons may explain this lack of empirical research on interactions between root hemiparasites and alien invaders. First, hemiparasites and alien invaders may not share the same habitats. For instance, in Central Europe, an analysis of habitats of hemiparasitic species identified natural and semi-natural communities as their principal habitats (Těšitel et al. 2015a). These habitats are simultaneously characterised by low levels of alien invasions (Pyšek et al. 2012). Second, establishing a parasitic association with alien invaders may be difficult. Although hemiparasites are mostly host generalists, host quality (i.e. the extent of support of parasite growth) varies between species (e.g. Rowntree et al. (2014); Matthies (2017, 2021)). Native hemiparasitic species lack a common evolutionary history with non-indigenous plants. The lack of experience with an alien host may limit a hemiparasite's efficiency of resource withdrawal on the one hand, but also the host's resistance or tolerance to parasitism on the other, as predicted by the biotic resistance hypothesis. Compatibility with a host may also be affected by high intra- and interpopulation genotypic variability of the annual hemiparasites (Mutikainen et al. 2000; Rowntree et al. 2011; Unachukwu et al. 2017; Rowntree and Craig 2019; Moncalvillo and Matthies 2023). The recognised ability to rapidly evolve ecotypes adapted to various environmental conditions (Zopfi 1993; Pleines et al. 2013) may further facilitate the interaction with novel host species.

In this paper, we investigated the interactions between root-hemiparasitic *Rhinan-thus alectorolophus* and *Melampyrum arvense* (Orobanchaceae) and the alien invasive clonal species *Solidago gigantea* and *Symphyotrichum lanceolatum* (Asteraceae). First, we examined the anatomy of haustoria to determine whether the hemiparasites can form functional parasitic connections with the novel hosts. Second, we set up a comprehensive pot experiment to study the effect of host identity on hemiparasite performance and the impact of hemiparasite infection on the two hosts. We expected to identify differences in vitality (measured by biomass production) of the two hemiparasite species (hypothesis 1), which should be reflected by a difference in host suppression (hypothesis 2). Specifically, we expected lower host suppression by *Rhinanthus*, given its general preference for grass or legume hosts (Matthies 2021), than in *Melampyrum*, which has been shown to flourish when attached to various forbs, including many Asteraceae

(Matthies 2017). Furthermore, we investigated the potential selection of hemiparasite lineages and their effect on host-hemiparasite interactions. To do so, we used seeds from hemiparasites that had grown for two years in monoculture stands of the two host species and compared their performance to plants from the original population from a species-rich grassland ('naïve' plants), i.e. all tested seed sources per hemiparasite species originated from a single hemiparasite population. We hypothesised that growth in a host monoculture might lead to a selection of lineages better adapted to the given host, reflected by improved hemiparasite growth and possibly a more deleterious effect on that host (hypothesis 3).

Materials and methods

Study species

Melampyrum arvense L. and Rhinanthus alectorolophus (Scop.) Pollich are annual xylemfeeding root-hemiparasitic species native to Europe. Melampyrum typically grows in dry grasslands and steppes, while *Rhinanthus alectorolophus* (Scop.) Pollich favours dry to mesic grasslands. *Solidago gigantea* Aiton and *Symphyotrichum lanceolatum* (Willd.) G. L. Nesom are perennial rhizomatous species from the Asteraceae family, originating from North America (Pyšek et al. 2012). They began spreading across Europe in the 19th century and have become serious invaders (Weber and Jakobs 2005; Jedlička and Prach 2006; Axmanová et al. 2021). Solidago and Symphyotrichum are considered typical wetland species, but they also occur in disturbed anthropogenic habitats, poorly-managed fields, pastures and meadows within their native range (Chmielewski and Semple 2001; Weber and Jakobs 2005). Solidago has a broader ecological niche in the invaded areas, also occupying drier and nutrient-poorer soils (Weber and Jakobs 2005). Both species have a perennial rhizome, which, in the spring, produces a cohort of shoots that start to flower in late summer (Solidago) or early autumn (Symphyotrichum) and yield numerous tiny wind-dispersed seeds. Jedlička and Prach (2006) noted the high viability of Symphyotrichum lanceolatum seeds, which, combined with the effective ability to penetrate established vegetation, triggers the high invasive potential of this species.

Haustorial connection

We initiated a pilot cultivation trial to examine the anatomy of haustorial connections between the hemiparasites and the two invasive hosts. The cultivation was set up in the experimental garden of the Department of Botany and Zoology at Masaryk University in Brno, Czech Republic. The hemiparasites' seeds were collected from species-rich vegetation in the summer of 2019 (see Suppl. material 1: appendix S1 for localisations). In the autumn of 2019, we transplanted rhizomes of host species into $15 \times 15 \times 20$ cm pots (narrower at the bottom, corresponding to 3.6 litres), filled with a mixture

of peat and garden soil (ratio 1:3). In October, we sowed 20 hemiparasite seeds on each pot. We established five replicates for each hemiparasite-host combination.

In June 2020, we rinsed the hosts' roots, harvested the haustoria and preserved them in 70% ethanol. Following the method of Soukup and Tylová (2014), we dehydrated the samples, transferred them to anhydrous butanol, infiltrated and then embedded them in paraffin. We prepared 12 μ m sections using a sliding microtome (Reichert, Wien, Austria) and de-waxed and stained them with phloroglucinol-HCl (Wiesner solution) (Liljegren 2010) to colour the lignified cell walls.

Cultivation experiment

We established the main pot experiment in autumn 2021 to investigate and quantify the outcome of the novel interactions for the hemiparasites and the extent of host suppression. For each hemiparasitic species, we used three seed sources: (i) seeds from a wild population growing in a species-rich grassland and (ii) seeds from plants originally obtained from the same populations as in (i), but which had been growing since 2019 in monostands of the two invasive host species. The aim was to investigate the potential selection of genotypes more adapted to the specific hosts. More specifically, the monostands were mown in the autumn of 2019, after which we sowed the hemiparasites' seeds. In 2020, the monostands with hemiparasites were mown in July and October. We collected ripe hemiparasite seeds from all populations from June to July 2021. The seeds were stored at room temperature before use. As both host species produce a dense rhizome network in the topsoil layer, we collected soil blocks with rhizomes from monostands of each host species to establish host cultivation in September 2021. First, we removed the above-ground biomass and then cut approx. 12×12 cm rhizome blocks with a spade. The rhizomes were then inserted into the same pots and soil substrate described in the chapter 'Haustorial connection'. See Suppl. material 1: appendix S1 for GPS coordinates of the sites of hemiparasites' seed and host plants' origin.

The experimental design comprised: (i) an uninfected control treatment (host species without hemiparasite seed addition) and three types of 'infected' treatments (with hemiparasite seeds addition), i.e. treatments (ii) 'naïve' (seeds of hemiparasites originating from a wild population), (iii) 'home' (seeds from hemiparasites growing for two years in a monostand of a host species and then sown with the same host species in the pot) and (iv) 'cross' (seeds from hemiparasites growing for two years in a monostand of one host species and then sown into the pot with the other host species) (see the scheme of the origin of hemiparasites' seeds in Fig. 1). Both hemiparasites were sown with both invasive species, resulting in 14 treatments. Each treatment consisted of 10 replicates of the pots, totalling 140 pots. Each pot in the 'infected' treatments (treatments ii–iv) received 40 seeds of one of the hemiparasitic species. Seeds were spread on the surface and gently mixed with the topsoil layer. The pots were then placed in the experimental garden in Brno, following a completely randomised design and irrigated. During spring 2022, the pots were irrigated as necessary. In April 2022, seedlings of



Figure 1. Scheme of origin of the hemiparasites' seeds used in the cultivation experiment. In October 2019, seeds of *Melampyrum arvense* and *Rhinanthus alectorolophus* from a single population per species, originating from a species-rich grassland, were sown in monostands of the host species *Solidago gigantea* and *Symphyotrichum lanceolatum*. By 2021, hemiparasite seeds collected from the host species' monostands and the original hemiparasite population were used in the cultivation experiment resulting in three types of hemiparasite seed sources: 'naïve', 'home' and 'cross'.

non-target species were removed from the pots. The pots were spaced 30 cm apart and their position within the experimental matrix was changed three times before harvest at the beginning of June 2022.

The experiment was harvested during hemiparasite flowering. We cut the aboveground biomass and counted the number of host shoots and hemiparasitic plants that survived in each pot. The hemiparasite and host biomass from each pot were dried separately at 80 °C and weighed. Schmid et al. (1995) revealed a strong dependency of sexual reproduction and clonal growth on plant size as well as a threshold size for sexual reproduction in *Symphyotrichum lanceolatum* and *Solidago canadensis*, a species closely related to *Solidago gigantea*. We thus expected the vegetative biomass to reflect host fitness and reproductive potential sufficiently.

Statistical analyses

Initially, we conducted an exploratory analysis of patterns in counts of hemiparasite individuals, host ramets and above-ground biomass production to identify pots that were not representative due to insufficient host or hemiparasite recruitment. Only pots with at least six host shoots and three hemiparasite individuals (in infected treatments) were subsequently included in the analysis (n = 132 out of 140 pots). Scatterplots of biomass vs. individual or shoot counts (Suppl. material 1: appendices S2, S3) demonstrated low correlations, indicating compensatory growth in pots retained for the analysis.

We used linear models to analyse the following parameters: hemiparasite aboveground biomass, the number of individuals, mean biomass per individual and host above-ground biomass, the number of shoots and mean biomass per shoot. All variables were log-transformed before analysis to improve the normality of residuals and homogeneity of variances. The analysis of each parameter, used as response variables, was conducted at two levels: (i) the species-level model included hemiparasite, host species and their interaction as predictors. Seed-source treatments were disregarded in this analysis; (ii) seed-source analysis consisted of a series of linear models, one for each host–hemiparasite combination, with seed-source treatment as a single predictor. In this analysis, we set treatment contrasts with the 'naïve' treatment as the baseline level, to which the two other treatments were compared. Only biomass data were tested in the seed-source level analysis.

We first built a saturated model for each analysis with all candidate predictors and interactions. Individual terms of the saturated models were tested by an F-test, the results of which are reported in ANOVA tables as in a classical two-way ANOVA with interactions. Non-significant (P > 0.05) terms were subsequently removed from the models in the backward predictor selection procedure. Non-significant main effects were retained if a predictor was involved in a significant interaction. The resulting minimal adequate models were then used to extract regression coefficients and their associated tests of significance. This approach was allowed by the nature of our data coming from a manipulative experiment with a balanced design, which implies the or-

thogonality of the predictors. We acknowledge that the orthogonality was not perfect because we removed a few pots with low establishment of hosts or parasites. Still, the collinearity between the tested effects (host and parasite predictors) was minimal as measured by the phi-coefficient ($\varphi = 0.026$; $\chi_1 = 0.0084$; P = 0.927), which justifies the validity of the interaction-term testing and supports backward selection as a suitable model-selection approach. All analyses were performed in R, version 4.2.2 (R Core Team 2022).

Results

Functional haustorial connection

Both hemiparasitic species formed fully developed haustoria on the roots and rhizomes of both host species. In all cases, the xylem bridge from hemiparasite haustoria reached the xylem vessels of the hosts. No signs of a defensive reaction by the hosts were observed (Fig. 2).

Host-Hemiparasite interaction on the species level

Hosts successfully resprouted from rhizomes in the transferred soil blocks; only four pots had to be omitted because of insufficient sprouting (Fig. 3, Suppl. material 1: appendix S4). The number of hemiparasite plants varied in the pots, but their establishment was generally successful, with only four pots omitted from the experiment due to poor hemiparasite establishment. On average, 10.9 Melampyrum plants were harvested in pots with both host species (max. 20 individuals). In contrast, significantly higher average numbers of *Rhinanthus* plants, 16.2 and 13.2, were harvested in pots with Solidago and Symphyotrichum (max. 23 individuals), respectively (Table 1, Fig. 4, Suppl. material 2 for the primary data). Hemiparasite biomass production differed between the two species and was also significantly affected by the host identity (Table 1). Specifically, Melampyrum grew larger than Rhinanthus (t₁₁₀ = 11.25, $P < 10^{-6}$) and Solidago supported a more vigorous hemiparasite growth than Symphyotrichum ($t_{110} = 10.12$, $P < 10^{-6}$). These effects were additive, i.e. the difference in the host quality had a similar impact on both hemiparasitic species (Fig. 4). Similar trends and significant interactions were also found concerning the average biomass of hemiparasite individuals (Table 1). Melampyrum individuals were consistently larger than *Rhinanthus* and both hemiparasitic species produced larger specimens on *Solidago* than on Symphyotrichum. However, this trend was less pronounced in Rhinanthus, i.e. Rhinanthus individuals growing with Symphyotrichum were larger than expected by additive effects ($t_{109} = 2.57$, P = 0.012; Fig. 4).

Regarding host suppression, we identified strong interactive effects of host and hemiparasite species identities on the host biomass (Table 1). The hemiparasitic spe-



Figure 2. Cross sections of haustorial connections between two root-hemiparasitic species and their hosts. In the hemiparasite haustoria (ha), there is a hyaline body (hb), the vascular core of the haustorium (vc) and a xylem bridge (xb) leading to host xylem vessels (xv) in the host root (hr); xx – xylem–xylem contact.

cies significantly reduced host biomass relative to uninfected controls (*Melampyrum*: $t_{126} = -10.1$, $P < 10^{-6}$, *Rhinanthus*: $t_{126} = -4.53$, $P < 10^{-4}$), but the suppression was significantly more pronounced in *Solidago* infected by *Melampyrum* ($t_{126} = 4.50$, $P < 10^{-5}$; Fig. 5). Overall, *Solidago* biomass was reduced by 77.6% and 49.1% on average when infected by *Melampyrum* and *Rhinanthus*, respectively. *Symphyotrichum* biomass was reduced by 31.6% and 35.2% on average by *Melampyrum* and *Rhinanthus*, respectively. Host biomass was reduced by decreasing the number of host shoots or reducing the average biomass of host shoots. While *Melampyrum* acted in both ways, *Rhinanthus* mainly decreased the average host shoot biomass (Fig. 5). In detail, *Melampyrum* reduced the number of host shoots per pot ($t_{128} = -4.05$, $P < 10^{-4}$) by 33% in *Solidago* and 21% in *Symphyotrichum*. The effect of *Rhinanthus* on the host shoot number was not significant ($t_{128} = -0.76$, P = 0.45). Both *Melampyrum* ($t_{126} = -7.07$, $P < 10^{-6}$) and *Rhinanthus* reduced the average shoot biomass of both hosts to a similar extent, *Melampyrum* was significantly less deleterious to *Symphyotrichum* than to *Solidago* ($t_{126} = 3.98$, P < 0.001).



Figure 3. Representative pots for each hemiparasite seed-source treatment ('cross', 'home', 'naïve') and the uninfected control. *Solidago gigantea* (left) and *Symphyotrichum lanceolatum* (right) are infected by *Melampyrum arvense* (top) or *Rhinanthus alectorolophus* (bottom). The bottom photo is flipped vertically for clarity of the experiment presentation. Photographic documentation of all experimental pots is provided in Suppl. material 1: appendix S4. Scale bars: 50 cm.

Effect of the hemiparasite seed origin on the interaction

We identified the significant effects of the hemiparasite seed-source treatments on some interactions. Total hemiparasite biomass was affected in the case of *Melampyrum* growing on *Solidago* (R² = 0.29, F_{2,24} = 4.98, P = 0.016) and *Rhinanthus* growing on *Symphyotrichum* (R² = 0.32, F_{2,25} = 5.81, P = 0.008). Specifically, *Melampyrum* plants

Response	Effect	df	Sum Sq.	F	Р
Hemiparasite biomass	Hemiparasite	1	13.72	121.95	< 10 ⁻⁶
	Host	1	11.65	103.48	< 10 ⁻⁶
	Hemiparasite × Host	1	0.24	2.10	0.15
	Residuals	109	12.28		
Hemiparasite count per pot	Hemiparasite	1	3.89	19.64	< 10 ⁻⁴
	Host	1	0.35	1.75	0.19
	Hemiparasite × Host	1	0.29	1.46	0.23
	Residuals	109	21.60		
Hemiparasite average	Hemiparasite	1	32.23	203.10	< 10 ⁻⁶
biomass	Host	1	7.97	50.22	< 10 ⁻⁶
	Hemiparasite × Host	1	1.05	6.61	0.011
	Residuals	109	17.30		
Host biomass	Hemiparasite*	2	16.11	46.12	< 10 ⁻⁶
	Host	1	0.02	0.13	0.72
	Hemiparasite × Host	2	7.69	22.00	< 10 ⁻⁶
	Residuals	126	22.00		
Host shoot count per pot	Hemiparasite*	2	2.53	13.09	< 10 ⁻⁵
	Host	1	5.07	52.42	< 10 ⁻⁶
	Hemiparasite × Host	2	0.31	1.59	0.21
	Residuals	126	12.18		
Host shoot average biomass	Hemiparasite [*]	2	7.28	18.35	< 10 ⁻⁶
	Host	1	5.77	29.10	< 10 ⁻⁶
	Hemiparasite × Host	2	5.04	12.71	< 10 ⁻⁵
	Residuals	126	24.99		

Table 1. Analysis of variance tables summarising the effects of hemiparasite and host species identity on the growth of hemiparasites and hosts.

*The hemiparasite effect on host biomass also comprises non-infected control as an extra level.

in the 'cross' treatment (seeds from plants previously grown with the alternative invasive host) produced significantly less biomass ($t_{24} = -2.80$, P = 0.010) compared to the 'naïve' treatment (seeds from species-rich vegetation), while the biomass of *Melampyrum* on *Solidago* from the 'home' (seeds from plants previously grown with the same host species) and 'naïve' treatment did not significantly differ (Fig. 6). Conversely, the biomass of *Rhinanthus* on *Symphyotrichum* was significantly higher in the 'home' treatment compared to the 'naïve' treatment ($t_{25} = 3.09$, P = 0.005) and the hemiparasite biomass in the 'cross' and 'naïve' treatment did not differ (Fig. 6).

Host biomass was significantly affected only in the case of *Solidago* infected by *Rhinanthus* ($R^2 = 0.27$, $F_{2,27} = 5.09$, P = 0.013) (Fig. 7). Here, *Rhinanthus* of 'home' and 'cross' treatments suppressed *Solidago* biomass more than 'naïve' *Rhinanthus* plants ($t_{27} = -2.73$, P = 0.011 and $t_{27} = -2.80$, P = 0.009 for 'home' and 'cross' treatments, respectively).



Figure 4. Effects of host species on the total biomass, number of individuals per pot and average biomass of the individuals of the two hemiparasitic species. Boxplots represent median, quartiles and ranges. See Table 1 for the ANOVA tables summarising significance tests. Note the logarithmic scale of the y-axes.


Figure 5. Effect of hemiparasite infection on total biomass, number of shoots per pot and average shoot biomass of the two host species. Boxplots represent median, quartiles and non-outlier ranges, with outliers displayed as points outside the non-outlier ranges. Note the logarithmic scale of the y-axes. See Table 1 for the ANOVA tables summarising significance tests.

Discussion

The outcome of the novel host-hemiparasite interactions

Both root-hemiparasitic species established a functional parasitic association with the two novel host species, as evidenced by functional haustorial connection, vital growth



Figure 6. Effect of seed-source treatments on hemiparasite biomass production categorised by the individual host-hemiparasite combinations. Boxplots represent median, quartiles and non-outlier ranges, with outliers displayed as points outside the non-outlier ranges. *P*-values indicate significant effects of seed-source treatments. Note the logarithmic scale of the y-axes.

and flowering of both parasites (Figs 2, 3). In line with our first hypothesis, the hemiparasite species differed in compatibility with the two invasive hosts from Asteraceae, with *Melampyrum* proving a more efficient parasite than *Rhinanthus*. This outcome is not surprising, as *Melampyrum* has previously been shown to thrive when attached to a series of forbs. Asteraceae species, such as *Achillea millefolium*, *Matricaria chamomilla* and *Taraxacum officinale*, were even amongst the top five hosts out of 27 potential hosts tested (Matthies 2017). The average biomass of *Melampyrum* individuals of ca. 500 mg on *Solidago* and 300 mg on *Symphyotrichum* classifies these species amongst the best or moderately good hosts, respectively (in comparison to Matthies (2017)). *Rhinanthus* spp. have been repeatedly reported to grow better when attached to grasses or legumes



Figure 7. Effect of seed-source treatments on host biomass production in infected pots categorised by the individual host–hemiparasite combinations. Boxplots represent median, quartiles and non-outlier ranges, with outliers displayed as points outside the non-outlier ranges. *P*-values indicate significant effects of seed-source treatments. Note the logarithmic scale of the y-axes.

than forbs (Cameron and Seel 2007; Rowntree et al. 2014; Matthies 2021). The biomass production of *Rhinanthus alectorolophus*, converted to values per 1 m², amounted to 136 g DW and 75.6 g DW when attached to *Solidago* and *Symphyotrichum*, respectively. These values are lower than those reported for the best grass hosts in a recent field cultivation experiment (Hejduk et al. 2020). Still, *Solidago* can be considered a similarly good host for *Rhinanthus alectorolophus* as *Lotus corniculatus*, the best host amongst legumes. *Symphyotrichum* is a host of lower quality, but still comparable to some grasses (*Festuca rubra*) or legumes (*Trifolium hybridum*) (Hejduk et al. 2020). Compared to pot cultivations, the two invasive hosts can also be considered of at least moderate quality for *Rhinanthus alectorolophus* with an average biomass of individuals of about 220 mg and 150 mg on *Solidago* and *Symphyotrichum*, respectively. The biomass values per individual may be up to five times higher with the best host species in greenhouse pot cultivations (Těšitel et al. 2015b; Matthies 2021). However, the hemiparasitic plants in these cultivation experiments could benefit from optimal greenhouse conditions, including sufficient soil nutrients and reduced intraspecific competition due to the presence of only a single hemiparasite individual in each pot (Matthies 2021).

Both hemiparasitic species significantly suppressed the growth of both host species, which is the first experimental demonstration of an adverse effect of root hemiparasites on invasive species. We expected that the growth of the hemiparasites would correlate with the level of host suppression (hypothesis 2), which was only partially supported. Both hemiparasitic species reduced Symphyotrichum above-ground biomass by a third despite a significant difference in hemiparasite biomass (Figs 4, 5). Conversely, Solidago growth was reduced by 80% and 50% when parasitised by Melampyrum and Rhinanthus, respectively, corresponding to the difference in biomass production of the two hemiparasitic species and also to maximal levels of host biomass suppression reported from previous pot experiments (70% and 65% by Melampyrum arvense and Rhinanthus alectorolophus, respectively; Těšitel et al. (2015b); Matthies (2017); Sandner and Matthies (2018)). The difference in the host suppression could be related to their clonal growth characteristics, specifically the persistence of ramet connection. The clonal connections of *Solidago* ramets may persist for several years, while connections amongst Symphyotrichum ramets decay after one year (Schmid et al. 1995; Klimešová and Klimeš 2006). Schmid and Bazzaz (1987) suggested stronger physiological integration in Solidago due to the larger effects of experimental rhizome severance on Solidago gigantea growth than Symphyotrichum. The persistent clonal spread was identified as a significant positive predictor of hemiparasite-induced growth reduction (Demey et al. 2015); thus, the putatively stronger integration of Solidago ramets could be one of the reasons for the more extensive damage inflicted by the parasites. Physiological integration may be a trait contributing to a species' susceptibility to plant parasitism.

Examining the interactions between clonal hosts and hemiparasites presents a challenging task. Pot experiments are necessary to isolate the interaction between the host and the generalist hemiparasites from the natural community context, ensuring no other plant serves as a host. Typically, hosts are grown from seeds in these experiments, with hemiparasites later germinating in the pots or being transplanted as pregerminated seedlings. Consequently, hemiparasites attach to young host individuals that have not yet developed clonal growth. Furthermore, arbitrary numbers of host and hemiparasite individuals (sometimes as low as one host with one hemiparasite) are used in most of the pot experiments (e.g. Cameron and Seel (2007); Rowntree et al. (2014); Těšitel et al. (2015b); Matthies (2017); Sandner and Matthies (2018); but see, for example, Matthies (1995) and Hejduk et al. (2020) for hemiparasite densitymanipulation experiments). These issues limit the potential of such experiments to elucidate the clonal host–hemiparasite interaction because, in natural communities, hemiparasite seedlings mostly attach to mature individuals of perennial plants with a fully-developed root system and clonal-growth organs. Thanks to transplanting whole soil blocks from the host population, our experiment maintains the host plant properties (developmental stage, ramet density) as close to natural conditions as possible. In addition, the hemiparasite seedlings were permitted to develop under natural climatic conditions, host phenological development and at densities close to realistic values (van Hulst et al. 1987; Mudrák and Lepš 2010). Hence, our experiment paves the way to more realistic pot experiments studying clonal host–hemiparasite interactions, which are of particular significance in European grassland ecosystem (Demey et al. 2015; Lepš and Těšitel 2015; Těšitel et al. 2017).

Conservation perspective

Pronounced biomass suppression of Solidago and Symphyotrichum is noteworthy from the restoration perspective. Both species are invasive, often achieving dominance and significantly impacting above-ground diversity (Hejda et al. 2021; Cubino et al. 2022; Fenesi et al. 2023). Solidago spp. also affected below-ground soil properties and the activity and biomass of soil bacteria and fungi (Zhang et al. 2009a, b; Scharfy et al. 2010; Pergl et al. 2023). Both species are listed in the second most serious category in the Black List of invasive species (BL2) with a massive environmental impact (Pergl et al. 2016). Hence, reducing their populations is crucial, particularly at sites of high conservation values. Mowing twice, a standard management technique for vegetation infested with Solidago gigantea, reduces the species' dominance. Cover reduction by 75% of the initial cover was reported over the long term, but the species is still persistent in the vegetation (Nagy et al. 2020; Szymura et al. 2022). A more pronounced suppression of Solidago may be achieved through cattle and sheep grazing or flooding (> 95% suppression; Nagy et al. (2020)). Despite the rapid spread of Symphyotrichum lanceolatum in wetland habitats of high natural value (Lanta et al. 2022), no information on managing this invasion is available. Biological control by introducing specialised insects or fungi from the species' native range has not been established yet in the invaded ranges, though several non-native insect enemies may be available in the case of *Solidago* (Fontes et al. 1994; Sheppard et al. 2006). Another biocontrol option available in subtropical regions may represent the widely-spread fungus Sclerotium rolfsii, causing the southern blight disease. Wilting of Solidago canadensis, induced by this fungus, has been reported from China (Tang et al. 2010) and the fungus application combined with soil disturbance led to 90% decrease in Solidago canadensis stem density (Zhang et al. 2019).

Using native hemiparasitic plants in combination with standard mowing management may offer another tool for the biocontrol of the two study species without any potential risks of introducing alien organisms to the ecosystems. The effects of hemiparasites on the invasive hosts observed in our experiment are comparable to the level of the suppression of *Calamagrostis epigejos* by *Rhinanthus alectorolophus* reported in previous research (Těšitel et al. 2017). The reduction and even elimination of this expansive grass by *Rhinanthus* have been established as a standard tool of biodiversity restoration in nature conservation in the Czech Republic (SPPK D02 002 2021). In contrast, *Melampyrum arvense* has not been used in ecological restoration so far, possibly because it is now considered a vulnerable species confined mainly to steppes and protected areas in Central Europe (Těšitel et al. 2015a). However, this species used to be a noxious weed in winter cereal fields (e.g. Rau (1970); Çetinsoy (1980); Matthies (1995)). It can increase its biomass by 1/3 in nutrient-rich soil and prefers hosts from nutrient-rich environments (Matthies 2017). Such ecological characteristics align with the ecology of *Solidago* and *Symphyotrichum*, sometimes called 'old-field perennials' (Schmid and Bazzaz 1987; Schmid et al. 1995), which efficiently colonise bare ground, fallows and disturbed urban areas and thrive in humid, nutrient-rich soils. Our experimental results demonstrate the ability of both hemiparasites to suppress the invasive species, but implementing this finding in ecological restoration requires further testing in the field conditions over longer periods.

Genotype adaptation

We identified transgenerational effects in hemiparasitic interactions thanks to using hemiparasite seeds of the same population origin, but cultivated for two generations (= years) with the target host. The effects were not universal across all host-hemiparasite combinations; however, where present, they generally supported our hypothesis 3. Specifically, when the hemiparasites were exposed to the target host species during two previous generations, the offspring plants produced relatively more biomass (Fig. 6) or were more detrimental to the host (Fig. 7) in some host-hemiparasite combinations. The effects were more pronounced on the hemiparasite side of the association, a pattern identified in a previous study on genotype effects in root-hemiparasitic interactions (Rowntree et al. 2014). Two mechanisms may be at play here: classical genetics and the selection of alleles that provide better compatibility with a host species or epigenetic (maternal) effects acting in the same way (Anastasiadi et al. 2021). We are not able to distinguish between these two with the current data. Even in model organism studies, the state-of-the-art methodology struggles to provide absolute separation of selection and epigenetics (Schmid et al. 2018). However, any adaptive process facilitating the association with novel hosts is crucial for the biotic resistance role of the parasites.

The existence of transgenerational effects in host-hemiparasite compatibility suggests that the breeding of genotypes more compatible with the target invasive hosts or exposing the mother plants to the novel host species may potentially increase the success of biocontrol applications, at least in the case of *Rhinanthus*. The feasibility of such an approach is also supported by the observations of rapid adaptive evolution in *Rhinanthus alectorolophus* in response to environmental conditions and host species (Zopfi 1993; Pleines et al. 2013; Moncalvillo and Matthies 2023). The genetic diversity of hemiparasites was also demonstrated to be a significant predictor of their establishment success and fitness when cultivated with multiple host species (Rowntree and Craig 2019). Therefore, while breeding hemiparasites in monospecific host stands may be efficient for specific purposes, it is equally important to preserve the genetic diversity of the populations of hemiparasitic species in nature and in seed production for ecological restoration; for instance, by cultivating hemiparasites with various host species from different plant functional groups so that the pool of genotypes efficient in various host-hemiparasites combinations is not depleted.

Acknowledgements

The authors thank Pavel Dřevojan, Zuzana Plesková, Zdenka Preislerová, Terezie Chamrátová, Maroš Šlachtič for help with the experiment setting and harvesting and David Watson and Ramiro Bustamante for their insightful comments on the manuscript. This work was supported by the Czech Science Foundation (project 21-22488S).

References

- Anastasiadi D, Venney CJ, Bernatchez L, Wellenreuther M (2021) Epigenetic inheritance and reproductive mode in plants and animals. Trends in Ecology & Evolution 36(12): 1124– 1140. https://doi.org/10.1016/j.tree.2021.08.006
- Axmanová I, Kalusová V, Danihelka J, Dengler J, Pergl J, Pyšek P, Večeřa M, Attorre F, Biurrun I, Boch S, Conradi T, Gavilán RG, Jiménez-Alfaro B, Knollová I, Kuzemko A, Lenoir J, Leostrin A, Medvecká J, Moeslund JE, Obratov-Petkovic D, Svenning J-C, Tsiripidis I, Vassilev K, Chytrý M (2021) Neophyte invasions in European grasslands. Journal of Vegetation Science 32(2): e12994. https://doi.org/10.1111/jvs.12994
- Blackburn TM, Essl F, Evans T, Hulme PE, Jeschke JM, Kühn I, Kumschick S, Marková Z, Mrugała A, Nentwig W, Pergl J, Pyšek P, Rabitsch W, Ricciardi A, Richardson DM, Sendek A, Vilà M, Wilson JRU, Winter M, Genovesi P, Bacher S (2014) A unified classification of alien species based on the magnitude of their environmental impacts. PLoS Biology 12(5): e1001850. https://doi.org/10.1371/journal.pbio.1001850
- Cameron DD, Seel WE (2007) Functional anatomy of haustoria formed by *Rhinanthus minor*: Linking evidence from histology and isotope tracing. The New Phytologist 174(2): 412– 419. https://doi.org/10.1111/j.1469-8137.2007.02013.x
- Çetinsoy S (1980) Studies on the Determination of Effective Chemicals Against *Melampyrum arvense* L., Harmful in Cereals Fields in Central Anatolia. Bölge Zirai Mücadele Arastima Enstitüsu, Ankara.
- Chmielewski JG, Semple JC (2001) The biology of Canadian weeds. 113. Symphyotrichum lanceolatum (Willd.) Nesom [Aster lanceolatus Willd.] and S. lateriflorum (L.) Löve & Löve. Canadian Journal of Plant Science 81(4): 829–849. https://doi.org/10.4141/P00-056 [Aster lateriflorus (L.) Britt.]
- Cubino JP, Těšitel J, Fibich P, Lepš J, Chytrý M (2022) Alien plants tend to occur in speciespoor communities. NeoBiota 73: 39–56. https://doi.org/10.3897/neobiota.73.79696
- Decleer K, Bonte D, Van Diggelen R (2013) The hemiparasite *Pedicularis palustris*: 'Ecosystem engineer' for fen-meadow restoration. Journal for Nature Conservation 21(2): 65–71. https://doi.org/10.1016/j.jnc.2012.10.004

- Demey A, De Frenne P, Baeten L, Verstraeten G, Hermy M, Boeckx P, Verheyen K (2015) The effects of hemiparasitic plant removal on community structure and seedling establishment in semi-natural grasslands. Journal of Vegetation Science 26(3): 409–420. https:// doi.org/10.1111/jvs.12262
- Divíšek J, Chytrý M, Beckage B, Gotelli NJ, Lososová Z, Pyšek P, Richardson DM, Molofsky J (2018) Similarity of introduced plant species to native ones facilitates naturalization, but differences enhance invasion success. Nature Communications 9(1): e4631. https://doi. org/10.1038/s41467-018-06995-4
- Fenesi A, Botta-Dukát Z, Miholcsa Z, Szigeti V, Molnár C, Sándor D, Szabó A, Kuhn T, Kovács-Hostyánszki A (2023) No consistencies in abundance–impact relationships across herbaceous invasive species and ecological impact metrics. Journal of Ecology 111(5): 1120–1138. https://doi.org/10.1111/1365-2745.14085
- Fontes EMG, Habeck DH, Slansky F (1994) Phytophagous insects associated with goldenrods (*Solidago* spp.) in Gainesville, Florida. The Florida Entomologist 77(2): 209–209. https:// doi.org/10.2307/3495506
- Gao F-L, Alpert P, Yu F (2021) Parasitism induces negative effects of physiological integration in a clonal plant. The New Phytologist 229(1): 585–592. https://doi.org/10.1111/ nph.16884
- Heer N, Klimmek F, Zwahlen C, Fischer M, Hölzel N, Klaus VH, Kleinebecker T, Prati D, Boch S (2018) Hemiparasite-density effects on grassland plant diversity, composition and biomass. Perspectives in Plant Ecology, Evolution and Systematics 32: 22–29. https://doi. org/10.1016/j.ppees.2018.01.004
- Hejda M, Sádlo J, Kutlvašr J, Petřík P, Vítková M, Vojík M, Pyšek P, Pergl J (2021) Impact of invasive and native dominants on species richness and diversity of plant communities. Preslia 93(3): 181–201. https://doi.org/10.23855/preslia.2021.181
- Hejduk S, Bitomský M, Pornaro C, Macolino S (2020) Establishment of a hemiparasite *Rhi-nanthus alectorolophus* and its density-dependent suppressing effect on a grass: A case study from golf roughs. Agronomy Journal 112(5): 3619–3628. https://doi.org/10.1002/agj2.20300
- Jedlička J, Prach K (2006) A comparison of two North-American asters invading in central Europe. Flora 201: 652–657. https://doi.org/10.1016/j.flora.2006.01.002
- Kavanová M, Gloser V (2005) The use of internal nitrogen stores in the rhizomatous grass Calamagrostis epigejos during regrowth after defoliation. Annals of Botany 95: 457–463. https://doi.org/10.1093/aob/mci054
- Klimešová J, Klimeš L (2006) Clo-Pla3–database of clonal growth of plants from Central Europe. https://clopla.butbn.cas.cz
- Lanta V, Liancourt P, Altman J, Černý T, Dvorský M, Fibich P, Götzenberger L, Hornych O, Miklín J, Petřík P, Pyšek P, Čížek L, Doležal J (2022) Determinants of invasion by single versus multiple plant species in temperate lowland forests. Biological Invasions 24(8): 2513–2528. https://doi.org/10.1007/s10530-022-02793-8
- Lepš J, Těšitel J (2015) Root hemiparasites in productive communities should attack competitive host, and harm them to make regeneration gaps. Journal of Vegetation Science 26(3): 407–408. https://doi.org/10.1111/jvs.12284

- Liljegren S (2010) Phloroglucinol stain for lignin. Cold Spring Harbor Protocols 2010.1: pdb. prot4954. https://doi.org/10.1101/pdb.prot4954
- Lukavský J (2020) Funguje nám kokrhel? Aneb zkušenosti z regionu. Ochrana přírody 2020: 11– 15. https://www.casopis.ochranaprirody.cz/pece-o-prirodu-a-krajinu/funguje-nam-kokrhel/
- Maron JL, Vilà M (2001) When do herbivores affect plant invasion? Evidence for the natural enemies and biotic resistance hypotheses. Oikos 95(3): 361–373. https://doi.org/10.1034/j.1600-0706.2001.950301.x
- Matthies D (1995) Host-parasite relations in the root hemiparasite *Melampyrum arvense*. Flora 190(4): 383–394. https://doi.org/10.1016/S0367-2530(17)30680-1
- Matthies D (2017) Interactions between a root hemiparasite and 27 different hosts: Growth, biomass allocation and plant architecture. Perspectives in Plant Ecology, Evolution and Systematics 24: 118–137. https://doi.org/10.1016/j.ppees.2016.12.006
- Matthies D (2021) Closely related parasitic plants have similar host requirements and related effects on hosts. Ecology and Evolution 11(17): 12011–12024. https://doi.org/10.1002/ece3.7967
- Moncalvillo B, Matthies D (2023) Performance of a parasitic plant and its effects on hosts depends on the interactions between parasite seed family and host species. AoB Plants 15(2): 1–11. https://doi.org/10.1093/aobpla/plac063
- Mudrák O, Lepš J (2010) Interactions of the hemiparasitic species *Rhinanthus minor* with its host plant community at two nutrient levels. Folia Geobotanica 45(4): 407–424. https:// doi.org/10.1007/s12224-010-9078-1
- Mutikainen P, Salonen V, Pustinen S, Koskela T (2000) Local adaptation, resistance, and virulence in a hemiparasitic plant-host plant interaction. Evolution; International Journal of Organic Evolution 54(2): 433–440. https://doi.org/10.1111/j.0014-3820.2000.tb00046.x
- Nagy DU, Rauschert ESJ, Henn T, Cianfaglione K, Stranczinger S, Pal RW (2020) The more we do, the less we gain? Balancing effort and efficacy in managing the *Solidago gigantea* invasion. Weed Research 60(3): 232–240. https://doi.org/10.1111/wre.12417
- Pergl J, Sádlo J, Petrusek A, Laštuvka Z, Musil J, Perglová I, Šanda R, Šefrová H, Šíma J, Vohralík V, Pyšek P (2016) Black, Grey and Watch Lists of alien species in the Czech Republic based on environmental impacts and management strategy. NeoBiota 28: 1–37. https://doi.org/10.3897/neobiota.28.4824
- Pergl J, Vítková M, Hejda M, Kutlvašr J, Petřík P, Sádlo J, Vojík M, Dvořáčková Š, Fleischhans R, Lučanová A, Pyšek P (2023) Plant-soil interactions in the communities dominated by alien and native plants. Perspectives in Plant Ecology, Evolution and Systematics 59: e125721. https://doi.org/10.1016/j.ppees.2023.125721
- Pleines T, Esfeld K, Blattner FR, Thiv M (2013) Ecotypes and genetic structure of *Rhinanthus alectorolophus* (Orobanchaceae) in southwestern Germany. Plant Systematics and Evolution 299(8): 1523–1535. https://doi.org/10.1007/s00606-013-0816-8
- Press MC, Press MC, Phoenix GK (2005) Impacts of parasitic plants on natural communities. The New Phytologist 166(3): 737–751. https://doi.org/10.1111/j.1469-8137.2005.01358.x
- Pyšek P, Chytrý M, Pergl J, Sádlo J, Wild J (2012) Plant invasions in the Czech Republic: Current state, introduction dynamics, invasive species and invaded habitats. Preslia 84: 575–629. https://www.preslia.cz/article/157

- R Core Team (2022) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna. http://www.R-project.org
- Rau E (1970) Changes in the species spectrum of the noxious plant flora. Gesunde Pflanzen 22: 164–165.
- Richardson DM, Pyšek P, Rejmánek M, Barbour MG, Dane Panetta F, West CJ (2000) Naturalization and invasion of alien plants: Concepts and definitions. Diversity & Distributions 6(2): 93–107. https://doi.org/10.1046/j.1472-4642.2000.00083.x
- Roiloa SR (2019) Clonal traits and plant invasiveness: The case of *Carpobrotus* N. E. Br. (Aizoaceae). Perspectives in Plant Ecology, Evolution and Systematics 40: e125479. https:// doi.org/10.1016/j.ppees.2019.125479
- Rowntree JK, Craig H (2019) The contrasting roles of host species diversity and parasite population genetic diversity in the infection dynamics of a keystone parasitic plant. Journal of Ecology 107(1): 23–33. https://doi.org/10.1111/1365-2745.13050
- Rowntree JK, Cameron DD, Preziosi RF (2011) Genetic variation changes the interactions between the parasitic plant-ecosystem engineer *Rhinanthus* and its hosts. Philosophical Transactions of the Royal Society of London, Series B, Biological Sciences 366(1569): 1380–1388. https://doi.org/10.1098/rstb.2010.0320
- Rowntree JK, Fisher Barham D, Stewart AJA, Hartley SE (2014) The effect of multiple host species on a keystone parasitic plant and its aphid herbivores. Functional Ecology 28(4): 829–836. https://doi.org/10.1111/1365-2435.12281
- Sandner TM, Matthies D (2018) Multiple choice: Hemiparasite performance in multi-species mixtures. Oikos 127(9): 1291–1303. https://doi.org/10.1111/oik.05148
- Scharfy D, Güsewell S, Gessner MO, Venterink HO (2010) Invasion of Solidago gigantea in contrasting experimental plant communities: Effects on soil microbes, nutrients and plant – soil feedbacks. Journal of Ecology 98(6): 1379–1388. https://doi.org/10.1111/j.1365-2745.2010.01722.x
- Schmid B, Bazzaz FA (1987) Clonal integration and population structure in perennials: Effects of severing rhizome connections. Ecology 68(6): 2016–2022. https://doi. org/10.2307/1939892
- Schmid B, Bazzaz FA, Weiner J (1995) Size dependency of sexual reproduction and of clonal growth in two perennial plants. Canadian Journal of Botany 73(11): 1831–1837. https:// doi.org/10.1139/b95-194
- Schmid MW, Heichinger C, Schmid DC, Guthörl D, Gagliardini V, Bruggmann R, Aluri S, Aquino C, Schmid B, Turnbull LA, Grossniklaus U (2018) Contribution of epigenetic variation to adaptation in Arabidopsis. Nature Communications 9(1): e4446. https://doi. org/10.1038/s41467-018-06932-5
- Sheppard AW, Shaw RH, Sforza R (2006) Top 20 environmental weeds for classical biological control in Europe: A review of opportunities, regulations and other barriers to adoption. Weed Research 46(2): 93–117. https://doi.org/10.1111/j.1365-3180.2006.00497.x
- Somodi I, Vadkerti Á, Těšitel J (2018) *Thesium linophyllon* parasitizes and suppresses expansive *Calamagrostis epigejos*. Plant Biology 20(4): 759–764. https://doi.org/10.1111/plb.12723
- Song Y, Yu F, Keser LH, Dawson W, Fischer M, Dong M, van Kleunen M (2013) United we stand, divided we fall: A meta-analysis of experiments on clonal integration and its re-

lationship to invasiveness. Oecologia 171(2): 317–327. https://doi.org/10.1007/s00442-012-2430-9

- Soukup A, Tylová E (2014) Essential methods of plant sample preparation for light microscopy. Methods in Molecular Biology 1080: 1–23. https://doi.org/10.1007/978-1-62703-643-6_1
- SPPK D02 002 (2021) Standardy Péče o Přírodu a Krajinu: Obnova Dlouhodobě Neobhospodařovaných Travních Společenstev (vč. likvidace náletových dřevin). AOPK ČR, Prague, 39 pp. https://nature.cz/documents/20121/1200108/02002_OBNOVA_ DLOUHODOBE_NEOBHOSPODAROVANYCH_TS.pdf
- Szymura M, Świerszcz S, Szymura TH (2022) Restoration of ecologically valuable grassland on sites degraded by invasive *Solidago*: Lessons from a 6-year experiment. Land Degradation & Development 33(12): 1985–1998. https://doi.org/10.1002/ldr.4278
- Tang W, Zhu YZ, He HQ, Qiang S (2010) First report of Southern blight on Canadian goldenrod (*Solidago canadensis*) caused by *Sclerotium rolfsii* in China. Plant Disease 94(9): 1172–1172. https://doi.org/10.1094/PDIS-94-9-1172B
- Teixeira-Costa L, Davis CC (2021) Life history, diversity, and distribution in parasitic flowering plants. Plant Physiology 187(1): 32–51. https://doi.org/10.1093/plphys/kiab279
- Těšitel J, Fibich P, De Bello F, Chytrý M, Lepš J (2015a) Habitats and ecological niches of roothemiparasitic plants: An assessment based on a large database of vegetation plots. Preslia 87: 87–108. https://www.preslia.cz/article/95
- Těšitel J, Těšitelová T, Fisher JP, Lepš J, Cameron DD (2015b) Integrating ecology and physiology of root-hemiparasitic interaction: Interactive effects of abiotic resources shape the interplay between parasitism and autotrophy. The New Phytologist 205(1): 350–360. https://doi.org/10.1111/nph.13006
- Těšitel J, Mládek J, Horník J, Těšitelová T, Adamec V, Tichý L (2017) Suppressing competitive dominants and community restoration with native parasitic plants using the hemiparasitic *Rhinanthus alectorolophus* and the dominant grass *Calamagrostis epigejos*. Journal of Applied Ecology 54(5): 1487–1495. https://doi.org/10.1111/1365-2664.12889
- Těšitel J, Mládek J, Fajmon K, Blažek P, Mudrák O (2018) Reversing expansion of *Calamagros*tis epigejos in a grassland biodiversity hotspot: Hemiparasitic *Rhinanthus major* does a better job than increased mowing intensity. Applied Vegetation Science 21(1): 104–112. https:// doi.org/10.1111/avsc.12339
- Těšitel J, Cirocco RM, Facelli JM, Watling JR (2020) Native parasitic plants: Biological control for plant invasions? Applied Vegetation Science 23(3): 464–469. https://doi.org/10.1111/ avsc.12498
- Unachukwu NN, Menkir A, Rabbi IY, Oluoch M, Muranaka A, Elzein A, Odhiambo G, Farombi EO, Gedil M (2017) Genetic diversity and population structure of *Striga hermonthica* populations from Kenya and Nigeria. Weed Research 57(5): 293–302. https://doi. org/10.1111/wre.12260
- van Hulst R, Shipley B, Thériault A (1987) Why is *Rhinanthus minor* (Scrophulariaceae) such a good invader? Canadian Journal of Botany 65(11): 2373–2379. https://doi.org/10.1139/b87-322
- Walder M, Armstrong JE, Borowicz VA (2019) Limiting similarity, biotic resistance, nutrient supply, or enemies? What accounts for the invasion success of an exotic legume? Biological Invasions 21(2): 435–449. https://doi.org/10.1007/s10530-018-1835-8

- Wan J-Z, Wang C-J, Zimmermann NE, Li M-H, Pouteau R, Yu F-H (2021) Current and future plant invasions in protected areas: Does clonality matter? Diversity & Distributions 27(12): 2465–2478. https://doi.org/10.1111/ddi.13425
- Wang Y, Chen D, Yan R, Yu F, van Kleunen M (2019) Invasive alien clonal plants are competitively superior over co-occurring native clonal plants. Perspectives in Plant Ecology, Evolution and Systematics 40: 125484. https://doi.org/10.1016/j.ppees.2019.125484
- Weber E, Jakobs G (2005) Biological flora of central Europe: Solidago gigantea Aiton. Flora 200(2): 109–118. https://doi.org/10.1016/j.flora.2004.09.001
- Zhang S, Jin Y, Tang J, Chen X (2009a) The invasive plant Solidago canadensis L. suppresses local soil pathogens through allelopathy. Applied Soil Ecology 41(2): 215–222. https://doi. org/10.1016/j.apsoil.2008.11.002
- Zhang CB, Wang J, Qian BY, Li WH (2009b) Effects of the invader Solidago canadensis on soil properties. Applied Soil Ecology 43(2–3): 163–169. https://doi.org/10.1016/j. apsoil.2009.07.001
- Zhang Y, Yang X, Zhu Y, Li L, Zhang Y, Li J, Song X, Qiang S (2019) Biological control of *Solidago* canadensis using a bioherbicide isolate of *Sclerotium rolfsii* SC64 increased the biodiversity in invaded habitats. Biological Control 139: e104093. https://doi.org/10.1016/j. biocontrol.2019.104093
- Zopfi H (1993) Ecotypic variation in *Rhinanthus alectorolophus* (Scopoli) Pollich (Scrophulariaceae) in relation to grassland management II. The genotypic basis of seasonal ecotypes. Flora 188: 153–173. https://doi.org/10.1016/S0367-2530(17)32261-2

Supplementary material I

Supplementary information

Authors: Tamara Těšitelová, Kateřina Knotková, Adam Knotek, Hana Cempírková, Jakub Těšitel

Data type: docx

Explanation note: appendix S1: Location of source localities of hemiparasite seeds and invasive host plants. appendix S2: Overview of the number of hemiparasite specimens and their biomass in the experimental treatments. appendix S3: Overview of host shoot counts and host biomass in the experimental treatments. appendix S4: Photographic documentation of all pots representing the experimental treatments.

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.113069.suppl1

Supplementary material 2

Primary data table

Authors: Tamara Těšitelová, Kateřina Knotková, Adam Knotek, Hana Cempírková, Jakub Těšitel

Data type: xlsx

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.113069.suppl2



Distribution and genetic diversity of the invasive pest Halyomorpha halys (Hemiptera, Pentatomidae) in Belgium

Olivier Hendrik Berteloot^{1*}, Alexandre Kuhn^{2*}, Gertie Peusens³, Tim Beliën³, Louis Hautier², Thomas Van Leeuwen¹, Patrick De Clercq¹

 Department of Plants and Crops, Faculty of Bioscience Engineering, Ghent University (UGent), Ghent, Belgium 2 Crops and Forest Health Unit, Life Sciences Department, Walloon Agricultural Research Centre (CRA-W), Gembloux, Belgium 3 Zoology Department, Research Centre for Fruit Cultivation (PCFruit), Sint-Truiden, Belgium

Corresponding author: Olivier Hendrik Berteloot (olivier.berteloot@ugent.be; olivier.berteloot@gmail.com)

Academic editor: A. Roques | Received 28 September 2023 | Accepted 5 December 2023 | Published 18 January 2024

Citation: Berteloot OH, Kuhn A, Peusens G, Beliën T, Hautier L, Van Leeuwen T, De Clercq P (2024) Distribution and genetic diversity of the invasive pest *Halyomorpha halys* (Hemiptera, Pentatomidae) in Belgium. NeoBiota 90: 123–138. https://doi.org/10.3897/neobiota.90.113421

Abstract

The brown marmorated stink bug, *Halyomorpha halys*, native to East Asia, is an invasive pest of economic importance. It has invaded North America and many European countries and is further expanding its range. In Belgium, it was first observed in 2011. *Halyomorpha halys* is known to cause severe damage in commercial fruit orchards and vegetable crops. A dramatic and unmitigated expansion of *H. halys* in its adventive range could lead to significant economic implications for agricultural production. In this study, occurrence data of *H. halys* since its first observation in Belgium was analysed together with molecular information to map the populations and evaluate the genetic diversity of this pest. The genetic diversity of *H. halys* in Belgium was compared to data from other invaded and native countries reported in previous studies to identify possible invasion routes. The analysis of 1176-bp of mitochondrial DNA cytochrome c oxidase I and II genes (COI and COII) led to the discovery of two novel COI-COII haplotype combinations currently unique to Belgium. The invasion of *H. halys* in Belgium is likely the result of multiple and ongoing introductions from its native region and from already invaded countries in Europe, particularly Italy. The expansion of the brown marmorated stink bug populations in Belgium is recent and ongoing. Presently, it appears to thrive best in northern Belgium.

^{*} These authors contributed equally to this work.

Keywords

brown marmorated stink bug, haplotype diversity, Heteroptera, population genetics

Introduction

Halyomorpha halys (Hemiptera: Pentatomidae) or the brown marmorated stink bug is a stink bug of economic importance, native to East Asia and an invasive pest species in Europe, North America, and other regions (Hoebeke and Carter 2003; Leskey et al. 2012; Lee et al. 2013; Haye et al. 2015; Zhu et al. 2016; Hamilton et al. 2018). In Europe, H. halys has already been detected as far north as The Netherlands (Aukema et al. 2019), Belgium (Claerebout et al. 2018) and the north of Germany (Hartung et al. 2022). Due to its polyphagous nature, *H. halys* can feed on virtually all primary fruit and vegetable crops in its invaded areas (Kuhar et al. 2012; Haye et al. 2015). For example, in northern Italy, it has become a key pest in fruit orchards, with an estimated economic impact of €588 million on the production of pear, apple, peach, and kiwi in 2019 (Bulgarini et al. 2020). Halyomorpha halys causes damage through its feeding activity. It inserts its stylets into the plant and injects saliva, pre-digesting the plant tissue before feeding on it (Rice et al. 2014). This feeding behaviour causes scarring, deformities, pitting, and discolouration of the product, rendering it unmarketable or even inedible (Rice et al. 2014; Bulgarini et al. 2020). Since its first record in Belgium in 2011, observations of H. halys on public citizen science databases (Observation International and local partners 2022) only appeared in 2017. Currently, it is assumed that H. halys has established univoltine overwintering breeding populations in Belgium (Claerebout et al. 2018). An observation of *H. halys* from the Haspengouw region in July 2021 indicated that *H. halys* is already present in commercial pip fruit orchards in Belgium. However, to date, no dramatic population increases, mass occurrences or damage has been reported in fruit production in Belgium (Berteloot et al. 2021). In 2022, in Belgium, 219.000 t of apples and 366.000 t of pears, some of the most vulnerable fruits to H. halys, were produced (European Commission 2022a, b). Apples and pears represent a combined revenue of more than € 148 million of which pear production is the most significant part with a revenue of € 119 million (Verbond van Belgische Tuinbouwcoöperaties 2022). An unmitigated expansion of H. halys in Belgium could therefore have a considerable impact on fruit production, especially in the Flanders region, which produces most of the apples, pears, and soft fruits in the country.

The increased accessibility and affordability of molecular technologies, as well as the expansion of databases containing publicly available DNA sequence data, have aided in the use of molecular tools to assess the genetic diversity and potential origin of invasive species (Cristescu 2016; Hamelin and Roe 2019). Due to its lack of recombination, relative neutrality and shorter coalescence time, mitochondrial DNA (mtDNA) is a highly popular molecular marker for examining genetic diversity and phylogeography of (invasive) animal species (Ficetola et al. 2008; Rollins et al. 2011; Bras et al. 2019; Ryan et al. 2019). Furthermore, for invasive species, the accumulation of easily reproducible genetic data in public databases facilitates the regular addition of new data from the invasion front to build a global pattern of invasion dynamics. For example, for *H. halys*, mtDNA sequences (portions of the mitochondrial gene cytochrome c oxidase subunits I and II – COI and COII, respectively) are already available for both native (China, Japan, Korea) and invaded countries (Gariepy et al. 2014; Xu et al. 2014; Cesari et al. 2015; Cesari et al. 2018; Lee et al. 2018; Yan et al. 2021).

In this study, we set out to fill one of the geographical knowledge gaps in Europe on the genetic diversity and distribution of *H. halys*. First, we investigated the genetic diversity of *H. halys* in Belgium by sequencing 99 specimens from 18 locations at two mitochondrial genes: COI and COII. Then, we compared those sequences to previously published *H. halys* sequences to infer the putative invasion routes to Belgium. Lastly, we used public citizen occurrence data to map the distribution of *H. halys* in Belgium from 2020 to 2022 and gain demographic insights into the Belgian populations.

Methods

Insect sampling

Halyomorpha halys specimens were collected from 18 locations in Belgium (Suppl. material 1) from agricultural sites and private gardens in 2021 and 2022 using baited traps (dual lure from Trécé Inc., Adair Oklahoma, USA) or by beating of the lower part of a tree trunk/shrub. The collected samples were stored dry or in 70% ethanol at -20 °C until used for molecular analysis.

DNA extraction & sequencing

DNA was extracted from two legs using a Chelex extraction method (Walsh et al. 1991). Briefly, the legs were crushed with a sterile pestle in 100 µl of 5% Chelex 100 (Bio-Rad, USA) solution. They were then incubated at 85 °C for 90 min, and the supernatant was collected after 3 min of centrifugation at 12,000 rpm. The cytochrome oxidase subunit I (COI) and II (COII) barcode regions of the mitochondrial DNA were amplified through PCR. For the COI region, the LCO 1490 (5'- GCT-CAACAAATCATAAAGATATTGG-3') and HCO 2198 (5'- TAAACTTCAGGGT-GACCAAAAAAATCA-3') primers (Folmer et al. 1994) were used. For COII, the HhalysCO2F2 (5'-TAACCCAAGATGCAAAATTCT-3') and HhalysCO2R2 (5'- CCATA-TATAATTCCTGGACGA-3') primers (Xu et al. 2014) were used. For both regions, the following PCR cycles were used: initial denaturation at 94 °C for 3 min, 38 cycles of denaturation at 94 °C for 30 sec, annealing at 48 °C for 7 min. The PCR product quality was checked on agarose gel. Both strands (forward and reverse) for each barcode region were sequenced (Eurofins, Germany GmbH) by Sanger sequencing.

Haplotype diversity analysis

Forward and reverse sequences were trimmed and assembled into a consensus sequence using CodonCode Aligner (version 10.0.2). COI and COII sequences obtained in this study were compared to COI and COII sequences from Yan et al. (2021) and Cesari et al. (2018), retrieved from GenBank and BOLD. Other previously conducted studies on the genetic diversity of H. halys only sequenced COI or COII fragments (or only reported unique haplotypes found without specifying which samples are associated with which haplotypes) (Gariepy et al. 2014; Xu et al. 2014; Valentin et al. 2017; Lee et al. 2018; Kapantaidaki et al. 2019). All individual COI and COII haplotypes found in this study have already been reported in other publications (see results) and were given the haplotype name used in those publications (Cesari et al. 2018; Yan et al. 2021). However, for COI, there were discrepancies between the sequences of haplotypes with the same name found in Cesari et al. (2018) and Yan et al. (2021). Haplotypes found in our study were named after the reference haplotypes of the study of Valentin et al. (2017) if a sequence match was found. Haplotype names from Cesari et al. (2018) were distinguished with the letter "c" (Table 1). The sequence of haplotype H41 from Yan et al. (2021) perfectly matched the H41 haplotype from Valentin et al. (2017) but corresponded to the sequence of H46 from Cesari et al. (2018). Therefore, the name of the sequence of this haplotype was retained in the present study as H41. Another sequence found in this study without a match in Valentin et al. (2017) or Yan et al. (2021) was already named H41 in Cesari et al. (2018) and was renamed "H41c" to distinguish it. For H40 and H43, the same haplotype names were given to different sequences by Cesari et al. (2018) and Yan et al. (2021). Again, those sequences were compared to the ones of Valentin et al. (2017). The H40 and H43 sequences from Yan et al. (2021) perfectly matched the identical name sequences from Valentin et al. (2017), and these names were retained in the present study. A letter "c" was added to the H40 and H43 sequences from Cesari et al. (2018) to distinguish them. Finally, the H42 and H49 sequences from Cesari et al. (2018) did not match the H42 and H49 haplotypes from Valentin et al. (2017) (no H42 and H49 haplotypes in Yan et al. (2021) for comparison). To be consistent, the letter "c" was added to the H42 and H49 sequences from Cesari et al. (2018).

Table 1. Summary of the discrepancies between sequences of COI-haplotypes with the same name in different studies and the names used in the present study.

Valentin et al. (2017)	Cesari et al. (2018)	Yan et al. (2021)	This study
H40	/	H40	H40
/	H40	/	H40c
H41	H46	H41	H41
/	H41	/	H41c
/	H42	/	H42c
H43	/	H43	H43
/	H43	/	H43c
/	H49	/	H49c

Sequences were aligned and analysed in R v4.0.2 (R Core Team 2020), using packages msa (Bodenhofer et al. 2015), adegenet (Jombart 2008) and pegas (Paradis 2010). The relationships among haplotypes within Belgium and between haplotypes from Belgium and other countries were investigated through haplotype networks and genetic diversity indexes (haplotype, Hd, and nucleotide diversity, π). We tested for the presence of a genetic structure in Belgium by performing a Mantel test (Euclidean) on genetic and geographic distance matrices between collected specimens.

Occurrence data

Occurrences from January 1st, 2017, to December 31st, 2022 were obtained from the publicly available citizen science database (Observation International and local partners 2022). For *H. halys*, the recorded occurrences on the public citizen science database are quality-controlled, and the taxonomic accuracy of each observation is confirmed or rejected by a specialist through photos. The occurrence data was additionally examined to check the validity of all the occurrences. Only occurrences with a high confidence level in the taxonomic identification of *H. halys* and with spatial coordinates were used. Additionally, specimens collected during this research were identified through morphological identification using the *H. halys* identification key from Maistrello et al. (2016) and confirmed by our molecular analysis. The occurrence data was visualised with QGIS 3.30.1 (QGIS Development Team 2023).

Results

DNA sequences

• COI: 99 *H. halys* sequences were retrieved out of 99 specimens collected. All specimens yielded a 658-bp DNA sequence. The DNA sequence data and specimen collection information were made accessible on GenBank with accession numbers OR581617–OR581715.

• COII: 93 sequences with a length of 518-bp were retrieved from 99 *H. halys* specimens collected. The DNA sequence data and specimen collection information were made accessible on GenBank with accession numbers OR602454–OR602546.

Haplotype diversity of H. halys

For the COI fragment individually, nine distinct haplotypes were found, consisting of 14 polymorphic sites (Fig. 1a). Three haplotypes were dominant (H01, H03 and H08) and were shared by roughly 80% of the individuals collected. Haplotype diversity was 0.77 ± 0.02 (mean \pm SD). The obtained COI sequences from this study were compared with 609 COI sequences from other European countries, Chile, the USA, China, and Japan. H01 and H03 haplotypes are shared with those in invaded areas in



Figure 1. Haplotype network using **A** COI **B** COII and **C** COI-COII fragments of *H. halys* in Belgium. The circle size is proportional to the square root of the frequency of the haplotypes. The tick marks on each line represent a base pair difference.

Europe and America, as well as with those in the native area of China, but not with Japan. Haplotype H08 is shared only with some invaded areas in Europe (Austria and Italy). The remaining haplotypes are shared with Italy (H40c, H41, H41c, H42c and H49c), Japan (H23 and H41) and Hungary (H41).

For the COII fragment individually, five distinct haplotypes were found, consisting of 5 polymorphic sites (Fig. 1b). Haplotype diversity was 0.70 ± 0.01 . Our COII sequences were compared with 651 COII sequences from other European countries, Chile, the USA, China, and Japan. Haplotypes h01, h03 and h11 accounted for 91 out of 95 haplotypes from Belgium. Haplotypes h01 and h03 are shared with invaded areas in Europe, the USA and China but not with Japan. Haplotypes h11 and h15 are only shared with some other European countries (Austria, Hungary, and Italy) and Japan. Haplotype h21 is shared only with Italy.

The resulting concatenated 1176-bp sequences rendered 12 distinct COI-COII haplotypes among 95 specimens consisting of 22 polymorphic sites in total (Fig.1c). Two novel COI-COII haplotypes were identified (H03h11 and H08h21) in our samples; these haplotypes are unique to Belgium. The ten other haplotype combinations were previously reported in the literature. The comparison of COI-COII haplotypes from Belgium to previously published sequences confirm the pattern observed for individual loci. The most frequent haplotype H01h01 is shared with all native and invaded countries included in this study (Table 2). H03h03, the second most frequent haplotype, has been recorded in Austria, Chile, China, Italy, and Hungary. Some of the remaining haplotypes (H03h11, H03h21, H41ch03 and H49ch03) are only shared with Italy (and Austria for H08h11) and are close to haplotypes from China. Others (H23h11, H41h15 and H41ch11) are shared with Italy (and Hungary for H41h15) and/or with (or close to) Japan (H46h15, H42h11, H23h11 and H08h11). Finally, the H08h21 and the H03h11 haplotypes are unique to Belgium and are close to the haplotypes from Italy (within the group of haplotypes found in most invaded countries and China).

Continent Country	Elect and al	N II.	III . CD	5D	Sec. 1
equences are listed).					
f haplotypes, h: haplotype c	liversity and π : nucleotic	de diversity (only counti	ies with availa	ble COI-COII
able 2. Summary table of a	ntDNA (COI-COII) di	iversity by co	untry. With	n N: sample siz	æ, Hn: number

Continent	Country	First record	Ν	Hn	Hd ± SD	$\pi \pm SD$	Study
Asia	China	Native	90	24	0.86 ± 0.02	0.0033 ± 0.0018	Yan et al. 2021
	Japan	Native	65	32	0.94 ± 0.01	0.0024 ± 0.0014	Yan et al. 2021
	Turkey	2017 (Güncan and Gümüş 2019)	11	1	0	0	Yan et al. 2021
Europe	Austria	2015 (Rabitsch and Friebe 2015)	15	4	0.69 ± 0.10	0.0021 ± 0.0013	Yan et al. 2021
	Belgium	2011 (Claerebout et al. 2018)	95	12	0.79 ± 0.02	0.0031 ± 0.0018	This study
	Georgia	2015 (Gapon 2016)	31	1	0	0	Yan et al. 2021
	Greece	2011 (Milonas and Partsinevelos 2014)	8	3	0.61 ± 0.16	0.0025 ± 0.0017	Cesari et al. 2018
	Hungary	2014 (Vétek et al. 2014)	90	3	0.11 ± 0.04	0.0003 ± 0.0003	Yan et al. 2021
	Italy	2012 (Maistrello et al. 2014)	16	18	0.72 ± 0.03	0.0028 ± 0.0016	Cesari et al. 2018;
							Yan et al. 2021
	Romania	2015 (Macavei et al. 2015)	30	1	0	0	Cesari et al. 2018
	Serbia	2015 (Šeat 2015)	13	5	0.61 ± 0.07	0.0014 ± 0.0008	Yan et al. 2021
	Slovenia	2017 (Rot et al. 2018)	15	3	0.51 ± 0.12	0.0012 ± 0.0008	Yan et al. 2021
North America	United States	2001 (Hoebeke and Carter 2003)	24	1	0	0	Yan et al. 2021
South America	Chile	2017 (Faúndez and Rider 2017)	31	2	0.06 ± 0.06	0.0001 ± 0.0002	Yan et al. 2021

The results of our Mantel test indicated a significant but weak correlation (r = 0.14, p = 0.001) between the genetic distance and the geographical distances for pairs of individuals. High haplotype diversity was observed in Belgium, Hd = 0.79 ± 0.02, with a nucleotide diversity value of $\pi = 0.0031 \pm 0.0018$ (Table 2). This level of genetic diversity was lower than that of native regions Japan and China (Hd = 0.94 ± 0.014 and 0.86 ± 0.023, respectively) but similar to Italy (Hd = 0.72 ± 0.033) (Table 2). Other invaded areas of *H. halys* typically had lower haplotype diversity but their estimation may be approximate due to shallower sampling depth (low sample size and/or few sampling locations; Table 2). The genetic data analysis of this research is publicly available through the following link: https://zenodo.org/records/10210286.

Distribution of H. halys in Belgium

o se

The 740 observations from 6 years (2017–2022) were checked for accuracy and completeness. Since the first record in 2011, occurrences in subsequent years initially remained low, without any public citizen database records or specimens collected until 2017. *Halyomorpha halys* was recorded once in 2017 and 2018, 5 times in 2019 and 35 times in 2020. In recent years, the number of observations of *H. halys* has increased dramatically. In 2021, 183 observations were recorded, followed by a substantial increase to 515 in 2022. Up until November 2023, the Belgian public citizen database reported more than 2200 observations, a more than tenfold increase compared to 2021. From the occurrences of *H. halys* in 2020–2022, most of the observations were made in northern Belgium, mainly around the urban areas of the cities of Gent, Leuven, and Mechelen and the region of Haspengouw (Fig. 2a–c).



Figure 2. Map of Belgium with recorded occurrences of *H. halys* in **A** 2020 **B** 2021, and **C** 2022. Each dot is an individual record, and density clouds indicate the level of density of occurrences in one area.

Discussion

This study uncovered 9 COI, 5 COII and 12 COI-COII distinct haplotypes from 99 *H. halys* specimens collected in Belgium. Among these, two new COI-COII haplotype combinations (H03h11 and H08h21), currently unique to Belgium, were observed.

The principal COI haplotypes present in Belgium were H01 (33%), H08 (24%) and H03 (23%). H01 and H03 are the most frequent haplotypes in China and in most invaded countries (Gariepy et al. 2014; Valentin et al. 2017; Cesari et al. 2018; Lee et al. 2018; Yan et al. 2021; Gariepy et al. 2021). As such, their presence in Belgium is to be expected and therefore does not help to determine the origin of *H. halys* in Belgium. Interestingly, however, H08 is the second most abundant haplotype in Belgium. So far, this haplotype has been exclusively detected in certain European countries: Austria, France, Italy, Switzerland and Serbia (Valentin et al. 2017; Cesari et al. 2018; Gariepy et al. 2021; Yan et al. 2021). This suggests that at least part of the H. halys population in Belgium originates from previously invaded territories in Europe (i.e., bridgehead effect; Lombaert et al. 2010). Remarkably, eight out of the nine COI haplotypes found in Belgium are shared with Italy. Furthermore, half of these haplotypes were reported exclusively in these two countries (H40c, H41c, H42c and H49c) suggesting that Italy is a direct region of origin of H. halys in Belgium. The rare H23 haplotype was only found once in Belgium and has been reported in Japan and the Western USA (Oregon) (Valentin et al. 2017). It was also detected in a shipment in Australia originating from Italy, suggesting that the H23 haplotype is also present in Italy (Horwood et al. 2019; Gariepy et al. 2021). Introductions may thus have originated from Japan or the USA in addition to Italy.

For the COII fragment, h01, h03 and h11 each accounted for 32% of the abundance. The h01 and h03 haplotypes are native to China and Korea (but have not been reported in Japan yet) and are the most frequent COII haplotypes in invaded countries (Xu et al. 2014; Cesari et al. 2018; Yan et al. 2021). The distribution of the h11 COII haplotype is more restricted and has only been reported in Austria, Italy, Japan, and Korea (Xu et al. 2014; Cesari et al. 2018; Yan et al. 2021). Consistent with Italy as a source of *H. halys* for Belgium, the h21 haplotype was isolated from two specimens collected in Belgium and is only shared with Italy (Cesari et al. 2018).

In this study, the combination of COI and COII fragments did not result in a significantly better geographic resolution to reveal possible origins of the invasion.

However, the presence of haplotype H03h21 provides some additional support for Italy as a source of the invasion since this combination is only shared with a sample from the Piedmont region of Italy (Cesari et al. 2018). Additionally, the inclusion of the COII fragment in this study revealed two novel haplotype combinations unique to Belgium: H08h21 and H03h11. The former is likely to originate from Italy since haplotype h21 has been exclusively reported there (Piedmont region, same sample as for H03h21), while H08 has been detected multiple times (Piedmont and Lombardy regions of Italy). H03h11 is more singular because it combines a COI haplotype native to China (H03) and a COII haplotype native to Korea or Japan (h11; see above). Such a combination could arise from recent COI or COII mutations. H03 is only two substitutions from the closest COI haplotype native to Japan (H27), and h11 is only one substitution from three COII haplotypes native to China (h03, h05 and h10). Alternatively, this combination could also exist in the regions of origin (China and/or Japan/Korea) but has not yet been sampled. Belgium's dominant haplotypes generally overlap with those from previously invaded European countries, such as Italy, Switzerland, Austria, and Hungary. In contrast, rarer haplotypes are exclusively shared with Italy and its native region of Japan (H23).

The genetic diversity found in Belgium is surprisingly high (Hd = 0.79, $\pi = 0.0018$, N = 95) compared to other invaded countries previously studied (Table 2), several distinct haplotypes were isolated from single sampling sites (Suppl. material 1). A reduced genetic diversity is usually expected for introduced species because of a limited number of founders associated with early genetic drift. However, genetic bottlenecks associated with introductions can be counteracted if many individuals are introduced at the same time or if repeated introductions occur (Dlugosch and Parker 2008). Similarly, Schuler et al. (2020) found a high haplotype diversity (Hd = 0.68, π = 0.0046, N = 156) in the H. halys population of South Tyrol in Northern Italy. The population in Tyrol was established by a secondary introduction from Eastern USA into the Emilia Romagna region of Italy and the spread of the founder population in Switzerland to the neighbouring countries. In contrast, Valentin et al. (2017) concluded that most invasive populations in North America and Europe were established from a direct introduction of H. halys from China with separate introductions into Eastern and Western USA and Canada, as well as into Switzerland and Greece. The high genetic diversity of *H. halys* populations in Belgium and the overlap of haplotypes with previously invaded European countries, such as Italy, Switzerland, Austria, and Hungary but also with Japan indicates that the Belgian populations probably originated from multiple invasions from already invaded European countries, mainly Italy, but also directly from Japan through inadvertent human-mediated transportation (often due to global trade of goods) (Valentin et al. 2017). The weak genetic structure detected by our Mantel test could be related to the recent local expansion of these genetically diverse clusters.

Previous studies have shown that citizen science provides valuable data to characterise the spread of *H. halys* (Maistrello et al. 2016; Stoeckli et al. 2020; Streito et al. 2021). From our data in Belgium, more *H. halys* observations are recorded in the nosrthern part of Belgium compared to the southern part of Belgium, with more urban areas than agricultural, forested, and rural areas. However, on a national scale, spatial biases are commonly associated with human population density, settlements, and road infrastructure (Kelling et al. 2015; Geldmann et al. 2016; Girardello et al. 2019. The bias towards urban areas in citizen-collected occurrence data and rising popularity of tools to make observations (e.g., smartphones) represents a challenge to infer true spatio-temporal patterns from our occurrence data (Bowler et al. 2022). Nevertheless, it also provides natural experimental gradients to examine the impacts of future environmental scenarios including climate change (Lahr et al. 2018). Urban areas are often seen as heat islands, offering more suitable refugia to survive winters or more favourable conditions throughout the seasons to develop and reproduce. Therefore, these urbanised areas often comprise a larger population of insects (Kaiser et al. 2016; Frank and Backe 2023). According to Kistner (2017), the northern part of Belgium lies near the latitudinal border of the climate in Europe suitable for the winter survival and summer reproduction of *H. halys*. In contrast, the southern part of Belgium is currently modelled to be an unsuitable eco-climatic region for *H. halys*, with colder summer and winter temperatures and fewer refugia to survive winter due to the lower urbanisation (Kistner 2017). Niche modelling through climate suitability by Streito et al. (2021) essentially confirms this, with the northern part of Belgium having more suitable areas for *H. halys* while the southern part is more marginal to unsuitable for its survival and expansion. Despite the possibility of spatio-temporal bias in our occurrence data, some temporal trends can still be derived from citizen-collected occurrence data (Powney et al. 2019; Outhwaite et al. 2020; Sheard et al. 2021; Zattara and Aizen 2021; Bowler et al. 2022). Our observational data shows an almost fifteenfold increase in observations from 2020 to 2022. However, to date, no damage related to *H. halvs* has been reported in agricultural production in Belgium (Berteloot et al. 2021). Based on the occurrence data, it is likely that the population expansion of *H. halys* in Belgium is very recent, as a relatively high number of observations were only made in 2021, 2022 and 2023. Streito et al. (2021) define three main phases in the invasion of *H. halys*: (1) the very beginning of the invasion, when populations of *H. halys* are low, and naturalists and official monitoring services who are excellent observers and expecting the emergence of H. halys can detect its presence, (2) when abundance increases, non-naturalist citizens are able to detect the species and start to provide information on the dynamics of the invasion and indirectly on the level of populations through citizen science platforms, and (3) when populations have expanded and become large enough, agricultural professionals take over and can predict and assess the population density and damages. From our occurrence data, the H. halys population in Belgium is seemingly under expansion, being numerously recorded in the citizen science databases.

Lastly, the univoltine *H. halys* population in Belgium likely expands in the summer and declines in the winter in current climatic conditions due to Belgium being situated at the northern latitudinal border of climatic suitability for *H. halys*, with winters cold enough to kill more individuals than winter temperatures in southern European countries like Italy, possibly delaying the fast expansion of its populations. However, both parts of Belgium are modelled to be suitable for the survival and development of *H. halys* by 2100 (Kistner 2017).

Conclusions

The results of this study provide haplotype information for *H. halys* from a newly invaded region. The haplotype diversity in Belgium is surprisingly high, with 9 COI, 5 COII and 12 COI-COII haplotypes found. The invasion of *H. halys* in Belgium likely occurred repeatedly and is assumed to be still ongoing through human-mediated transportation from other invaded European countries and directly from its native regions in Eastern Asia. A significant overlap between Belgian and Italian haplotypes points to Italy as the most probable source for a significant proportion of haplotypes currently present in Belgium. By combining the citizen-collected occurrence data with the molecular data, we assume the population expansion of *H. halys* is recent and ongoing.

Acknowledgements

This research was in part supported by funds from VLAIO (Flanders Innovation and Entrepreneurship) through LA-traject HBC.2018.2224 ("SOS Penta") and by the DA-TAPESTFRU project (RI 2020-A340) funded by the Belgian Federal Public Service (FPS) Health, Food Chain Safety and Environment. We thank Stéphane Claerebout for providing us one sample collected in 2018 in Mouscron.

References

- Aukema B, Bieman den K, Lommen G, Maat van de G, Troisfontaine L, Vossen P (2019) Nieuwe en interessante nederlandse Wantsen VI (Hemiptera: Heteroptera). Nederlandse Faunistische Mededelingen 46: 57–86.
- Berteloot OH, Vervaet L, Huayan C, Talamas EJ, Van Leeuwen T, De Clercq P (2021) First record in Belgium of *Trissolcus basalis* (Hymenoptera, Scelionidae), an egg parasitoid of economically important stink bugs (Hemiptera, Pentatomidae). Belgian Journal of Zoology 151: 139–148. https://doi.org/10.26496/bjz.2021.90
- Bodenhofer U, Bonatesta E, Horejs-Kainrath C, Hochreiter S (2015) msa: An R package for multiple sequence alignment. Bioinformatics 31(24): 3997–3999. https://doi. org/10.1093/bioinformatics/btv494
- Bowler DE, Callaghan CT, Bhandari N, Henle K, Barth MB, Koppitz C, Klenke R, Winter M, Bruelheide H, Bonn A (2022) Temporal trends in the spatial bias of species occurrence records. Ecography 8(8): e06219. https://doi.org/10.1111/ecog.06219
- Bras A, Avtzis DN, Kenis M, Li H, Vétek G, Bernard A, Courtin C, Rousselet J, Roques A, Auger-Rozenberg M-A (2019) A complex invasion story underlies the fast spread of the invasive box tree moth (*Cydalima perspectalis*) across Europe. Journal of Pest Science 92(3): 1187–1202. https://doi.org/10.1007/s10340-019-01111-x
- Bulgarini G, Badra Z, Leonardi S, Maistrello L (2020) The predatory ability of generalist predators on eggs, young nymphs and adults of the invasive *Halyomorpha halys* in southern Europe. BioControl 66(3): 355–366. https://doi.org/10.1007/s10526-020-10066-3

- Cesari M, Maistrello L, Ganzerli F, Dioli P, Rebecchi L, Guidetti R (2015) A pest alien invasion in progress: Potential pathways of origin of the brown marmorated stink bug *Halyomorpha halys* populations in Italy. Journal of Pest Science 88(1): 1–7. https://doi.org/10.1007/ s10340-014-0634-y
- Cesari M, Maistrello L, Piemontese L, Bonini R, Dioli P, Lee W, Park C, Partsinevelos GK, Rebecchi L, Guidetti R (2018) Genetic diversity of the brown marmorated stink bug *Ha-lyomorpha halys* in the invaded territories of Europe and its patterns of diffusion in Italy. Biological Invasions 20(4): 1073–1092. https://doi.org/10.1007/s10530-017-1611-1
- Claerebout S, Haye T, Olafsson E, Pannier E, Bultot J (2018) Premières occurrences de *Halyo-morpha halys* (Stål, 1855) pour la Belgique et actualisation de sa répartition en Europe (Hemiptera: Heteroptera: Pentatomidae). Bulletin de la Société Royale Belge d'Entomologie 154: 205–227.
- Commission EU (2022a) The apple market in the EU: Production, yields and areas. EU Fruit and Vegetables market observatory, pip fruit subgroup. https://agriculture.ec.europa.eu/ system/files/2022-10/apples-production_en.pdf
- Commission EU (2022b) The pear market in the EU: Production, yields and areas. EU Fruit and Vegetables market observatory, pip fruit subgroup. https://agriculture.ec.europa.eu/ system/files/2022-10/pears-production_en.pdf
- Cristescu ME (2016) Genetic reconstructions of invasion history. In: Barret SCH, Colautti RI, Dlugosch KM, Rieseberg LH (Eds) Invasion Genetics. John Wiley & Sons, Ltd., Chichester, 267–282. https://doi.org/10.1002/9781119072799.ch15
- Dlugosch KM, Parker IM (2008) Founding events in species invasions: Genetic variation, adaptive evolution, and the role of multiple introductions. Molecular Ecology 17(1): 431–449. https://doi.org/10.1111/j.1365-294X.2007.03538.x
- Faúndez EI, Rider DA (2017) The brown marmorated stink bug *Halyomorpha halys* (Stål, 1855) (Heteroptera: Pentatomidae) in Chile. Arquivos Entomolóxicos, 305–307.
- Ficetola GF, Bonin A, Miaud C (2008) Population genetics reveals origin and number of founders in a biological invasion. Molecular Ecology 17(3): 773–782. https://doi.org/10.1111/ j.1365-294X.2007.03622.x
- Folmer O, Black M, Hoeh W, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3(5): 294–299.
- Frank SD, Backe KM (2023) Effects of urban heat islands on temperate forest trees and arthropods. Current Forestry Reports 9(1): 48–57. https://doi.org/10.1007/s40725-022-00178-7
- Gapon DA (2016) First records of the brown marmorated stink bug *Halyomorpha halys* (Stål, 1855) (Heteroptera, Pentatomidae) in Russia, Abkhazia, and Georgia. Entomological Review 96(8): 1086–1088. https://doi.org/10.1134/S001387381608011X
- Gariepy TD, Haye T, Fraser H, Zhang J (2014) Occurrence, genetic diversity, and potential pathways of entry of *Halyomorpha halys* in newly invaded areas of Canada and Switzerland. Journal of Pest Science 87(1): 17–28. https://doi.org/10.1007/s10340-013-0529-3
- Gariepy TD, Musolin DL, Konjević A, Karpun NN, Zakharchenko VY, Zhuravleva EN, Tavella L, Bruin A, Haye T (2021) Diversity and distribution of cytochrome oxidase I (COI) haplotypes of the brown marmorated stink bug, *Halyomorpha halys* Stål (Hemiptera,

Pentatomidae), along the eastern front of its invasive range in Eurasia. NeoBiota 68: 53–77. https://doi.org/10.3897/neobiota.68.68915

- Geldmann J, Heilmann-Clausen J, Holm TE, Levinsky I, Markussen B, Olsen K, Rahbek C, Tøttrup AP (2016) What determines spatial bias in citizen science? Exploring four recording schemes with different proficiency requirements. Biodiversity Research 22(11): 1139–1149. https://doi.org/10.1111/ddi.12477
- Girardello M, Chapman A, Dennis R, Kaila L, Borges PAV, Santangeli A (2019) Gaps in butterfly inventory data: A global analysis. Biological Conservation 236: 289–295. https://doi. org/10.1016/j.biocon.2019.05.053
- Güncan A, Gümüş E (2019) Brown marmorated stink bug, *Halyomorpha halys* (Stål, 1855) (Hemiptera: Heteroptera, Pentatomidae), a new and important pest in Turkey. Entomological News 128(2): 204–210. https://doi.org/10.3157/021.128.0208
- Hamelin RC, Roe AD (2019) Genomic biosurveillance of forest invasive alien enemies: A story written in code. Evolutionary Applications 13(1): 95–115. https://doi.org/10.1111/eva.12853
- Hamilton GC, Ahn JJ, Bu W, Leskey TC, Nielsen AL, Park YL, Rabitsch W, Hoelmer KA (2018) *Halyomorpha halys* (Stål). In: McPherson JE (Ed.) Invasive Stink Bugs and Related Species (Pentatomoidea): Biology, Higher Systematics, Semiochemistry, and Management. CRC Press, Taylor & Francis, Boca Raton, 243–292. https://doi.org/10.1201/9781315371221-4
- Hartung V, Heller M, Husemann M (2022) First record of the invasive true bug (Heteroptera) Halyomorpha halys (Stål, 1855) from Hamburg with other records from Northern Germany and possible implications for its range expansion. Evolutionary Systematics 6(1): 57–64. https://doi.org/10.3897/evolsyst.6.80744
- Haye T, Gariepy T, Hoelmer K, Rossi J-P, Streito J-C, Tassus X, Desneux N (2015) Range expansion of the invasive brown marmorated stinkbug, *Halyomorpha halys*: An increasing threat to field, fruit and vegetable crops worldwide. Journal of Pest Science 88(4): 665–673. https://doi.org/10.1007/s10340-015-0670-2
- Hoebeke ER, Carter ME (2003) Halyomorpha halys (Stål) (Heteroptera: Pentatomidae): a polyphagous plant pest from Asia newly detected in North America. Proceedings of the Entomological Society of Washington 105: 225–237.
- Horwood M, Milnes JM, Cooper WR (2019) Brown marmorated stink bug, *Halyomorpha halys* (Hemiptera: Pentatomidae), detections in Western Sidney, New South Wales, Australia. Austral Entomology 58(4): 857–865. https://doi.org/10.1111/aen.12421
- Jombart T (2008) adegenet: A R package for the multivariate analysis of genetic markers. Bioinformatics 24(11): 1403–1405. https://doi.org/10.1093/bioinformatics/btn129
- Kaiser A, Merckx T, Van Dyck H (2016) The Urban Heat Island and its special scale dependent impact on survival and development in butterflies of different thermal sensitivity. Ecology and Evolution 6(12): 4129–4140. https://doi.org/10.1002/ece3.2166
- Kapantaidaki DE, Evangelou VI, Morrison WR III, Leskey TC, Brodeur J, Milonas P (2019) Halyomorpha halys (Hemiptera: Pentatomidae) Genetic diversity in North America and Europe. Insects 10(6): e174. https://doi.org/10.3390/insects10060174
- Kelling S, Fink D, La Sorte FA, Johnston A, Bruns NE, Hochachka WM (2015) Taking a 'Big Data' approach to data quality in a citizen science project. Ambio 44(S4): 601–611. https://doi.org/10.1007/s13280-015-0710-4

- Kistner EJ (2017) Climate change impacts on the potential distribution and abundance of the brown marmorated stink bug (Hemiptera: Pentatomidae) with special reference to North America and Europe. Environmental Entomology 46(6): 1212–1224. https://doi. org/10.1093/ee/nvx157
- Kuhar TP, Kamminga KL, Whalen J, Dively GP, Brust G, Hooks CRR, Hamilton G, Herbert DA (2012) The pest potential of brown marmorated stink bug on vegetable crops. Plant Health Progress 13(1): 1–41. https://doi.org/10.1094/PHP-2012-0523-01-BR
- Lahr EC, Dun RR, Frank SD (2018) Getting ahead of the curve: Cities as surrogates for global change. Proceedings of the Royal Society B, Biological Sciences 285(1882): e20180643. https://doi.org/10.1098/rspb.2018.0643
- Lee D-H, Short BD, Joseph SV, Bergh JC, Leskey TC (2013) Review of the biology, ecology, and management of *Halyomorpha halys* (Hemiptera: Pentatomidae) in China, Japan, and the Republic of Korea. Environmental Entomology 42(4): 627–641. https://doi. org/10.1603/EN13006
- Lee W, Guidetti R, Cesari M, Gariepy TD, Park Y-L, Park C-G (2018) Genetic diversity of *Halyomorpha halys* (Hemiptera, Pentatomidae) in Korea and Comparison with COI sequence datasets from East Asia, Europe, and North America. The Florida Entomologist 101(1): 49–54. https://doi.org/10.1653/024.101.0110
- Leskey TC, Hamilton GC, Nielsen AL, Polk DF, Rodriguez SC, Bergh JC, Herbert DA, Kuhar TP, Pfeiffer D, Dively GP, Hooks CRR, Raupp MJ, Shrewsbury PM, Krawczyk G, Shearer PW, Whalen J, Koplinka LC, Myers E, Inkley D, Hoelmer KA, Lee D-H, Wright SE (2012) Pest status of the brown marmorated stink bug, *Halyomorpha halys* in the USA. Outlooks on Pest Management 23(5): 218–226. https://doi.org/10.1564/23oct07
- Lombaert E, Guillemaud T, Comuet J-M, Malausa T, Facon B, Estoup A (2010) Bridgehead effect in the worldwide invasion of the biocontrol harlequin ladybird. PLoS ONE 5(3): e9743. https://doi.org/10.1371/journal.pone.0009743
- Macavei LI, Baetan R, Oltean I, Florian T, Varga M, Costi E, Maistrello L (2015) First detection of *Halyomorpha halys* Stål, a new invasive species with a high potential of damage on agricultural crops in Romania. Lucrări Științifice 58: e2015.
- Maistrello L, Dioli P, Vaccari G, Nannini R, Bortolotti P, Caruso S, Costi E, Montermini A, Casoli L, Bariselli M (2014) Primi rinvenimenti in Italia della cimice esotica *Halyomorpha halys*, una nuova minaccia per la frutticoltura. Giornate Fitopatologiche 1: 283–288.
- Maistrello L, Dioli P, Bariselli M, Mazzoli GL, Giacalone-Forini L (2016) Citizen science and early detection of invasive species: Phenology of first occurrences of *Halyomorpha halys* in Southern Europe. Biological Invasions 18(11): 3109–3116. https://doi.org/10.1007/ s10530-016-1217-z
- Milonas PG, Partsinevelos GK (2014) First report of brown marmorated stink bug *Halyo-morpha halys* Stål (Hemiptera: Pentatomidae) in Greece. Bulletin OEPP. EPPO Bulletin. European and Mediterranean Plant Protection Organisation 44(2): 183–186. https://doi. org/10.1111/epp.12129
- Observation International and local partners (2022) Observation International and local partners [waarnemingen.be/observations.be dataset *Halyomorpha halys*]. [retrieved June 5, 2023, from] https://waarnemingen.be/species/197583/

- Outhwaite CL, Gregory RD, Chandler RE, Collen B, Isaac NJB (2020) Complex long-term biodiversity change among invertebrates, bryophytes and lichens. Nature Ecology & Evolution 4(3): 384–392. https://doi.org/10.1038/s41559-020-1111-z
- Paradis E (2010) pegas: An R package for population genetics with an integrated-modular approach. Bioinformatics 26(3): 419–420. https://doi.org/10.1093/bioinformatics/btp696
- Powney GD, Carvell C, Edwards M, Morris RKA, Roy HE, Woodcock BA, Isaac NJB (2019) Widespread losses of pollinating insects in Britain. Nature Communications 10(1): e1018. https://doi.org/10.1038/s41467-019-08974-9
- QGIS Development Team (2023) QGIS Geographic Information System. Open Source Geospatial Foundation Project. http://qgis.osgeo.org
- R Core Team (2020) R: A Language and Environment for statistical Computing. R Foundation for Statistical Computing, Vienna. https://www.R-project.org/
- Rabitsch W, Friebe G (2015) From the west and from the east? First records of *Halyomorpha halys* (Stål, 1855) (Hemiptera: Heteroptera: Pentatomidae) in Vorarlberg and Vienna, Austria. Beiträge zur Entomofaunistik 16: 115–139.
- Rice KB, Bergh CJ, Bergmann EJ, Biddinger DJ, Dieckhoff C, Dively G, Fraser H, Gariepy T, Hamilton G, Haye T, Herbert A, Hoelmer K, Hooks CR, Jones A, Krawczyk G, Kuhar T, Martinson H, Mitchell W, Nielsen AL, Pfeiffer DG, Raupp MJ, Rodriguez-Saona C, Shearer P, Shrewsbury P, Venugopal PD, Whalen J, Wiman NG, Leskey TC, Tooker JF (2014) Biology, ecology, and management of brown marmorated stink bug (Hemiptera: Pentatomidae). Journal of Integrated Pest Management 5(3): 1–13. https://doi. org/10.1603/IPM14002
- Rollins LA, Woolnough AP, Sinclair R, Mooney NJ, Sherwin WB (2011) Mitochondrial DNA offers unique insights into invasion history of the common starling: Mitochondrial DNA in invasive populations. Molecular Ecology 20(11): 2307–2317. https://doi.org/10.1111/ j.1365-294X.2011.05101.x
- Rot M, Devetak M, Carlevaris B, Žežlina J, Žežlina I (2018) First record of brown marmorated stink bug (*Halyomorpha halys* Stål, 1855) (Hemiptera: Pentatomidae) in Slovenia. Acta Entomologica Slovenica 26: 5–12. https://doi.org/10.3390/insects12060505
- Ryan SF, Lombaert E, Espeset A, Vila R, Talavera G, Dincă V, Doellman MM, Renshaw MA, Eng MW, Hornett EA, Li Y, Pfrender ME, Shoemaker D (2019) Global invasion history of the agricultural pest butterfly *Pieris rapae* revealed with genomics and citizen science. Proceedings of the National Academy of Sciences of the United States of America 116(40): 20015–20024. https://doi.org/10.1073/pnas.1907492116
- Schuler H, Elser D, Fischnaller S (2020) Population genetics of the brown marmorated stink bug *Halyomorpha halys* in the early phase of invasion in South Tyrol (Northen Italy). Bulletin of Entomological Research 111(4): 394–401. https://doi.org/10.1017/ S0007485320000553
- Šeat J (2015) *Halyomorpha halys* (Stål, 1855) (Heteroptera: Pentatomidae) a new invasive species in Serbia. Acta Entomologica Serbica 20: 167–171.
- Sheard JK, Rahbek C, Dunn RR, Sanders NJ, Isaac NJB (2021) Long-term trends in the occupancy of ants revealed through use of multi-sourced datasets. Biology Letters 17(10): e20210240. https://doi.org/10.1098/rsbl.2021.0240

- Stoeckli S, Felber R, Haye T (2020) Current distribution and voltinism of the brown marmorated stink bug, Halyomorpha halys, in Switzerland and its response to climate change using a high-resolution CLIMEX model. International Journal of Biometeorology 64(12): 2019–2032. https://doi.org/10.1007/s00484-020-01992-z
- Streito J-C, Chartois M, Pierre E, Dusoulier F, Armand J-M, Gaudin J, Rossi J-P (2021) Citizen science and niche modeling to track and forecast the expansion of the brown marmorated stinkbug *Halyomorpha halys* (Stål, 1855). Scientific Reports 11(1): e11421. https:// doi.org/10.1038/s41598-021-90378-1
- Valentin RE, Nielsen AL, Wiman NG, Lee D-H, Fonseca DM (2017) Global invasion network of the brown marmorated stink bug, *Halyomorpha halys*. Scientific Reports 7(1): e9866. https://doi.org/10.1038/s41598-017-10315-z
- Verbond van Belgische Tuinbouwcoöperaties (2022) Verbond van Belgische Tuinbouwcoöperaties. Annual report. https://www.vbt.eu/en/annual-reports
- Vétek G, Papp V, Haltrich A, Rédei D (2014) First record of the brown marmorated stink bug, *Halyomorpha halys* (Hemiptera: Heteroptera: Pentatomidae), in Hungary, with description of the genitalia of both sexes. Zootaxa 3780: 194–200. https://doi.org/10.11646/zootaxa.3780.1.8
- Walsh PS, Metzger DA, Higuchi R (1991) Chelex 100 as a medium for simple extraction of DNA for PCR-based typing from forensic material. BioTechniques 10: 506–513.
- Xu J, Fonseca DM, Hamilton GC, Hoelmer KA, Nielsen AL (2014) Tracing the origin of US brown marmorated stink bugs, *Halyomorpha halys*. Biological Invasions 16(1): 153–166. https://doi.org/10.1007/s10530-013-0510-3
- Yan J, Pal C, Anderson D, Vétek G, Farkas P, Burne A, Fan Q-H, Zhang J, Gunawardana DN, Balan RK, George S, Li D (2021) Genetic diversity analysis of brown marmorated stink bug, *Halyomorpha halys* based on mitochondrial COI and COII haplotypes. BMC Genomic Data 22(1): 1–7. https://doi.org/10.1186/s12863-021-00961-8
- Zattara EE, Aizen MA (2021) Worldwide occurrence records suggest a global decline in bee species richness. One Earth 4(1): 114–123. https://doi.org/10.1016/j.oneear.2020.12.005
- Zhu G-P, Ye Z, Du J, Zhang D-L, Zhen Y, Zheng C, Zhao L, Li M, Bu W-J (2016) Range wide molecular data and niche modeling revealed the Pleistocene history of a global invader (*Halyomorpha halys*). Scientific Reports 6(1): e23192. https://doi.org/10.1038/srep23192

Supplementary material I

Metadata of all samples

Authors: Olivier Hendrik Berteloot, Alexandre Kuhn

Data type: csv

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.113421.suppl1

RESEARCH ARTICLE



Field studies of the ecological impacts of invasive plants in Europe

Montserrat Vilà^{1,2}, Alejandro Trillo¹, Pilar Castro-Díez³, Belinda Gallardo⁴, Sven Bacher⁵

I Estación Biológica de Doñana (EBD-CSIC), 41092 Sevilla, Spain 2 Department of Plant Biology and Ecology, Universidad de Sevilla, 41012 Sevilla, Spain 3 Universidad de Alcalá, Facultad de Ciencias, Departamento de Ciencias de la Vida, Área Ecología, Grupo de Investigación en Invasiones Biológicas, Campus Científico-Tecnológico, E-28805, Alcalá de Henares, Madrid, Spain 4 Applied and Restoration Ecology Group, Pyrenean Institute of Ecology (IPE-CSIC), 50059 Zaragoza, Spain 5 University of Fribourg, Department of Biology, Unit of Ecology and Evolution, 1700 Fribourg, Switzerland

Corresponding author: Montserrat Vilà (montse.vila@ebd.csic.es)

Academic editor: Sven Jelaska | Received 7 September 2023 | Accepted 24 December 2023 | Published 25 January 2024

Citation: Vilà M, Trillo A, Castro-Díez P, Gallardo B, Bacher S (2024) Field studies of the ecological impacts of invasive plants in Europe. NeoBiota 90: 139–159. https://doi.org/10.3897/neobiota.90.112368

Abstract

The impacts of invasive species can vary widely across invaded sites and depend on the ecological variable of study. In this paper, we describe the first harmonised database that compiles scientific evidence of the ecological impacts of invasive plant species at continental scale. We summarise results from 266 publications reporting 4259 field studies on 104 invasive species in 29 European countries. For each study, we recorded whether the effects were statistically significant and noted their direction (i.e. decrease or increase in the response variable when compared to uninvaded sites). We classified studies, based on the impacts on the levels of ecological organisation (species, communities and ecosystems), taxa and trophic level. More than half of the studies were conducted in temperate and boreal forests and woodlands and temperate grasslands. Notably, one third of the studies focused on just five invasive species. Most studies were on native species followed by studies on communities. Impacts on plants were more frequently studied than impacts on other taxa and trophic groups. Overall, 43% of the studies reported significant impacts, with more significant decreases (26%) than increases (17%) in the response variables. Significant impacts were more frequent on species and communities than on ecosystems; and on plants than on animals or microbes. This database is of interest for academic, management and policy-related purposes.

Keywords

Biological invasions, context-dependence, diversity, ecological organisation, ecosystem properties, European Regulation on IAS, non-native plants, trophic level

Copyright Montserrat Vilà et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

Many non-native species introduced by human agency outside their original area of distribution invade natural areas and cause ecological impacts to native species, communities and ecosystems (Simberloff et al. 2013). Ecological impacts are defined in this paper as any statistically significant ecological change occurring when an invasive species is present compared to when the invasive species is absent. This change can be a decrease (i.e. negative impact) or an increase (i.e. positive impact) of any ecological attribute of the invaded ecosystem (Jeschke et al. 2014). Thus, it is important to note that negative and positive impacts are independent of ethical and societal human values (Vimercati et al. 2020).

Information on the impacts of invasive species is of fundamental importance to assist management and policy (Vilà et al. 2019). In particular, empirical studies of ecological impacts provide essential scientific evidence to underpin risk assessment of invasions that are often used to rank and prioritise management actions. Despite the fast increase in the number of field studies testing for invasive species impacts, the majority focus on a few species and regions. Consequently, there are still important biases and gaps in knowledge that preclude our capacity to provide information for management and policy actions (IPBES 2023). It is thus of paramount importance to synthesise the scientific evidence on impacts to identify which are the most studied invasive species, the most studied habitats and the most studied impact types.

Meta-analyses have shown a strong context-dependency not only in the magnitude, but also in the direction of the impacts (Pyšek et al. 2012; Gallardo et al. 2016; Volery et al. 2021; Romero-Blanco et al. 2023). For example, an invasive N-fixing plant may strongly increase soil fertility in a recipient ecosystem with N-deficient soils and lacking native N-fixing species, but may have negligible impacts in communities including native N-fixing plants or in soils otherwise rich in N (Vitousek and Walker 1989; Castro-Díez et al. 2014, 2016). Moreover, invasive species can cause multiple, sometimes contrasting, impacts at different levels of ecological organisation (species, communities and ecosystems). For example, an invasive N-fixing plant may increase N soil availability and this can favour the establishment of some native plant species at the expense of others, with a neutral effect on species richness (Marchante et al. 2011). Thus, the impact of an invasive species can vary, presenting a neutral, negative or positive effect. This variability depends on factors such as the identity of the native species under study or whether the focus is on particular native species or the entire community. Therefore, to guide management decisions and biodiversity conservation efforts, it is important to document a broad spectrum of numerical increases and decreases in ecological responses following invasion. Given the conservation interest in native species and communities, adopting a value-laden perspective, their decrease may be considered deleterious, while an increase may be considered beneficial (Vimercati et al. 2020).

The environmental assessment of the impacts of invasive species requires the analysis of the full range of ecological changes after invasion. To this end, we conducted a comprehensive review of field studies reporting ecological impacts of invasive plant species in Europe to identify the most studied species, countries and habitats and to describe the frequency and direction of impacts. We classified impacts according to four criteria: i) the ecological level at which the impact is measured, i.e. species, communities and ecosystems; ii) the affected taxonomic level, i.e. microbes, plants and animals; and iii) the trophic level of the affected taxa. Beyond identifying the most-studied invasive plant species and habitats and their most-studied impacts, our database also enables the exploration of differences in the frequencies and directions of impact types studied. Specifically, we explored if there were differences in the frequency of impacts amongst levels of organisation and taxa.

Material and methods

We started from the studies conducted in Europe extracted from the databases constructed and analysed in Pyšek et al. (2012) and in Castro-Díez et al. (2019). Additionally, we searched for new publications on the Web of Science (https://www.webofscience.com/ wos/alldb/basic-search) database on 31 December 2022 with no restriction on publication year, using the following search term combinations: (plant inva* OR exotic plant OR alien plant OR non-native plant) AND (impact* OR effect*) AND (community structure* OR diversity* OR ecosystem process* OR competition*). Amongst the retrieved documents, we first screened titles and abstracts to identify all publications on the impacts of invasive plants conducted in Europe. We then examined each publication and constructed a database of impacts according to the following selection criteria:

1. The studies had to be in natural or semi-natural field conditions in Europe. The habitat type of the study was classified according to the IPBES unit of analysis (IPBES 2018) with the exception that, in this study, coastal areas was considered for terrestrial plants. The evidence of impact was based on observational or experimental (i.e. removal or addition of target species) field studies comparing simultaneously invaded or uninvaded sites where the identity of single invasive species of study was explicitly mentioned. We excluded tree plantations. We also excluded those referring to impacts by several invasive species.

2. When the same publication examined different response variables, different invasive species, different ecosystem types or geographically different localities, we considered each as different entries in the database (i.e. study, hereafter).

3. When a response variable was measured at different times (e.g. sampling species diversity across years), we made an informed decision on whether to take the mean value across times or to consider each measure as independent. However, when the variable was repeated across short periods (e.g. sampling N availability in different weeks), we only used the final measurement or the most representative (e.g. when the soil activity was the highest).

4. When the study manipulated other environmental factors in addition to invasion, we only considered results from the non-manipulated plots.

5. When the study investigated the effects of different degrees of invasion and different residence times (i.e. old vs. recent invasions), we examined differences between the least invaded sites and the most invaded sites and differences between uninvaded sites and sites with the longest time since invasion. As all studies dealt with established non-native plant species and their threats to biological diversity and/or ecosystems, for simplicity, we refer to them as "invasive species" through out the text.

Following Vilà et al. (2011), impacts were classified according to the affected level of ecological organisation as follows: impacts to native species (e.g. abundance, performance, biomass), to communities (e.g. abundance, biomass or diversity) and to ecosystem properties (e.g. soil C/N, nutrient fluxes, decomposition rates, pH, nutrient pools, resource availability, soil minerals, soil organic matter and soil salinity/cation exchange capacity) (Table 1, Suppl. material 1: table S1). Furthermore, when the information was available, the impacts to species and communities were also classified according to the affected taxa (i.e. microbes, plants and animals) and to the trophic level of the affected taxa (i.e. impacts to herbivores, parasites, plants, pollinators, predators, omnivores, decomposers and symbionts) (Table 2). If the native species of concern belongs to different trophic levels along its life history, we considered the one during the stage of the study. In total, the database considered 23 impact types, which integrate the main biodiversity and ecosystem changes after invasion and allow for comparing impacts across studies (see Tables 1, 2).

For each impact, we recorded the statistical significance (no/yes) and direction (increase/decrease) of differences between invaded and uninvaded plots. For the purpose of this analysis, the direction does not mean a desirable/undesirable impact, but a significant increase or decrease of the response variable analysed in the invaded compared to the uninvaded control treatment, respectively.

To search for differences in the frequency of significant impacts across different levels of organisation (species, community, ecosystem) and taxa (animals, microbes, plants), we summed the number of responses – whether significant or non-significant – for each impact type. Responses were grouped, based on the identity of the invasive species and the respective publication.

We employed generalised linear mixed models (GLMMs) with a binomial (logit link function) error distribution family (lmerTest package; Kuznetsova et al. (2017)). The response variable was a two-column matrix generated using the 'cbind' function to combine the counts of significant and non-significant impacts. In each model, we included as a fixed factor the levels of ecological organisation or taxa. To account for the non-independence of data, we included the publication and the identity of the invader as random factors. Post-hoc Tukey tests (emmeans package; Russell (2018)) were then applied to evaluate differences in impact frequencies amongst levels of ecological organisation and taxa. To visualise these differences, we used the package ggeffects (Lüdecke 2018). All analyses were performed in R (v. 4.2.1, R Core Team (2022)).

Open research statement

All data employed in this research are archived in Figshare repository https://doi. org/10.6084/m9.figshare.23579082.

Table 1. Ecological impacts of invasive plant species studied in field conditions in Europe classified by categories of ecological organization (species, communities, ecosystems), with indication of the response variables examined in the literature. In parenthesis, the sample size (number of field studies testing for impacts). See Table S1 for definitions of the impact type classification.

Level of ecological	Impact type	Variables related to
organization		
Species (576)	Animal (176)	Animal abundance (143), activity (10), fitness (4),
		performance (19)
	Microbial (5)	Microbial abundance (5)
	Plant (395)	Plant abundance (223), biomass (34), fitness (66),
		performance (72)
Community (2541)	Animal (1142)	Animal abundance (682), activity (3), biomass (11),
		diversity (446)
	Microbial (370)	Microbial abundance (111), activity (150), biomass
		(17), diversity (92)
	Plant (1016)	Plant abundance (254), biomass (130), diversity (632)
Ecosystem (1155)	Soil carbon to nitrogen ratio (C/N) (74)	C/N (74)
	Nutrient fluxes (25)	C fluxes (11), N fluxes (14)
	Decomposition rates (39)	Litter decomposition (38), soil organic matter
		mineralization (1)
	pH (134)	pH (134)
	Nutrient pools (402)	C pools (114), N pools (194), P pools (94)
	Resource availability (83)	Light (19), moisture (60), soil temperature (4)
	Soil minerals (264)	Soil minerals (264)
	Soil organic matter (85)	Soil organic matter (85)
	Soil salinity/cation exchange capacity (CEC) (49)	Soil CEC (1), salinity (3), salinity/CEC (45)

Table 2. Ecological impacts of invasive plant species studied in field conditions in Europe classified by the trophic level of affected species (i.e. decomposers, herbivores, omnivores, parasites, plants, pollinators, predators, symbionts) with indication of the response variables examined in the literature. In parenthesis, sample size (number of field studies testing for impacts).

Trophic level of the affected	Variables related to			
taxa				
Decomposer (269)	Decomposer abundance (189), biomass (7), diversity (62), activity (11)			
Herbivore (100)	Herbivore abundance (62), diversity (36), performance (2)			
Omnivore (47)	Omnivore abundance (41), diversity (3), fitness (2), performance (1)			
Parasite (50)	Parasite abundance (44), biomass (2), diversity (4)			
Plant (1411)	Plant abundance (477), biomass (164), diversity (632), fitness (66), performance (72)			
Pollinator (353)	Pollinator abundance (190), activity (9), diversity (142), fitness (2), performance (10)			
Predator (287)	Predator abundance (224), activity (4), biomass (1), diversity (54), performance (4)			
Symbiont (23)	Symbiont abundance (16), biomass (1), diversity (6)			

Results

Our final database included 266 publications describing 4259 field studies of 104 invasive plant species in Europe (Fig. 1, Suppl. material 1: tables S2, S3).



Figure 1. Total number of field studies testing for impacts in Europe classified by invasive plant species, ecological organisation and impact type. The grey shading legend indicates whether the impact is on animals, microbes, plants or ecosystems.
Impacts of invasive plants are widely studied across Europe, although around 50% of studies were carried out in six countries (Spain, Poland, Czech Republic, Germany, Italy and Portugal) and there are some countries without any studies (e.g. Albania, Bulgaria, Estonia and Latvia) (see Fig. 2). Most studies were conducted in temperate and boreal forests and woodlands (33%) and temperate grasslands (26%), followed by coastal areas (14%) and Mediterranean forests and woodlands (12%) (Fig. 3).



Figure 2. Map of locations (red dots) of field studies on the ecological impacts of invasive plant species in Europe. Twelve publications described studies in multiple countries and were represented by a dot in each country.



Figure 3. Percentage of publications on field studies testing for impacts classified by the invaded habitat in Europe. Habitats were classified according to the IPBES unit of analysis (IPBES 2018).

While the number of invasive plant species studied has increased linearly since about 2005, the number of publications on impacts have increased exponentially (Fig. 4a). One third of the publications examined the impacts of only five species (Revnoutria japonica, Impatiens glandulifera, Solidago gigantea, Carpobrotus edulis and Robinia pseudoacacia) out of 104 (Fig. 5). The studies on these five species have been concentrated in the last two decades and are still increasing to date (Fig. 4b).

The most studied impacts are on the abundance of species followed by impacts on the abundance and diversity of communities. Impacts on plants have been more studied than impacts on other taxa and trophic groups (Fig. 6). The second most studied impacted group is that of pollinators, followed by predators and decomposers (Fig. 6d). Impacts on microbial communities, although less frequently studied, have increased in the last few years (Fig. 6b). The number of field studies testing for impacts to ecosystem properties have increased one order of magnitude in the last decade (Fig. 4c), with impacts on nutrients pools and soil minerals being the most common (Fig. 6c).

Overall, 43% of studies found significant impacts of invasive plants with more decreases (26%) than increases (17%) on the response variables. Although more than half of the species (58 out of 104) have impacts in both directions, 10% of the invasive species showed only increase responses and 30% decrease responses (Fig. 5).



Figure 4. Cumulative total number of publications on impacts and invasive plant species studied (**a**), number of field studies testing for impacts on the five most studied species (**b**) and across ecological levels of organisation (**c**) in Europe. See Table 1 for impact type classification.

Invasive plant species	Studies	Publications	Frequency (%)
Reynoutria japonica	395	19	39.2 - +
Impatiens glandulifera	333	35	40.2
Solidago gigantea	316	21	39.6
Carpobrotus edulis	224	9	45.1
Robinia pseudoacacia	215	19	41.9
Prunus serotina	206	12	33.5
Acacia longifolia	191	13	66.5
Rosa rugosa	170	7	38.8
Acacia dealbata	158	9	50.0
Solidago canadensis	126	14	41.3
Ailanthus altissima	115	12	47.8
Lupinus polyphyllus	100	11	41.0
Heracleum mantegazzianum	96	8	30.2
Nicotiana glauca	90	1	44.4
Asclepias syriaca	76	8	13.2
Heracleum sosnowskyi	76	4	63.2
Oxalis pes-caprae	72	4	27.8
Pinus mugo	53	7	71.7
Phytolacca americana	51	1	35.3
Quercus rubra	48	4	29.2
Amorpha fruticosa	46	5	43.5
Senecio inaequidens	43	3	37.2
Acacia melanoxylon	43	1	60.5
Erigeron canadensis	42	3	2.4
Reynoutria x bohemica	42	4	59.5
Rhododendron ponticum	41	7	36.6
Symphyotrichum lanceolatum	40	3	40.0
Rudbeckia laciniata	38	4	28.9
Impatiens parviflora	35	6	40.0
Erigeron annuus	29	2	6.9
Campylopus introflexus	28	3	82.1
Prunus laurocerasus	28	1	35.7
Symphyotrichum novi-belgii	27	4	18.5
Sporobolus pumilus	25	2	64.0
Solanum elaeagnifolium	24	4	12.5
Arundo donax	24	2	50.0
Genista aetnensis	23	2	91.3
Acer negundo	22	6	68.2
Carpobrotus acinaciformis	20	5	45.0
Buddleja davidii	20	3	30.0

Figure 5. List of invasive plant species with the total number of field studies testing for impacts, publications and impact frequency (i.e. percentage of significant responses). Blue and orange bars indicate the proportion of decreases and increases, respectively.

Arctotheca calendula	20	2	50.0 - +
Erigeron bonariensis	20	2	40.0
Opuntia dillenii	20	2	40.0
Echinocystis lobata	19	1	31.6
Baccharis halimifolia	18	3	83.3
Echinops sphaerocephalus	18	1	11.1
Parthenocissus quinquefolia	18	1	44.4
Humulus scandens	16	1	31.3
Ambrosia artemisiifolia	15	3	0.0
Hippophae rhamnoides	15	1	0.0
Oenothera drummondii drummondii	15	1	46.7
Pinus pinaster	14	1	57.1
Helianthus tuberosus	13	3	7.7
Cortaderia selloana	12	2	83.3
Rorippa austriaca	12	1	66.7
Sporobolus cryptandrus	12	1	75.0
Gaillardia aristata	11	1	18.2
Lupinus nootkatensis	11	1	72.7
Parthenocissus inserta	11	1	100.0
Vinca minor	11	1	90.9
Malephora crocea	10	1	60.0
Mesembrynthemum crystallinum	10	1	50.0
Gunnera tinctoria	9	3	100.0
Erythranthe guttata	9	2	44.4
Bunias orientalis	9	1	66.7
Sporobolus montevidensis	9	1	33.3
Opuntia stricta	8	2	12.5
Bromus erectus	8	1	75.0
Aizoon pubescens	8	1	100.0
Sulla coronaria	7	2	71.4
Acacia saligna	7	1	28.6
Heracleum persicum	7	1	85.7
Reynoutria sachalinensis	6	2	100.0
Achillea ptarmica	6	1	100.0
Artemisia verlotiarum	6	1	66.7
Epilobium ciliatum subsp. ciliatum	6	1	66.7
Gallium album	6	1	16.7
Phyla nodiflora var. minor	6	1	83.3
Trifolium hybridum	6	1	33.3
Elymus athericus	5	2	100.0
Ulmus pumila	5	2	80.0
Calamagrostis arenaria	5	1	60.0

Acacia pycnantha	4	1	100.0 🗖	+
Symphyotrichum × salignum	4	1	100.0 💻	
Pteridium aquilinum	4	1	50.0	
Sorbaria sorbifolia	4	1	75.0	
Impatiens balfourii	3	1	66.7	
Ludwigia grandiflora	3	1	33.3	
Paspalum distichum	3	1	33.3	
Agave americana	2	1	50.0	
Bothriochloa ischaemum	2	1	100.0	
Calamagrostis epigejos	2	1	100.0	
Carpinus betulus	2	1	100.0	
Castanea sativa	2	1	50.0	
Imperatoria ostruthium	2	1	50.0	
Juglans regia	2	1	100.0	
Rumex alpinus	2	1	100.0	
Sporobolus anglicus	2	1	100.0	
Ficus carica	1	1	100.0	
Paspalum thunbergii	1	1	0.0	
Phacelia tanacetifolia	1	1	100.0	
Pinus sylvestris	1	1	100.0	
Silphium perfoliatum	1	1	0.0	
Xanthium orientale var. albinum	1	1	0.0	

Figure 5. Continued.



Figure 6. Cumulative number of field studies testing for impacts to species (**a**) and communities (**b**) by taxa, ecosystem properties (**c**) and amongst trophic levels (**d**) in Europe. See Tables 1, 2 for impact type classification.

Results on the frequency of significant impacts and their direction can be found in Fig. 7. In studies where the affected level is species, 41% of the impacts (n = 576) were significant, with more decreases (25%) than increases (16%) on the response variables. At the community level, 47% of impacts (n = 2528) were significant, with two times more decreases (32%) than increases (15%). At the ecosystem level, 38% of impacts (n = 1155) were significant, with fewer decreases (15%) than increases (23%). When impacts were classified by the affected trophic levels, altogether 45% of impacts (n = 2807) were significant, with two times more decreases (30%) than increases (15%).

The frequency of significant impacts was similar between the species and community levels (z=0.17, p= 0.99), but higher than at the ecosystem level (z=2.32, p = 0.05 and z = 3.94, p < 0.001, respectively). Additionally, the frequency of significant impacts was similar between animals and microbes (z=0.17, p=0.99), but lower than for plants (z=3.86, p< 0.001 and z=2.94, p< 0.01, respectively) (Fig. 8).



Figure 7. Frequency of significant plant invader impacts vs. percentage of non-significant impacts (grey bars) studied in field conditions in Europe. Blue and orange bars indicate the percentage of decreases and increases, respectively. See Tables 1, 2 for impact type classification.



Figure 8. Frequency of significant plant invader impacts studied in field conditions in Europe across levels of ecological organisation (left) and taxa (right). Open circles are observed values (i.e. the proportion of significant impacts grouped by invader species and publication). Black dots are predicted values obtained from the models and their 95% confidence intervals.

Discussion

Evidence about the impacts of invasive plants on different properties of the recipient ecosystems is scattered across many different studies and technical reports (Kumschick et al. 2015), preventing its efficient transfer to managers and policy-makers. Here, we provide the first harmonised database synthesising results from field studies about the ecological impacts of invasive plants at a continental scale. However, a great proportion of studies focuses on a few invasive species in temperate central European countries or in southern Mediterranean countries. As already indicated ten years ago (Hulme et al. 2013), bias continues to be the norm in the study of impacts, probably reflecting the academic interest of research groups on the most common species in their countries. This database can be of scientific, management and policy use at different national and international scales.

The exploration of impact studies indicates that the main geographic gap of knowledge in Europe corresponds to Baltic and Balkan countries. The least represented habitats in impact studies are desert and xeric shrublands, high mountains and subtropical forests. In Europe, subtropical forests of major conservation status are located in Macaronesian islands, where non-native species invasion is prevalent. Many of these islands exhibit a higher proportion of non-native than native plant species in their flora (Kueffer et al. 2010), emphasising the crucial need to identify invasive species causing major impacts (Silva et al. 2008). The impacts of invasive plants in desert and semi-arid habitats are also poorly studied, despite an increasing number of dry-tolerant invasive plants promoted by ornamental xero-gardening (Morente-López et al. 2023). In the face of climate change, it is also imperative to focus more on the impacts of invasive plants in mountain regions. Climate warming is expected to enhance the dispersal and establishment of invasive species at higher altitudes, making this an area of critical concern (Carboni et al. 2018). Ecological impacts were statistically heterogeneous in their significance and direction. Significant impacts were more frequent on species and communities than on ecosystems. Any change in ecosystem properties can be considered adverse, as it modifies ecosystem functioning (Strayer 2012; Vilà and Hulme 2017; Castro-Díez et al. 2019). Compared to impacts on ecosystems, the impacts on species and communities are more directly linked to changes in biodiversity. According to our database, there were two times more studies reporting negative effects than positive effects on the studied species and community response variables. Negative effects indicate a decrease in native species abundance, fitness or diversity after invasion and are, therefore, considered detrimental for nature conservation. On the contrary, positive effects indicate the reverse and, thus, can be assumed to be beneficial. However, even increasing effects on native species and communities can have cascading effects, depending on the position of the native species in the trophic network (Gallardo et al. 2016).

While the correspondence from value-free to value-laden effects of invaders on biodiversity is not always straightforward (Vimercati et al. 2020), our database on impacts studied in Europe contributes significantly to the global assessment on impacts of invasive species. This comprehensive database aligns with the broader finding that invasive species globally tend to cause more harm than benefits on nature (Bacher et al. 2023).

Significant impacts were more frequently reported on native plants than on native animals or microbes. In general, it seems that invasive species most frequently impact native species from the same broad taxonomic group (Bacher et al. 2023). For plants, this is an expected result because the main mechanism of interaction amongst plants is resource competition or facilitation, while the mechanisms of impact of invasive plants on animals are more diverse and often indirect, depending on the type of interaction, feeding mode and trophic position. Furthermore, impact studies on microbes are relatively recent and predominantly focused on soil bacteria and fungi (Dawson and Schrama 2016). However, it is important to note that, since our focus was on field studies, our review may not have captured all the impacts on plant-soil feedbacks, which are often mediated by microorganisms, such as pathogens or symbionts. It is largely unknown how the strength of plant-soil feedbacks compares with plant-plant competition. This is an area of research which deserves more attention because such interactions influence the co-occurrence of native and invasive species (Lekberg et al. 2018).

Other areas of research interest might include the analysis of the major causes of the variation in impacts and improving their prediction. For this purpose, the information provided in our database could be associated with other aspects of biological invasions (Strayer 2012). For instance, links with their pathways of introduction (Pergl et al. 2017), their local or regional abundances (Bradley et al. 2019), the species traits and the biotic and abiotic characteristics of the invaded habitats (Pyšek et al. 2012; Sapsford et al. 2020) or their invasion history (Lenzner et al. 2022). The frequency and direction of impacts could also be compared to those of invasive animals and amongst invaded areas to determine taxonomic differences in impact across regions.

Causal impacts, together with the probability of arrival and establishment, is one of the main requested information to identify potential invasive species. Therefore,

from a management point of view, the database displays and harmonises the available peer-reviewed publications that can be used for horizon scanning to identify potential invasive species in countries where they are not yet present (e.g. Lucy et al. (2020); Cano-Barbacil et al. (2023)). The information from the database can also be used to populate standardised impact assessments, such as the EICAT-IUCN (Blackburn et al. 2014) and to assist species management prioritisation, based on the magnitude of their impacts on biodiversity.

From a policy perspective, it is important to highlight that, although our analysis screened all European countries, the database does not include information for 29 of the 39 invasive plant species of EU concern (European Union 2014, 2017). Moreover, of the 20 most studied species according to our database, only three are regulated, namely *Impatiens glandulifera*, *Heracleum mantegazzianum* and *H. sosnowskyi*. These mismatches can be explained by some of the features of the EU Regulation (Carboneras et al. 2018). Some regulated species are not yet present in the EU (e.g. many aquatic plant species), but, if introduced, would be capable of establishing self-sustainable populations. On the other hand, some species are present, form self-sustainable populations and cause significant adverse impacts on biodiversity and ecosystem services in Europe, but listing the species will not prevent, minimise or mitigate their impacts and are therefore not listed.

Conclusion

Our first comprehensive European database of the field studies reporting on the ecological effects of invasive non-native plants indicates that invasive plants cause impacts to species, communities and ecosystem processes of a wide range of taxa at different trophic levels. Major gaps in knowledge are found in Baltic and Balkan countries, in desert and semi-arid shrublands, subtropical forests and high mountains. To improve the knowledge of the impacts of invasive plant species, we also advocate for more studies on species that are still locally rare and with restricted distribution, and on how they modify plant-soil-microbe interactions.

The information provided in this database is of interest for academic, management and policy-related purposes at the national, European and international scale. We acknowledge that our database may not encompass all relevant studies. The Web of Science has been the most widely used database for bibliometric analysis, offering more comprehensive coverage of older literature compared to Scopus. However, Scopus includes a larger list of journals than the Web of Science (Mongeon and Paul-Hus 2016). Further extensions of the database should also include a broader keywords string. For example, although restoration studies may not be explicitly designed to detect impacts, they can offer valuable insights into ecological differences between invaded plots before and after intervention in removal plots. Our database will have to be updated as new field studies on the ecological impacts of invasive species are published.

Acknowledgements

We thank T. Heger, S. Jelaska and an anonymous reviewer for valuable comments on a previous version of this paper. We thank IPBES for inspiring the construction of this database. José M. Espinar assisted on data extraction.

This research was supported by the 2017–2018 Belmont Forum–BiodivERsA International joint call project InvasiBES under the BiodivScen ERA-Net COFUND programme and with the following funding organisations: the Spanish Ministry of Science and Innovation (PCI2018-092939, PCI2018-092986, MCI/AEI/FEDER, RED2022-134338-T) and the Swiss National Science Foundation (31BD30-184114 and 31003A_179491); also by EXARBIN (RTI2018-093504-B-100), RADIOPOPO (PDI2021-122690OB-100); the European Union FPS COST Actions FP1403 NNEXT; and through the European Regional Development Fund (SUMHAL, LIFE-WATCH-2019-09-CSIC-4, POPE 2014-2020).

References

- Bacher S, Galil BS, Nuñez MA, Ansong M, Cassey P, Dehnen-Schmutz K, Fayvush G, Hiremath AJ, Ikegami M, Martinou AF, McDermott SM, Preda C, Vilà M, Weyl OLF, Fernandez RD, Ryan-Colton E (2023) Chapter 4: Impacts of invasive alien species on nature, nature's contributions to people, and good quality of life. In: Roy HE, Pauchard A, Stoett P, Renard Truong T (Eds) Thematic Assessment Report on Invasive Alien Species and Their Control of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. IPBES secretariat, Bonn. https://doi.org/10.5281/zenodo.7430731
- Blackburn TM, Essl F, Evans T, Hulme PE, Jeschke JM, Kühn I, Kumschick S, Marková Z, Mrugała A, Nentwig W, Pergl J, Pyšek P, Rabitsch W, Ricciardi A, Richardson DM, Sendek A, Vilà M, Wilson JRU, Winter M, Genovesi P, Bacher S (2014) A unified classification of alien species based on the magnitude of their environmental impacts. PLoS Biology 12(5): e1001850. https://doi.org/10.1371/journal.pbio.1001850
- Bradley BA, Laginhas BB, Whitlock R, Allen JM, Bates AE, Bernatchez G, Diez JM, Early R, Lenoir J, Vilà M, Sorte CJB (2019) Disentangling the abundance-impact relationship for invasive species. Proceedings of the National Academy of Sciences of the United States of America 116(20): 9919–9924. https://doi.org/10.1073/pnas.1818081116
- Cano-Barbacil C, Carrete M, Castro-Díez P, Delibes-Mateos M, Jaques JA, López-Darias M, Nogales M, Pino J, Ros M, Traveset A, Turon X, Vilà M, Altamirano M, Álvarez I, Arias A, Boix D, Cabido C, Cacabelos E, Cobo F, Cruz J, Cuesta JA, Dáder B, del Estal P, Gallardo B, Gómez Laporta M, González-Moreno P, Hernández JC, Jiménez-Alfaro B, Lázaro Lobo A, Leza M, Montserrat M, Oliva-Paterna FJ, Piñeiro L, Ponce C, Pons P, Rotchés-Ribalta R, Roura-Pascual N, Sánchez M, Trillo A, Viñuela E, García-Berthou E (2023) Identification of potential invasive alien species in Spain through horizon scanning. Journal of Environmental Management 345: e118696. https://doi.org/10.1016/j.jenvman.2023.118696

- Carboneras C, Genovesi P, Vilà M, Blackburn TM, Carrete M, Clavero M, D'hondt B, Orueta JF, Gallardo B, Geraldes P, González-Moreno P, Gregory RD, Nentwig W, Paquet JY, Pyšek P, Rabitsch W, Ramírez I, Scalera R, Tella JL, Walton P, Wynde R (2018) A prioritised list of invasive alien species to assist the effective implementation of EU legislation. Journal of Applied Ecology 55(2): 539–547. https://doi.org/10.1111/1365-2664.12997
- Carboni M, Guéguen M, Barros C, Georges D, Boulangeat I, Douzet R, Dullinger S, Klonner G, van Kleunen M, Essl F, Bossdorf O, Haeuser E, Talluto MV, Moser D, Block S, Conti L, Dullinger I, Münkemüller T, Thuiller W (2018) Simulating plant invasion dynamics in mountain ecosystems under global change scenarios. Global Change Biology 24(1): e289– e302. https://doi.org/10.1111/gcb.13879
- Castro-Díez P, Godoy O, Alonso A, Gallardo A, Saldaña A (2014) What explains variation in the impacts of exotic plant invasions on the nitrogen cycle? A meta-analysis. Ecology Letters 17(1): 1–12. https://doi.org/10.1111/ele.12197
- Castro-Díez P, Pauchard A, Traveset A, Vilà M (2016) Linking the impacts of plant invasion on community functional structure and ecosystem properties. Journal of Vegetation Science 27(6): 1233–1242. https://doi.org/10.1111/jvs.12429
- Castro-Díez P, Vaz AS, Silva JS, van Loo M, Alonso Á, Aponte C, Bayón Á, Bellingham PJ, Chiuffo MC, DiManno N, Julian K, Kandert S, La Porta N, Marchante H, Maule HG, Mayfield MM, Metcalfe D, Monteverdi MC, Núñez MA, Ostertag R, Parker IM, Peltzer DA, Potgieter LJ, Raymundo M, Rayome D, Reisman-Berman O, Richardson DM, Roos RE, Saldaña A, Shackleton RT, Torres A, Trudgen M, Urban J, Vicente JR, Vilà M, Ylioja T, Zenni RD, Godoy O (2019) Global effects of non-native tree species on multiple ecosystem services. Biological Reviews of the Cambridge Philosophical Society 94(4): 1477–1501. https://doi.org/10.1111/brv.12511
- Dawson W, Schrama M (2016) Identifying the role of soil microbes in plant invasions. Journal of Ecology 104: 1211–1218. https://doi.org/10.1111/1365-2745.12619
- European Union (2014) Regulation (EU) No 1143/2014 of the European Parliament and of the Council of 22 October 2014 on the prevention and management of the introduction and spread of invasive alien species. OJ L 317, 35–55.
- European Union (2017) Commission Implementing Regulation (EU) 2017/1263 of 12 July 2017 updating the list of invasive alien species of Union concern established by Implementing Regulation (EU) No 2016/1141 pursuant to Regulation (EU) No 1143/2014 of the European Parliament and of the Council. OJ L 182, 37–39.
- Gallardo B, Clavero M, Sánchez MI, Vilà M (2016) Global ecological impacts of invasive species in aquatic ecosystems. Global Change Biology 22(1): 151–163. https://doi.org/10.1111/gcb.13004
- Hulme PE, Pyšek P, Jarošík V, Pergl J, Schaffner U, Vilà M (2013) Bias and error in understanding plant invasion impacts. Trends in Ecology & Evolution 28(4): 212–218. https://doi. org/10.1016/j.tree.2012.10.010
- IPBES (2018) The IPBES regional assessment report on biodiversity and ecosystem services for Europe and Central Asia. In: Rounsevell M, Fischer M, Torre-Marin Rando A, Mader A (Eds) Secretariat of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services, Bonn, 892 pp. https://doi.org/10.5281/zenodo.3237429

- IPBES (2023) Summary for Policymakers of the Thematic Assessment Report on Invasive Alien Species and their Control of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. IPBES secretariat, Bonn. https://doi.org/10.5281/ zenodo.7430692
- Jeschke JM, Bacher S, Blackburn TM, Dick JTA, Essl F, Evans T, Gaertner M, Hulme PE, Kühn I, Mrugała A, Pergl J, Pyšek P, Rabitsch W, Ricciardi A, Richardson DM, Sendek A, Vilà M, Winter M, Kumschick S (2014) Defining the impact of non-native species. Conservation Biology 28(5): 1188–1194. https://doi.org/10.1111/cobi.12299
- Kueffer C, Daehler CC, Torres-Santana CW, Lavergne C, Meyer JY, Otto R, Silva L (2010) A global comparison of plant invasions on oceanic islands. Perspectives in Plant Ecology, Evolution and Systematics 12(2): 145–161. https://doi.org/10.1016/j.ppees.2009.06.002
- Kumschick S, Gaertner M, Vilà M, Essl F, Jeschke JM, Pysek P, Ricciardi A, Bacher S, Blackburn TM, Dick JTA, Evans T, Hulme PE, Kühn I, Mrugala A, Pergl J, Rabitsch W, Richardson DM, Sendek A, Winter M (2015) Ecological impacts of alien species: Quantification, scope, caveats, and recommendations. Bioscience 65(1): 55–63. https://doi.org/10.1093/ biosci/biu193
- Kuznetsova A, Brockhoff PB, Christensen RHB (2017) ImerTest package: Tests in linear mixed effects models. Journal of Statistical Software 82(13): 1–26. https://doi.org/10.18637/jss. v082.i13
- Lekberg Y, Bever JD, Bunn RA, Callaway RM, Hart MM, Kivlin SN, Klironomos J, Larkin BG, Maron JL, Reinhart KO, Remke M, van der Putten WH (2018) Relative importance of competition and plant–soil feedback, their synergy, context dependency and implications for coexistence. Ecology Letters 21(8): 1268–1281. https://doi.org/10.1111/ele.13093
- Lenzner B, Latombe G, Schertler A, Seebens H, Yang Q, Winter M, Weigelt P, van Kleunen M, Pyšek P, Pergl J, Kreft H, Dawson W, Dullinger S, Essl F (2022) Naturalized alien floras still carry the legacy of European colonialism. Nature Ecology & Evolution 6(11): 1723–1732. https://doi.org/10.1038/s41559-022-01865-1
- Lucy FE, Davis E, Anderson R, Booy O, Bradley K, Britton JR, Byrne C, Caffrey JM, Coughlan NE, Crane K, Cuthbert RN, Dick JTA, Dickey JWE, Fisher J, Gallagher C, Harrison S, Jebb M, Johnson M, Lawton C, Lyons D, Mackie T, Maggs C, Marnell F, McLoughlin T, Minchin D, Monaghan O, Montgomery I, Moore N, Morrison L, Muir R, Nelson B, Niven A, O'Flynn C, Osborne B, O'Riordan RM, Reid N, Roy H, Sheehan R, Stewart D, Sullivan M, Tierney P, Treacy P, Tricarico E, Trodd W (2020) Horizon scan of invasive alien species for the island of Ireland. Management of Biological Invasions : International Journal of Applied Research on Biological Invasions 11(2): 155–177. https://doi.org/10.3391/mbi.2020.11.2.01
- Lüdecke D (2018) ggeffects: Tidy data frames of marginal effects from regression models. Journal of Open Source Software 3(26): e772. https://doi.org/10.21105/joss.00772
- Marchante H, Freitas H, Hoffmann JH (2011) The potential role of seed banks in the recovery of dune ecosystems after removal of invasive plant species. Applied Vegetation Science 14(1): 107–119. https://doi.org/10.1111/j.1654-109X.2010.01099.x
- Mongeon P, Paul-Hus A (2016) The journal coverage of Web of Science and Scopus: A comparative analysis. Scientometrics 106(1): 213–228. https://doi.org/10.1007/s11192-015-1765-5

- Morente-López J, Arjona Y, Salas-Pascual M, Reyes-Betancort JA, del Arco-Aguilar MJ, Emerson BC, García-Gallo A, Jay-García LS, Naranjo-Cigala A, Patiño J (2023) Biogeographic origins and drivers of alien plant invasions in the Canary Islands. Journal of Biogeography 50(3): 576–590. https://doi.org/10.1111/jbi.14556
- Pergl J, Pyšek P, Bacher S, Essl F, Genovesi P, Harrower CA, Hulme PE, Jeschke JM, Kenis M, Kühn I, Perglová I, Rabitsch W, Roques A, Roy DB, Roy HE, Vilà M, Winter M, Nentwig W (2017) Troubling travellers: Are ecologically harmful alien species associated with particular introduction pathways? NeoBiota 32: 1–20. https://doi.org/10.3897/neo-biota.32.10199
- Pyšek P, Jarošík V, Hulme PE, Pergl J, Hejda M, Schaffner U, Vilà M (2012) A global assessment of invasive plant impacts on resident species, communities and ecosystems: The interaction of impact measures, invading species' traits and environment. Global Change Biology 18(5): 1725–1737. https://doi.org/10.1111/j.1365-2486.2011.02636.x
- R Core Team (2022) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna.
- Romero-Blanco A, Castro-Díez P, Lázaro-Lobo A, Molina-Venegas R, Cruces P, Pyšek P (2023) Searching for predictors of the variability of impacts caused by non-native trees on regulating ecosystem services worldwide. The Science of the Total Environment 877: e162961. https://doi.org/10.1016/j.scitotenv.2023.162961
- Russell L (2018) Emmeans: estimated marginal means, aka least-squares means. R package version 2: 1–1.
- Sapsford SJ, Brandt AJ, Davis KT, Peralta G, Dickie IA, Gibson II RD, Green JL, Hulme PE, Nuñez MA, Orwin KH, Pauchard A, Wardle DA, Peltzer DA (2020) Towards a framework for understanding the context dependence of impacts of non-native tree species. Functional Ecology 34(5): 944–955. https://doi.org/10.1111/1365-2435.13544
- Silva L, Land EO, Luis J, Luengo R, Borges P, Oliveira P, Jardim R (2008) Invasive alien species in Macaronesia. In: Silva L, Land EO, Luis J (Eds) Invasive Terrestrial Flora & Fauna of Macaronesia. TOP 100 in Azores, Madeira and Canaries, Ponta Delgada, 159–165.
- Simberloff D, Martin JL, Genovesi P, Maris V, Wardle DA, Aronson J, Courchamp F, Galil B, García-Berthou E, Pascal M, Pyšek P, Sousa R, Tabacchi E, Vilà M (2013) Impacts of biological invasions: What's what and the way forward. Trends in Ecology & Evolution 28(1): 58–66. https://doi.org/10.1016/j.tree.2012.07.013
- Strayer DL (2012) Eight questions about invasions and ecosystem functioning. Ecology Letters 15(10): 1199–1210. https://doi.org/10.1111/j.1461-0248.2012.01817.x
- Vilà M, Hulme PE (2017) Impact of Biological Invasions on Ecosystem Services. Springer International Publishing, Cham. https://doi.org/10.1007/978-3-319-45121-3
- Vilà M, Espinar JL, Hejda M, Hulme PE, Jarošík V, Maron JL, Pergl J, Schaffner U, Sun Y, Pyšek P (2011) Ecological impacts of invasive alien plants: A meta-analysis of their effects on species, communities and ecosystems. Ecology Letters 14(7): 702–708. https:// doi.org/10.1111/j.1461-0248.2011.01628.x
- Vilà M, Gallardo B, Preda C, García-Berthou E, Essl F, Kenis M, Roy HE, González-Moreno P (2019) A review of impact assessment protocols of non-native plants. Biological Invasions 21(3): 709–723. https://doi.org/10.1007/s10530-018-1872-3

- Vimercati G, Kumschick S, Probert AF, Volery L, Bacher S (2020) The importance of assessing positive and beneficial impacts of alien species. NeoBiota 62: 525–545. https://doi. org/10.3897/neobiota.62.52793
- Vitousek PM, Walker LR (1989) Biological invasion by Myrica Faya in Hawai'i: Plant demography, nitrogen fixation, ecosystem effects. Ecological Monographs 59(3): 247–265. https://doi.org/10.2307/1942601
- Volery L, Jatavallabhula D, Scillitani L, Bertolino S, Bacher S (2021) Ranking alien species based on their risks of causing environmental impacts: A global assessment of alien ungulates. Global Change Biology 27(5): 1003–1016. https://doi.org/10.1111/gcb.15467

Supplementary material I

Supplementary information

Authors: Montserrat Vilà, Alejandro Trillo, Pilar Castro-Díez, Belinda Gallardo, Sven Bacher

Data type: docx

- Explanation note: table S1. Definitions of the response variables used to classify impact types on native species (i.e. individuals of the same taxon) and communities (i.e. individuals of several species at a site). table S2. Publication level information in PLAN-TIMPACTSEUROPE_publicationLevel. xlsx. The PLANTIMPACTSEUROPE database can be accessed at https://figshare.com/s/0a890d22bf5632fe5cb5.
 table S3. Invasive plant information and field studies testing for impacts in PLAN-TIMPACTSEUROPE_impactsDatabase. xlsx. The PLANTIMPACTSEUROPE database can be accessed at https://figshare.com/s/0a890d22bf5632fe5cb5.
 Storage location and medium: The PLANTIMPACTSEUROPE database can be accessed at https://figshare.com/s/0a890d22bf5632fe5cb5.
 Storage location and medium: The PLANTIMPACTSEUROPE database can be accessed at https://figshare.com/s/0a890d22bf5632fe5cb5.
 Storage location and medium: The PLANTIMPACTSEUROPE database can be accessed at https://figshare.com/s/0a890d22bf5632fe5cb5.
 Storage location and medium: The PLANTIMPACTSEUROPE database can be accessed at https://figshare.com/s/0a890d22bf5632fe5cb5.
 (1) PLANTIMPACT-SEUROPE_publicationLevel. xlsx: 266 publications with indication of countries, habitats and study locations, 312 entries (rows excluding the header), 8 columns, 59 KB. (2) PLANTIMPACTSEUROPE_impactsDatabase. xlsx: 4259 impacts (rows excluding the header), 16 columns, 348 KB.
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.112368.suppl1

RESEARCH ARTICLE



Recent invasion of Ponto-Caspian amphipods in the Masurian Lakeland associated with human leisure activities

Krzysztof Podwysocki¹, Andrea Desiderato¹, Tomasz Mamos¹, Tomasz Rewicz¹, Michał Grabowski¹, Alicja Konopacka¹, Karolina Bącela-Spychalska¹

I Department of Invertebrate Zoology and Hydrobiology, University of Lodz, Lodz, Poland

Corresponding author: Karolina Bącela-Spychalska (karolina.bacela@biol.uni.lodz.pl)

Academic editor: Eric Larson | Received 8 July 2023 | Accepted 14 December 2023 | Published 29 January 2024

Citation: Podwysocki K, Desiderato A, Mamos T, Rewicz T, Grabowski M, Konopacka A, Bącela-Spychalska K (2024) Recent invasion of Ponto-Caspian amphipods in the Masurian Lakeland associated with human leisure activities. NeoBiota 90: 161–192. https://doi.org/10.3897/neobiota.90.109221

Abstract

Non-indigenous species (NIS) contribute to the decrease of native species' diversity on a local and global scale. One of Europe's most significant donors of freshwater invasions is the Ponto-Caspian Region. Following the construction of artificial canals connecting isolated waterbodies and the resulting heavy boat traffic, the Ponto-Caspian Amphipoda started to spread in Europe. Four amphipod species: *Dikerogammarus villosus, Pontogammarus robustoides* and *Chaetogammarus ischnus* have invaded the Masurian Lakeland (North-eastern Poland). Based on literature and our data, we studied their distribution in 22 lakes in the region during the years 2001–2016. We analysed their distribution against several water quality parameters and levels of anthropogenic pressure. Our results also present the first records of two new invaders, *D. villosus* and *C. ischnus*, in the studied area. We show that the relative abundance and frequency of these two species rapidly increase and, simultaneously, the populations of the earlier invaders, i.e. *D. haemobaphes* and *P. robustoides*, decrease. The native species – *Gammarus lacustris* – seems to be negatively affected by NIS richness, as well as by the proximity of towns. The spread of NIS in the lakes appears to be facilitated by boating and the lower complexity of the shoreline. Our study shows how anthropogenic pressure, especially tourism, can facilitate bioinvasion, jeopardising native biodiversity unless appropriate regulations are implemented.

Keywords

assemblage succession, biological invasions, lakes, propagule pressure, recreational boating, time series, tourist pressure

Copyright Krzysztof Podwysocki et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

Biological invasions are perceived as the second strongest threat to biodiversity on a local and global scale, right after habitat degradation (e.g. CBD (2000); Sala et al. (2000); Dudgeon et al. (2006); Kettunen et al. (2009); Strayer and Dudgeon (2010); Lambertini et al. (2011); Mačić et al. (2018); Lipták et al. (2019); Igbal et al. (2021); Yang et al. (2021); Vantarová et al. (2023)). Many non-indigenous species (NIS) cause declines in abundance and diversity of native species, which is particularly apparent in Europe and the USA (e.g. Pinkster et al. (1992); Dick and Platvoet (1996, 2000); Ricciardi and MacIsaac (2000); Ricciardi (2006); Bellard et al. (2016); Panlasigui et al. (2018); Albano et al. (2021); Haubrock et al. (2021); Yang et al. (2021)) and is more prominent in freshwater ecosystems than in marine and terrestrial ones (Strayer and Dudgeon 2010). Many studies show high economic costs incurred by biological invasions on a global scale (Pyšek and Richardson 2010; Cuthbert et al. 2021a, b; Kouba et al. 2022). The average annual costs of preventing biological invasions and reversing their effects globally reach \$76 billion (Bradshaw et al. 2016); however, the costs of prevention of invasion are much lower than post-invasion management (Cuthbert et al. 2021a). Globally, the economic costs of aquatic bioinvasions have been estimated at \$23 billion (Cuthbert et al. 2021b). The costs of amphipod invasions constitute a small part of the global costs of aquatic crustacean invasions (\$180,000 out of an estimated \$271 million); however, these costs are underestimated (Kouba et al. 2022).

Even though surface freshwaters represent only 0.01% of the Earth's water resources and constitute 0.80% of the Earth's surface, they are inhabited by ca. 6% of the world's species (Dudgeon et al. 2006; Strayer and Dudgeon 2010). Therefore, freshwater ecosystems are precious from environmental, economic, sanitary, cultural and scientific perspectives and also constitute a valuable spot for tourism (Dudgeon et al. 2006; Hall and Härkönen 2006). Unfortunately, these ecosystems are in crisis, as indicated by stronger biodiversity loss than in terrestrial ecosystems (Dudgeon et al. 2006). According to the Water Framework Directive (European Environment Agency 2000), every waterbody in the EU should have achieved a high or at least good ecological and chemical status by 2015. However, the latest reports indicate that only 40% of such waters have achieved a satisfactory, healthy status (European Environment Agency 2018). Land use and agriculture are amongst the most important factors in aquatic ecosystems' declining conditions globally (Foley et al. 2005; Feld et al. 2016). Thus, although freshwater ecosystems constitute only a tiny fraction of the Earth's surface, high anthropogenic pressure results in a more pronounced negative impact of invaders on native species than in marine ecosystems (Ricciardi and Kipp 2008).

One of the richest European sources of species invading inland waters is the Ponto-Caspian Region (Ricciardi and MacIsaac 2000; Bij de Vaate et al. 2002; Galil et al. 2008; Panov et al. 2009; Copilaș-Ciocianu et al. 2023a). This region covers the coastal area of the Caspian, Black, Aral and Azov Seas, with their brackish limans and deltas of rivers discharging into these seas (Jażdżewski 1980). The Ponto-Caspian basin constitutes a hotspot of crustacean diversity, particularly in the case of amphipod crustaceans (Cristescu and Hebert 2005; Väinölä et al. 2008; Copilas-Ciocianu and Sidorov 2022; Copilaș-Ciocianu et al. 2022). Ponto-Caspian amphipods comprise around 10% of European freshwater invasive species (Pöckl et al. 2011). One of the main significant causes fuelling the bioinvasions of Ponto-Caspian species is the construction of canals that connect previously isolated watersheds (e.g. Jażdżewski (1980); Bij de Vaate et al. (2002); Nehring (2005); Galil et al. (2008); Arbačiauskas et al. (2010); Minchin et al. (2019); Jażdżewska et al. (2020)). Another important factor is translocations of species in ballast waters (Jażdżewski 1980; Pinkster et al. 1992; Bij de Vaate et al. 2002; Zhulidov et al. 2018). However, a more important vector of bioinvasions in freshwater ecosystems is transporting on biofouled hulls, filters and other submerged parts of vessels (Nehring 2005; Hewitt et al. 2009; Bącela-Spychalska et al. 2013; Anderson et al. 2014, 2015; De Ventura et al. 2016; Rewicz et al. 2017; Rodríguez-Rey et al. 2021). Biofouling of vessels by species resistant to desiccation enables their subsequent overland transport and the colonisation of isolated waterbodies (Bacela-Spychalska et al. 2013; Rachalewski et al. 2013; De Ventura et al. 2016). Fishing and diving equipment can also be an effective vector of invasions (Bacela-Spychalska et al. 2013; Anderson et al. 2014; Smith et al. 2020). Moreover, many species are also intentionally introduced into freshwater ecosystems (Grigorovich et al. 2002; Nehring 2005).

Seven species of Ponto-Caspian gammarids (Amphipoda, Gammaroidea) have already been recorded from Polish freshwaters: Chaetogammarus ischnus (Stebbing, 1899), Dikerogammarus haemobaphes (Eichwald, 1841), Dikerogammarus villosus (Sowinsky, 1894), Obesogammarus crassus (G.O. Sars, 1894), Pontogammarus robustoides (Sars, 1894), Spirogammarus major (Cărăușu, 1943) (former European population of Echinogammarus trichiatus) and Chelicorophium curvispinum (G.O. Sars, 1895) (Konopacka 1998; Gruszka 1999; Jażdżewski and Konopacka 2000; Konopacka and Jażdżewski 2002; Jażdżewski et al. 2005; Grabowski et al. 2007; Rachalewski et al. 2013; Copilaș-Ciocianu et al. 2023b). These species are already widely distributed in European inland waters, where they arrived through well-defined migration corridors: northern, central and southern (Bij de Vaate et al. 2002; Panov et al. 2009). Not only have they colonised the major rivers and canals constituting the invasions corridors, but also spread to the watersheds of these rivers, as well as many European lakes, for example, the Alpine Lakes (Rewicz et al. 2017) and the Great Masurian Lakes in Poland (Jażdżewski 2003; Jażdżewska and Jażdżewski 2008). An extensive up-to-date description of the distribution of alien freshwater amphipods in Europe can be found in Copilaș-Ciocianu et al. (2023a). As the dynamics of invasion in terms of species and ecosystem vulnerability varies and the impact of NIS depends on their invasion process (i.e. propagule pressure, species interactions), there is a constant need for monitoring and estimating trends and threats regarding invasions. The impact of invasive species on aquatic ecosystems is profound (Kurashov et al. 2012). Their introduction may lead to drastic changes in the macroinvertebrate community structure and affect the functioning of whole ecosystems (Jones et al. 1994; Jones et al. 1997; Lambertini et al. 2011). NIS can modify habitats as well as food chains and contribute to changes in

energy flows – benthic communities can be transformed from being energy suppliers to upper trophic levels becoming major consumers of ecosystem energy (Nalepa et al. 2009; Kurashov et al. 2012).

Lakes seem to be particularly susceptible to biological invasions, as many of them are under high tourist pressure, resulting in a higher probability of alien species introduction, even if the lakes are not directly connected with the invasion corridor (Bacela-Spychalska et al. 2013; Bacela-Spychalska 2016; De Ventura et al. 2016; Rewicz et al. 2017). One such region is the Masurian Lakeland. It is the most popular area for yachting in Poland and one of central Europe's main inland yachting regions. The region is extensively used for associated recreational activities, particularly angling and camping (Kistowski and Śleszyński 2010; Ulikowski et al. 2021). Unfortunately, the level of knowledge about the risks of spreading invasive Amphipoda in this region is poor and out of date (Jażdżewski 2003; Jażdżewska and Jażdżewski 2008). Previous studies were based on sampling from only a few lakes, provided mainly presence/ absence data and predated the effect of increased recreational pressure. Knowledge about the role of tourism, shipping and other factors in biological invasions in the Masurian Lakeland is poor and demands study. Given the significance of these factors in other regions, it is likely that their influence in the Masurian Lakeland is also considerable. The intensity of shipping and, therefore, its effect on biological invasions will increase with time (Sardain et al. 2019). Thus, it is crucial to understand these mechanisms in the study area. We also do not know how the invasion of amphipods affected native amphipods in the region. With regards to the faunistic data about the native amphipod species in the Lakeland, Jażdżewski and Konopacka (1995) mention two widely distributed lacustrine species, namely Gammarus lacustris G.O. Sars, 1863 and Pallasiola quadrispinosa (G.O. Sars, 1867). However, these data are old and require updating.

The aims of our study were: i) to update the knowledge on the distribution and expansion of the Ponto-Caspian amphipod fauna in the Masurian Lakeland; ii) to assess the distribution of native vs. invasive Ponto-Caspian amphipods in the context of biotic and abiotic characteristics of the lakes and anthropogenic pressure in this region, using both historical and newly-obtained data. Based on observed trends in other regions (e.g. Dick and Platvoet (2000); Grabowski et al. (2006); Van der Velde et al. (2009); Meßner and Zettler (2021)), we assumed that some invasive amphipods are replaced by stronger competitors and that native species are not able to co-exist with the invasive species. We hypothesise that high tourist pressure contributes to the dispersion of invasive amphipods, while the occurrence of the native species is linked to isolated lakes.

We tracked the distribution of invasive Amphipoda in the Masurian Lakeland since 2001, based on literature and our data. To explore the relationship between the structure of amphipod assemblages and lake characteristics, including human tourist pressure in the years 2014 and 2016, we collected data on the relative abundance of amphipods, measured basic water parameters, implemented hydromorphological data and estimated the tourist pressure.

Materials and methods

Study area

The Masurian Lakeland (Pojezierze Mazurskie in Polish) is a lake district (macroregion) in North-eastern Poland with a surface area of 52,000 km² including seven mesoregions, amongst others, the Land of the Great Masurian Lakes (Kraina Wielkich Jezior Mazurskich in Polish) and the Ełckie Lakeland (Pojezierze Ełckie in Polish) (Kondracki 2002). The landscape was formed between 16,000 and 11,000 BP (at the end of the last glaciation) and is characterised by strong latitude differentiation, dominantly with moraine hills (Hillbricht-Ilkowska et al. 2000; Ulikowski et al. 2021) and with glacial tills as a dominant component of the soil substratum (Hillbricht-Ilkowska et al. 2000). The lakes are mainly surrounded by a mosaic of agricultural areas and forests giving similar input of allochthonous organic and mineral matter to each lake (Chróst and Siuda 2006; Ejsmont-Karabin et al. 2020). Most lakes of this region are dimictic with summer thermal stratification (Ulikowski et al. 2021). They are connected with main European watersheds via artificial canals and small rivers: the River Pisa (flowing into the River Narew and then into the River Vistula) and the River Wegorapa (flowing into the River Pregolya and then into the Vistula Lagoon) (Bajkiewicz-Grabowska 2008; Jażdżewska and Jażdżewski 2008; Ulikowski et al. 2021). This connectivity increases the probability of invasive amphipods spreading in the region. For this study, we selected lakes with historical faunistic data, based on Jażdżewski and Konopacka (1995), as well as along a gradient of tourist pressure, including more natural and isolated lakes. We also selected sampling points on the rivers, i.e. the River Wegorapa, the River Pisa and the River Narew, which connect the Masurian Lakeland with major rivers, for example, the River Vistula and the River Neman (Fig. 1; see also Suppl. material 1).

Sampling and data collection

Our dataset consists of two types of data: (*i*) published, including the years between 2001 and 2007 (Jażdżewski 2003; Jażdżewska and Jażdżewski 2008) and (*ii*) new data coming from field surveys in 2008, 2009, 2014 and 2016. Additionally, to facilitate the monitoring of the amphipod expansion and to model the distribution of native *Gammarus lacustris*, we incorporated records from several lakes and the River Narew, which are situated outside of the study area (see Suppl. material 1). The studies that were conducted between 2001 and 2009 only have a qualitative character (i.e. presence/absence of amphipod species), while for 2014 and 2016, the species abundances are available. Generally, sampling was done through "kick-sampling" with a benthic hand-net with a mesh size of 0.5 mm, used for 45 min at each station, performed by two people with equal effort, from all available littoral habitats (sand, mud, gravel, stones and submerged macrophytes) at depths from 0.05 to 0.5 m. Such a semi-quantitative method gives reliable and comparable results for all sampling points and all study years/periods (Jażdżewski et al. 2002; Grabowski et al. 2006). The amphipods



Figure 1. The sites in the Masurian Lakeland. Sites were divided into previously unpublished (records of this study) and published (Jażdżewski 2003; Jażdżewska and Jażdżewski 2008). Mesoregions are delimited according to Kondracki (2002). The two-letter acronyms for particular lakes were used in further Figures and Suppl. material 1.

were preserved in 96% ethanol and then identified in the laboratory to the species level, based on the available literature (Mordukhai-Boltovskoi 1964; Eggers and Martens 2001). This collection and preservation protocol was used at all studied sites and in all study years.

To detect the potential role of biotic and abiotic factors, as well as human pressure on the presence of invasive amphipods in the lakes sampled in 2014 and 2016, we used topological and anthropogenic variables, such as the surface-volume ratio or the distance from town. As a proxy of the level of anthropogenic pressure, we used the water quality status (water QS) from Soszka et al. (2016). This index categorises the waterbodies into six water quality categories (ranging from excellent – class I, to very poor – class VI), based on species assemblages and chemical and physical parameters of water according to the Water Framework Directive (European Environment Agency 2000). We presume that lower values of this variable (lower water class), indicating increased species diversity and reduced levels of nutrients and heavy metals in the water (better water quality), correspond to lower levels of anthropogenic pressure on the lake (European Environment Agency 2000; Sánchez et al. 2007; Lobato et al. 2015). Environmental heterogeneity creates more niches that can be occupied by co-occurring species (Chesson 2000). Thus, we used two indices: shoreline development (shoreline length to surface area ratio) from mojemazury.pl and surface area to volume ratio (A/V ratio) from Soszka et al. (2016). The shoreline development index is the ratio of the actual shoreline length of a lake to the circumference of a perfectly circular lake with the same area (Aronow 1982). High values indicate a more complex shoreline, retaining a higher load of nutrients from land (Cole 1975) and providing more niches for the biota (Chesson 2000). The surface area to volume ratio combines information about the depth and size of the lake and can be positively correlated with the productivity of the lake (Fee 1979). Smaller waterbodies (lower A/V ratio) may play the role of refugia for native species (Grabowski et al. 2009). The density of boats (i.e. the number of boats divided by the lake surface in ha), was obtained as the maximum possible number of moored boats in marinas (Johnson and Padilla 1996; Vander Zanden and Olden 2008; Ros et al. 2013). We assumed that the higher the density of boats in the lakes, the higher the tourist pressure and the higher the probability of transport of invasive species by vessels (Johnson and Padilla 1996; Vander Zanden and Olden 2008; Bacela-Spychalska et al. 2013; Ros et al. 2013). The maximum capacity of marinas was obtained from websites: mazury24.eu and skorupki.mazury.info.pl. Tourist infrastructure is mainly localised in urban areas (Kulczyk et al. 2016). Thus, we used the distance between the sampling point and towns (i.e. centroid) as an estimation of anthropogenic pressure. Moreover, land use in the vicinity of water-bodies can impact the temporal variations in amphipod assemblages (Cereghetti 2023). The distance was measured as a linear distance in km from the centroid of the closest town to the sampling point using QGIS software. Towns were designated according to the ESRI shapefile "UIA World Countries Boundaries", available at: https://hub.arcgis.com/datasets/UIA::uia-world-countries-boundaries. All spatial analyses and their visualisation were conducted using QGIS 3.10.13 (QGIS Development Team 2020).

Data analysis

Using all unpublished records since 2008 from the lakes and the rivers, including sites outside the study area (see Suppl. material 1), we modelled the presence of the only native gammarid (i.e. *Gammarus lacustris*) according to the number of NIS and the relative distance of each sampling site from town. We included this variable as a proxy of the anthropogenic propagule pressure (i.e. the introduction of NIS by human activities) of NIS at each site (i.e. inversely correlated). We used generalised linear mixed models (GLMMs) to include the random variable of the sampling year. Given the presence/absence nature of the data, we used a Bernoulli distribution fitted with glmmTMB (link = logit) with the homonymous package (Brooks et al. 2017). The possible inclusion of the interaction between NIS richness (i.e. number of species) and the distance from the closest town was also tested using the Akaike Information Criterion (AIC; Bozdogan (1987)). After fitting the model, we validated it by simulating its residuals using the package DHARMa (Hartig 2022). We also confirmed the absence of spatial autocorrelation of the residuals using the Spatial Autocorrelation function of the DHARMa package.

Using samples collected in 2014 and 2016, we first explored the variability of the environmental parameters of the sites and lakes, grouping them according to their geographical position and connectivity (i.e. I: northern, II: southern, III: eastern; Fig. 2B, see also Suppl. material 1). We hypothesised that nearby and interconnected lakes would exhibit comparable gammarid assemblages. This assumption is supported by findings from the Great Lakes in the USA, where the likelihood of species invasion was found to be the highest near the mouth of canals connecting the lakes (Grigorovich et al. 2005). To explore and visualise the environmental variability of the study area, we used a principal component analysis (PCA) with standardised values with prcomp of the package vegan (Oksanen et al. 2022). We analysed the gammarid assemblage using a permutational multivariate analysis of the covariance (PERMANCOVA) with an orthogonal design with two fixed factors (i.e. lake groups with three levels – I, II, III; time with two levels - 2014 and 2016) and five covariates: water QS, A/V ratio, shoreline development, density of boats and distance from the town. To control the possible sampling differences (i.e. being semi-quantitative), Hellinger distances were used to compare the abundances of the different species. To account for the excess of zero values, a dummy variable of 0.0001 was added to the whole dataset. We used first adonis2 of the package vegan with 9999 permutations and pairwise.adonis of the package pairwiseAdonis, with Holm correction and 9999 permutations, for the post hoc analysis between levels of the significant factors (Martinez Arbizu 2020). To visualise and corroborate the results of the PERMANCOVA, we finally used a constrained ordination using distance-based redundancy analysis (dbRDA), based on Legendre and Anderson (1999), with capscale (package vegan) and Hellinger distances, as for Permancova, including the covariates of the PERMANCOVA as constraining variables. All the analyses were performed in the R environment 4.3.0 version (R Core Team 2023).

Results

Temporal and spatial distribution of invasive species

We recorded four invasive gammarid species from 12 lakes and the Rivers Węgorapa and Pisa and one native species (*Gammarus lacustris*) from 16 lakes (Fig. 2A, Suppl. material 1). The first recorded invasive species was *Dikerogammarus haemobaphes* found in 2001 (Jażdżewski 2003) and the second was *Pontogammarus robustoides*, which was first observed in 2007 (Jażdżewska and Jażdżewski 2008). The spread of invasive species can be observed over time (Fig. 2B). Between 2014 and 2016, *D. haemobaphes* spread to one more lake and is observed now in nine of them. *Pontogammarus robustoides* did not colonise new lakes in 2016, compared to 2014. In 2014, we noticed the first appearance of the other two invaders: *C. ischnus* and *D. villosus* (Fig. 2A). The previous species was found in two lakes in 2014 and expanded to five further lakes in 2016, while the latter one was already found in five lakes in 2014 and expanded to two further lakes in 2016 (Fig. 2A). Although *Chaetogammarus ischnus* was recorded in the River Pisa in 2014 and 2016, *D. villosus* was not found in any of the studied rivers



Figure 2. A the distribution of invasive and native amphipod species in studied lakes since 2001, based on published and new data (locality codes according to Suppl. material 1). Table at each lake showing the assemblage (colours in rectangles according to different species, see legend) variation in time (symbols for sampling years: 1 - 2001; 2 - 2002; 3 - 2007; 4 - 2008; 5 - 2009; 6 - 2014; 7 - 2016). Only years of samplings from each lake and river are shown. Colourless rectangles indicate that no amphipods were recorded during the sampling. The dashed black line indicates country borders; the dashed red line indicates the Masurian tourist boat route. Black lines delimit mesoregions according to Kondracki (2002) **B** the assemblage composition of the amphipod fauna in studied lakes in the years 2014 and 2016 (locality codes according to Suppl. material 1). Pie charts show the relative abundances of each species. An empty circle means no amphipods were recorded. Black lines delimit mesoregions according to Kondracki (2002). Coloured dotted lines around the pie charts correspond to the lake groups: orange – I, green – II, blue – III.

(Fig. 2A) The relative abundance of new invaders (*D. villosus* and *C. ischnus*) increased with time, while it decreased for *D. haemobaphes* and *P. robustoides* (Fig. 2B). In Lake Nidzkie, we did not record any amphipod species (Fig. 2A, B).

The modelled occurrence of native Gammarus lacustris

Generally, the native species – *Gammarus lacustris* – was not found in lakes inhabited by invasive species, apart from Lake Dobskie, where the native and invasive gammarids cooccurred in 2014 with a low number of *G. lacustris* (two individuals vs. 194 individuals of invasive species) (Fig. 2A, B; Suppl. material 1). The GLMM for the presence of *G. lacustris* showed the significant negative effect of NIS richness (p-value = 0.002) and the positive effect of the distance from town (p-value = 0.024), but not their interaction (Fig. 3). The inclusion of the year as a random effect barely increased the R² (Marginal 0.733 – Conditional 0.808), supporting the effectiveness in sampling efforts (Suppl. materials 2, 4).

Environmental factors and amphipod assemblage

The first three components of the PCA explained 85.5% of the variance amongst the environmental variables (Fig. 4A, B). According to PC1 and PC3 (~ 57% variance explained), the lakes further from the tourist route (i.e. group III) are, indeed, characterised by a lower number of boats, higher complexity of the shore and a greater distance from town. The PC2 was more related to the water quality status (water class) and the surface-volume ratio showing a general trend of better water quality (lower class of water quality status) and deeper waters for group I (highest class of water status – lowest water quality for group II). The PERMANCOVA results showed significant effects (p-values < 0.05) of shoreline development (F = 22.096, p < 0.001), the number of boats (F = 10.788, p < 0.001) and water quality status – water class (F = 3.794, p = 0.035) on the assemblage of amphipods (Suppl. material 3). Even though the relative abundance of species changed with time, i.e. increased in *D. villosus* and *C. ischnus* and decreased



Figure 3. The predicted probability of occurrence of *G. lacustris* dependent on the richness of NIS (A) and the distance of the sampling point from town (B). The grey area delimits the 95% confidence intervals.



Figure 4. Biplots displaying the first three axes of the PCA of the environmental variables of the lakes sampled in 2014 and 2016 (**A** PC1-2 **B** PC1-3). The colours refer to the different lake groups: orange circles (I), green triangles (II) and blue squares (III). The lengths of the arrows are proportional to the loading of each variable, dashed lines = 0. The acronyms of lakes are according to Fig. 1 and Suppl. material 1.

in *D. haemobaphes*, *P. robustoides* and *G. lacustris*, the time factor was not significant. The differences in amphipod assemblages between lake groups (determined, based on the geographical position and interconnections between the lakes) were marginally significant, i.e. F = 2.680, p = 0.057) and the post hoc analysis showed a significant difference (p.adjusted < 0.001) between the group III (i.e. eastern group) and the others, but not between the first two (p.adjusted > 0.4).

The first two axes of the dbRDA fitted 90.1% of 52.1% of the total variation explained (Fig. 5). The presence of the native *G. lacustris* appeared more correlated to lakes with more complex shorelines. The occurrence of *D. villosus* was mainly explained by the increasing number of boats and proximity to town. The other three species (i.e. *P. robustoides*, *D. haemobaphes* and *C. ischnus*) seemed to be related to simpler shorelines and average values for the other variables, which was generally the opposite to *D. villosus*.



Figure 5. Canonical analysis of principal coordinates (CAPSCALE) derived from the Bray-Curtis dissimilarities of the gammarid assemblages and the environmental variables of the studied lakes in the years 2014 and 2016. The colours of the dots refer to the different lake groups: orange circles (I), green circles (II) and blue circles (III).

Discussion

Our study shows that, between 2001 and 2016, the number of invasive amphipod species in the study area increased drastically from one (*D. haemobaphes*) to four (three more species recorded: *D. villosus, P. robustoides, C. ischnus*). Simultaneously, a continuous decrease in the occurrence of native *Gammarus lacustris* was recorded. Our study reveals that the presence of NIS in lakes is primarily facilitated by three key factors: recreational boating activities, proximity to urban areas and simplified lake shorelines.

Distribution of Gammarus lacustris

According to our results, the presence of more than one NIS significantly affects the presence of the native G. lacustris, bringing the probability of its presence almost to zero already with three NIS (Fig. 3A). The species disappeared several years after the expansion of invasive amphipod species in several lakes (Fig. 2A, Suppl. material 1). For instance, the species was widely distributed until the last record in 2001 in Lake Kisajno (Jażdżewski 2003), in 2007 in Lake Tałty (Jażdżewska and Jażdżewski 2008), in 2008 in Lake Niegocin and in 2009 in Lake Śniardwy. Older data mention the presence of Gammarus lacustris in Lake Mamry (Jażdżewski 1975). In these lakes, the disappearance of G. lacustris coincided with the invasion of alien species. In 2014, G. lacustris was co-occurring with invasive species in only one lake (Lake Dobskie). One potential explanation could be the limited tourist activity in Lake Dobskie, as well as low species introduction probabilities, resulting from the absence of direct connections between this lake and other lakes situated along the Masurian tourist routes. Moreover, in 2014, the invasion of C. ischnus and P. robustoides in Lake Dobskie was still in its early stage. However, in 2014 the abundance of G. lacustris in this lake was very low and we did not record this species in 2016. Additionally, in 2002, we recorded the species co-occurring with D. haemobaphes in Lake Mikołajskie, but the presence of G. lacustris in this lake in subsequent years is unknown. In general, most of the records of G. lacustris in the Masurian Lakeland come from isolated lakes where invasive amphipods did not spread. In 2016, we found this species only in four isolated lakes, i.e. Dejguny, Ełckie, Łaśmiady and Łaźno (Fig. 2A; Suppl. material 1).

These four lakes (three of them in the eastern group of lakes) are characterised by low tourist pressure (low number of boats, long distance from the tourist routes) (Fig. 5). The low level of tourist pressure in these lakes and lack of direct connections with the Great Masurian Lakes (central part of the Masurian Lakeland), where all the invasive amphipods are present, may create a refuge for native species. Furthermore, we found that *G. lacustris* is associated with lakes distanced from towns (Figs 3B, 5). The proximity of the lakes to the urban areas results in their pollution and declining quality of water (Mishra et al. 2023). Although *G. lacustris* has a broad tolerance to environmental factors (Matafonov and Bazova 2014), its populations decline in polluted water, for example, with high acidity (Okland 1969) and pesticides (Gerhardt et al. 2011). Hence, it can be anticipated that *G. lacustris* will primarily be distributed in lakes with low anthropogenic pressure. Our results of CAPSCALE analysis show that higher classes of water quality status (lower water quality) characterise mainly lakes inhabited by *C. ischnus*, *D. haemobaphes* and *P. robustoides* (e.g. Lake Śniardwy, Lake Roś, Lake Święcajty), where we did not record *G. lacustris* (apart from Lake Śniardwy in 2009) (Fig. 5).

Instead, we recorded *G. lacustris* in lakes characterised by a high level of shoreline development. Lakes with higher shoreline complexity may provide higher habitat diversity, resulting in lower competition rates between species on environmental resources and, consequently, promoting the possible co-existence of many species, both native and invasive amphipods (Chesson 2000; Amarasekare 2003). While in the lakes with the lower value of this index, native amphipods may be unable to compete with invasive species and could become extinct. However, our results do not confirm this assumption. We found *G. lacustris* in lakes with high shoreline development (e.g. Lake Ełckie), but no invasive amphipods were found there. Isolation of these lakes and low tourist pressure could result in the lack of conditions for their invasion. Nonetheless, in the event of their invasion, we can suppose that the high shoreline complexity of these lakes would promote the co-existence of native and invasive amphipods.

The declining populations of *G. lacustris* in our studies are similar to the general tendency observed in Europe. This species seems to be one of the weakest competitors amongst European freshwater amphipods giving way to the Ponto-Caspian species of genera: *Chaetogammarus*, *Dikerogammarus* and *Pontogammarus* (Meßner and Zettler 2021). *Gammarus lacustris* occurs in a wide range of habitats; nevertheless, in the last few decades, the species has been pushed to the relict range of occurrence (Hesselschwerdt et al. 2008; Meßner and Zettler 2021). Nowadays, the species is present almost exclusively in isolated waterbodies and continues to decline (Meßner and Zettler 2021). The population decline is also attributed to the hydromorphological and hydrochemical changes that occur in aquatic ecosystems (Okland 1969; Matafonov and Bazova 2014).

Similarly, we did not record another native amphipod, *Pallasiola quadrispinosa*, also recorded as declining in the freshwater ecosystems due to invasive amphipods (Żmudziński 1995; Jażdżewski et al. 2004). According to Jażdżewski and Konopacka (1995), this species was found in several lakes of the Masurian Lakeland, i.e. Dargin, Dobskie, Ełckie, Kisajno, Łaśmiady, Mamry, Mikołajskie, Mokre, Niegocin, Śniardwy and Tałty. In some of these lakes, we collected *G. lacustris* without invasive species which suggests also the possible presence of *P. quadrispinosa* in these lakes. *Pallasiola quadrispinosa* thrives in colder temperatures and typically resides in deeper waters during the summer months, which may explain why the species was not recorded during our summer samplings.

Our findings report the set of lake features promoting the distribution of native amphipod species in the studied lakes. As the study area is highly impacted by tourist activities, our results can be useful for better comprehension of the threats to native amphipods in other regions with similar levels of anthropogenic pressure and biological invasions. Our conclusions may highlight the need to protect isolated lakes from tourism and urban area development.

Distribution of invasive amphipods

Freshwater NIS can easily spread with tourist activities, including yachting and angling in particular. Our results show that the number of boats is one of the factors which best explains the distribution of D. villosus (Fig. 5). The main part of the Masurian Lakeland with a high abundance of this species covers the area of high tourist activities, i.e. lakes from group I in the northern part of the Lakeland (Figs 2, 4). Yachting is a very significant component of tourism in the Masurian Lakeland, reaching 37% of total tourist activities in the region (Kulczyk et al. 2016). Masurian tourist routes run through these lakes, thus, tourist boat activity supplements yachting. In 2016, we recorded D. villosus in all these lakes. A good example is Lake Niegocin, which has a high level of tourist pressure and a rapid invasion of *D. villosus* was observed in 2016. In 2014, the species was absent in this Lake, while in 2016, it constituted 81% of all sampled amphipods. Lake Niegocin is located between the other lakes with high tourist pressure and the Masurian tourist route runs through this lake. The evidence of high tourist activity in this lake can be the high number of car parks per km of shoreline and one of the highest, amongst the Masurian lakes, number of beds in accommodation establishments in 2014 (Kulczyk et al. 2016).

Similar findings were done in other tourist lakes. In Alpine lakes, with higher yachting activity than in the Masurian Lakeland, the expansion of *D. villosus* was caused by yachting and using diving equipment (Bącela-Spychalska et al. 2013; Rewicz et al. 2017). Many species using boat biofouling to invade new waterbodies have broad tolerance to desiccation (Bącela-Spychalska et al. 2013; Glisson et al. 2020). Likewise, *D. villosus* has a high tolerance to air exposure (Rewicz et al. 2014). Moreover, the species is usually associated with another invasive species – zebra mussel (*Dreissena polymorpha*) and can survive up to six days out of the water between mussels fouling the boats (Martens and Grabow 2008). Similarly, the species can be transported with algae and macrophytes (Minchin et al. 2019). It enables them to expand rapidly in new waterbodies, including those isolated from others, by overland transport of boats and yachting equipment.

Overland transport of boats may explain the invasion of *D. villosus* in our study area. Although *D. villosus* was found in most of the recently studied lakes, the species was not found in the River Pisa and the River Węgorapa. These rivers connect the Masurian Lakeland with large rivers, where *D. villosus* is present. It suggests the possible expansion of this species in the Masurian Lakeland by overland transport apart from these rivers. In certain lakes, we did not record *D. villosus*. These lakes have no direct contact with the invaded lakes and low tourist activity almost excludes the possibility of overland boat transport. In contrast to the Alpine lakes, we did not expect diving and angling (using waders) equipment to play a significant role in invading isolated waterbodies by *D. villosus* in the studied area.

Another strong factor explaining the distribution of *D. villosus* is the distance from town. Our results show that this species occurs mainly in the lakes with towns nearby. Proximity to the town and tourist activities are correlated with each other. Most of the marinas are located in towns with well-developed tourist facilities. Indeed, the

proximity to the ports and marinas is an important factor in promoting the expansion of *D. villosus* (Minchin et al. 2019). Higher tourist activities in proximity to urban areas may explain the distribution of *D. villosus* in the study area.

Distribution of other invasive species in the Masurian Lakeland, i.e. P. robustoides, D. haemobaphes and C. ischnus, concerns mainly the lakes with less developed shoreline and rather low water quality (higher class of water status). Predominantly, they are present in lakes with different conditions compared to those where D. villosus was found (Figs 4, 5). Dikerogammarus haemobaphes is the first Ponto-Caspian invasive amphipod recorded in the Masurian Lakeland (Jażdżewski 2003). This species was recorded in most of the studied lakes, as well as in the Rivers Wegorapa and Pisa. The presence of this species in the Rivers Bug and Narew suggests its invasion in the Masurian Lakeland from the east - from the River Dnieper. Despite the broad distribution of D. haemobaphes in the lakeland, this species was quickly over-dominated by P. robustoides. The latter species was first recorded in the study area in 2007 (Jażdżewska and Jażdżewski 2008). Three hypothetical routes of *P. robustoides* invasion to this region were proposed - from Kaliningrad (Russia) via the Pregel and the Wegorapa Rivers; from Lithuania via the Augustów Canal; from the Baltic Sea via the River Vistula and its tributaries (Jażdżewska and Jażdżewski 2008). However, since the first record of D. villosus in the region in 2014, the abundance of both species - D. haemobaphes and P. robustoides drastically decreased until 2016. In 2016, D. villosus became the most abundant species in the lakes studied. These results are not surprising as the latter species is a strong competitor and successfully eliminates other invasive and native amphipods (Dick and Platvoet 2000; Platvoet et al. 2007; Bacela-Spychalska et al. 2012; Rewicz et al. 2014; Mathers et al. 2023). Especially, two of them – D. haemobaphes and P. robustoides – are weaker competitors than D. villosus, occurring in different habitats and occupying different niches (Bacela-Spychalska et al. 2012; Kobak et al. 2016; Poznańska-Kakareko et al. 2021; Copilaș-Ciocianu and Sidorov 2022). PCA and CAPSCALE results did not show a strong pattern in the distribution of these species, contrary to D. villosus, which suggests that D. haemobaphes and P. robustoides avoid niches occupied by D. villosus (Figs 4, 5). Dikerogammarus haemobaphes and Pontogammarus robustoides have high desiccation resistance, enabling their overland transport with vessels (Poznańska et al. 2013). Although sailing and angling activities may play an important role in their spreading (Bacela-Spychalska 2016; Csabai et al. 2020), tourist activities probably play a minor role in their distribution in the Masurian Lakeland (Fig. 5).

Another species rapidly spreading in the Masurian Lakeland is *Chaetogammarus ischnus*. In the study area, this species was recorded for the first time in 2014 (Fig. 2A; Suppl. material 1). In two years, its increasing abundance coincided with the decline of the abundance of *D. haemobaphes* and *P. robustoides* (Fig. 2B). In 2016, *C. ischnus* constituted more than half of the collected individuals in Lake Śniardwy. An especially high abundance of this species was observed in the southern group of lakes (group II), contrary to *D. villosus* occurring mainly in the northern group (group I). Moreover, we recorded *Chaetogammarus ischnus* in the River Pisa and did not record this species in the River Wegorapa. These results may suggest that *C. ischnus* invaded the Masurian

Lakeland from the southern direction, i.e. from the River Narew and then via the River Pisa. However, in the lakes where we recorded D. villosus and C. ischnus co-occurring, the abundance of both species increased. Chaetogammarus ischnus usually occupies similar habitats to D. villosus, i.e. sites with hard substrate, particularly covered by D. polymorpha (Żytkowicz and Kobak 2008; Copilas-Ciocianu and Sidorov 2022). The coexistence of both species may be attributed to the small body size of C. ischnus, which enables this species to occupy microhabitats without interfering with D. villosus (Borza et al. 2018). This microhabitat-scale differentiation allows for both species to exist within the same habitat. Between C. ischnus and P. robustoides, the habitats also overlap, but usually P. robustoides limits the occurrence of C. ischnus because of its larger body size and more predatory diet (Żytkowicz and Kobak 2008). Therefore, we can hypothesise that D. villosus eliminates P. robustoides in the lakes studied and then C. ischnus refills the empty niche. In several lakes, for example, Dargin and Kisajno, we observed that C. ischnus reached a similar abundance in 2016 as P. robustoides had in 2014 (Fig. 2B; Suppl. material 1). Similar rapid invasion of C. ischnus and elimination of native species was observed in the Great Lakes in the USA (Dermott et al. 1998) and River Rhine in Europe (Van der Velde et al. 2000), where rapid range extension of D. villosus was observed as well (Bollache et al. 2004). Chaetogammarus ischnus can disperse over great distances (Witt et al. 1997). This species is capable of utilising natural water connections between different water-bodies, but it can also be transported through shipping (Nalepa et al. 2001). Witt et al. (1997) noted that the euryhaline nature of the species enables it to be transported even via ballast waters. However, the understanding of the invasion process of *C. ischnus* is limited and demands further studies.

Our results constitute an important contribution to the long-term observation of expansion dynamics of Ponto-Caspian amphipods and can be part of global databases monitoring invasive species. Rapid expansions underline the importance of regular, annual samplings in lakes and watersheds connecting them with invasion corridors. We show the very important role of tourist activities in lakes in the expansion of alien amphipods, in particular of D. villosus. These findings underline the important role of permanent monitoring of yachting and shipping vessels. Our predictions can be applicable in other tourist freshwater areas and help designate protection zones limiting boating. Our results can be also valuable to studies on other biofouling taxa. The significance of the town's proximity for the amphipod invasion is due to well-developed tourist facilities in urban areas and possibly water pollution; thus, lakes shorelines and marinas should be controlled as well. Water connections between lakes also should be regularly monitored. Our records of Chelicorophium curvispinum in the River Narew in 2014 and 2016 suggest that this species may be the next recorded invasive amphipod in the Masurian Lakeland (see Suppl. material 1). Some studies show a rapid expansion of C. curvispinum in freshwater ecosystems with the presence of Dreissena polymorpha and shipping (Van den Brink et al. 1993; Jażdżewski and Konopacka 2002). The current distribution of other invasive amphipods, for example, Obesogammarus crassus and Gammarus tigrinus, suggest no direct risk of their expansion in the Masurian Lakeland soon, but permanent monitoring of their expansion is necessary.

Limitations of our study

Although our data come from several years, it is important to indicate that the most recent data come from 2016; thus, the current invasion status in the study area can be worse than what we present here. The lack of lakes where native and invasive amphipods co-occur makes some of our findings difficult to interpret and partially speculative.

One of the crucial findings of our study is the impact of boating on the invasions. However, we need to remember that the methods we used have some limitations. We used the maximum capacity of marinas as the number of boats in use. Although on busy days the percentage of used boats in the total number of moored boats is high, as shown in Ros et al. (2013), these data are not precise and might be an under-representation of reality. We need to remember that, the association between boat density and the propagation of invasive amphipods has not been established through direct observation of vessel biofouling communities.

Lakes, especially those with high shoreline complexity, provide many habitats which various species can occupy. Therefore, analysing the data based on one sampling per lake may not depict the real diversity of the amphipod communities. Especially, the lack of records of amphipods in Lake Nidzkie, which has a connection with other lakes and Masurian tourist routes, suggests not enough efficient sampling. We need to be aware that the absence of a species in one sample does not exclude the possible occurrence of this species in other habitats of the same lake. Thus, our results, showing the replacement of native species by invasive species, present interesting trends, but are insufficient to conclude the extinction of certain species. Therefore, our findings should be perceived as predictions, not postulates.

Future directions

Future studies would benefit from utilising a more thorough and systematic sampling to provide a more accurate and reliable picture of the invasion process.

Our results show the importance of the proximity of sampling points to towns for invasions. Although we assume that this correlation is connected with tourist facilities and pollution, implementation of more data is needed in the future. Particularly, the distance between sampling points or lake centroid and marinas should be implemented in the analyses (Cole et al. 2019; Minchin et al. 2019). A significant effect of distance to marinas on invasions was noted by Minchin et al. (2019). Marinas are critical entry points for many invasive species and may play the role of reservoirs for newly-introduced invaders (Glasby et al. 2007; Ros et al. 2013; Fernández-Rodríguez et al. 2022).

Additionally, using the actual number of boats in use in the area would be advisable as was done in studies by Bacela-Spychalska et al. (2013) and Keramidas et al. (2018). Moreover, our knowledge about what part of the vessels are fouled by amphipods, which particular species can be transported and on what maximum distance is still scarce. Future studies would also benefit from including inspections of boats and ropes to identify potential vectors for amphipods, such as algae and mussels. Dikerogammarus villosus, Chaetogammarus ischnus and Chelicorophium curvispinum can be transported with zebra mussels. Therefore, it is advisable to incorporate data on the occurrence of *D. polymorpha* in lakes and on vessels for future research. The type of vessel can also be an important factor. For instance, motorboats can be vectors of invasions, while canoeing does not play this role (Venohr et al. 2018). In this context, the presence/absence of silent zones, i.e. lakes or their parts where using boats with motors is forbidden, should complete the analyses. Knowledge about the success of the "check, clean and dry" strategy in the study area is missing. We expect that none of these methods is implemented as the local law does not demand their respecting. The method to prevent transporting invasive species on boats, as described in Mohit et al. (2021), should be tested in the Masurian Lakeland. Surveys amongst fishermen and tourists are worth collecting and analysing (Cole et al. 2019). To gain deeper insights into these dynamics, we recommend the establishment of an inter-lakes traffic registry. This registry would provide crucial data regarding boat traffic and potential pathways for the introduction of invasive species. Prevention measures and facilities for anglers should also be studied (Smith et al. 2023). To better understand which species can be transported by vessels, it is important to experimentally test the resistance of different invasive species like C. ischnus and C. curvispinum to desiccation.

As far as the financial and technical situation allows, samples should be collected from a large number of points on each lake. Additionally, studying a greater set-up of lakes would allow better tracking of invasions and more accurate detection of all amphipod species in the lakes. Finally, tracking of the invasion process can be supported by molecular studies (e.g. Mamos et al. (2021)).

Conclusions

The rapid expansion of the invasive Ponto-Caspian amphipods observed in this study aligns with a general trend along European freshwater basins. The contraction of the range and niche of native species when faced with more aggressive (e.g. *D. villosus*) and/ or generalist (e.g. *C. ischnus*) species is something expected and confirmed by our findings. Even though many lakes seem to be still free from amphipod invaders, this may be for a short time considering the abrupt increase we have registered in just two years.

Our study emphasises the need for a comprehensive approach to understanding and addressing the dispersal of alien species through human activity. Our findings highlight the important role of boats in the spread of invasive amphipods within lake systems. The invasion process of *Dikerogammarus villosus* especially suggests the possible impact of overland boat transport in spreading this species in new lakes.

Furthermore, it is essential to raise awareness amongst lake users about the negative consequences of biological invasions and the necessity of implementing a "check, clean and dry" policy. By educating and engaging lake users, we can foster a sense of responsibility and cooperation in preventing the spread of invasive species. Implementing these measures collectively will contribute to better biosecurity practices and safeguard the ecological integrity of lakes against invasive species.

Acknowledgements

We would like to thank Krzysztof Jażdżewski, Ewa Janowska, Mariusz Surlejewski, Olga Antczak-Orlewska, Paula Krzywoźniak, Remi Wattier, Aleksandra Jabłońska and Anna Jażdżewska for help during sampling and their valuable suggestions. We would like to thank Stephen Venn for linguistic corrections and for providing valuable suggestions that improved the overall quality of the manuscript. We are grateful to Denis Copilaș-Ciocianu for the suggestions concerning the taxonomic status of *Spirogammarus major*.

References

- Albano PG, Steger J, Bakker PAJ, Bogi C, Bošnjak M, Guy-Haim T, Huseyinoglu MF, La-Follette PI, Lubinevsky H, Mulas M, Stockinger M, Azzarone M, Sabelli B (2021) Numerous new records of tropical non-indigenous species in the Eastern Mediterranean highlight the challenges of their recognition and identification. ZooKeys 1010: 1–95. https://doi. org/10.3897/zookeys.1010.58759
- Amarasekare P (2003) Competitive coexistence in spatially structured environments: A synthesis. Ecology Letters 6(12): 1109–1122. https://doi.org/10.1046/j.1461-0248.2003.00530.x
- Anderson LG, White PCL, Stebbing PD, Stentiford GD, Dunn AM (2014) Biosecurity and vector behaviour: Evaluating the potential threat posed by anglers and canoeists as pathways for the spread of invasive non-native species and pathogens. PLOS ONE 9(4): 1–10. https://doi.org/10.1371/journal.pone.0092788
- Anderson LG, Rocliffe S, Haddaway NR, Dunn AM (2015) The Role of tourism and recreation in the spread of non-native species: A systematic review and meta-analysis. PLOS ONE 10(10): e0140833. https://doi.org/10.1371/journal.pone.0140833
- Arbačiauskas K, Rakauskas V, Virbickas T (2010) Initial and long-term consequences of attempts to improve fish-food resources in Lithuanian waters by introducing alien peracaridan species: A retrospective overview. Journal of Applied Ichthyology 26: 28–37. https:// doi.org/10.1111/j.1439-0426.2010.01492.x
- Aronow S (1982) Shoreline development ratio. Beaches and Coastal Geology. Encyclopedia of Earth Sciences Series. Springer, New York, 754–755. https://doi.org/10.1007/0-387-30843-1_417
- Bącela-Spychalska K (2016) Attachment ability of two invasive amphipod species may promote their spread by overland transport. Aquatic Conservation 26(1): 196–201. https:// doi.org/10.1002/aqc.2565
- Bącela-Spychalska K, Wattier RA, Genton C, Rigaud T (2012) Microsporidian disease of the invasive amphipod *Dikerogammarus villosus* and the potential for its transfer to local invertebrate fauna. Biological Invasions 14(9): 1831–1842. https://doi.org/10.1007/s10530-012-0193-1
- Bącela-Spychalska K, Grabowski M, Rewicz T, Konopacka A, Wattier R (2013) The 'killer Shrimp' *Dikerogammarus villosus* (Crustacea, Amphipoda) invading Alpine lakes: Overland
transport by recreational boats and scuba-diving gear as potential entry vectors? Aquatic Conservation 23(4): 606–618. https://doi.org/10.1002/aqc.2329

- Bajkiewicz-Grabowska E (2008) Obieg wody w systemie Wielkich Jezior Mazurskich [Water circulation in the Great Masurian Lake System]. In: Jasser I, Robak S, Zdanowski B (Eds) Ochrona i Rekultywacja wód Wielkich Jezior Mazurskich Narzędziem Rozwoju Naukowego, Gospodarczego, Społecznego i Kulturowego Regionu [Protection and Reclamation of Water in the Great Masurian Lakes as a Tool of Scientific, Economic Social and Cultural Development of the Region]. IRS, Olsztyn, 19–29. [In Polish]
- Bellard C, Cassey P, Blackburn M (2016) Alien species as a driver of recent extinctions. Biology Letters 12(2): e20150623. https://doi.org/10.1098/rsbl.2015.0623
- Bij de Vaate A, Jażdżewski K, Ketelaars HAM, Gollasch S, Van der Velde G (2002) Geographical patterns in range extension of Ponto-Caspian macroinvertebrate species in Europe. Canadian Journal of Fisheries and Aquatic Sciences 59(7): 1159–1174. https://doi. org/10.1139/f02-098
- Bollache L, Devin S, Wattier R, Chovet M, Beisel JN, Moreteau JC, Rigaud T (2004) Rapid range extension of the Ponto-Caspian amphipod *Dikerogammarus villosus* in France: Potential consequences. Archiv für Hydrobiologie 160(1): 57–66. https://doi.org/10.1127/0003-9136/2004/0160-0057
- Borza P, Huber T, Leitner P, Remund N, Graf W (2018) How to coexist with the 'killer shrimp' Dikerogammarus villosus? Lessons from other invasive Ponto-Caspian peracarids. Aquatic Conservation 28(6): 1441–1450. https://doi.org/10.1002/aqc.2985
- Bozdogan H (1987) Model selection and Akaik"s Information Criterion (AIC): The general theory and its analytical extensions. Psychometrika 52(3): 345–370. https://doi.org/10.1007/ BF02294361
- Bradshaw C, Leroy B, Bellard C, Roiz D, Albert C, Fournier A, Barbet-Massin M, Salles J-M, Simard F, Courchamp F (2016) Massive yet grossly underestimated global costs of invasive insects. Nature Communications 7(1): e12986. https://doi.org/10.1038/ncomms12986
- Brooks ME, Kristensen K, van Benthem KJ, Magnusson A, Berg CW, Nielsen A, Skaug HJ, Maechler M, Bolker BM (2017) glmmTMB Balances Speed and Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling. The R Journal 9(2): 378–400. https://doi.org/10.32614/RJ-2017-066
- CBD (2000) Global strategy on invasive alien species. Convention on Biological Diversity, UNEP/CBD/SBSTTA/6/INF/9: 1–52.
- Cereghetti E (2023) Stream ecosystems and terrestrial resources: a temporal perspective on detritus cycling and keystone shredders. PHD Thesis. Universität Zürich.
- Chesson P (2000) General theory of competitive coexistence in spatially-varying environments. Theoretical Population Biology 58(3): 211–237. https://doi.org/10.1006/tpbi.2000.1486
- Chróst RJ, Siuda W (2006) Microbial production, utilization, and enzymatic degradation of organic matter in the upper trophogenic layer in the pelagial zone of lakes along a eutrophication gradient. Limnology and Oceanography 51(1part2): 749–762. https://doi. org/10.4319/lo.2006.51.1_part_2.0749
- Cole GA (1975) Textbook of Limnology. C. V. Mosby Co., St. Louis, 283 pp.

- Cole E, Keller RP, Garbach K (2019) Risk of invasive species spread by recreational boaters remains high despite widespread adoption of conservation behaviors. Journal of Environmental Management 229: 112–119. https://doi.org/10.1016/j.jenvman.2018.06.078
- Copilaș-Ciocianu D, Sidorov D (2022) Taxonomic, ecological and morphological diversity of Ponto-Caspian gammaroidean amphipods: A review. Organisms, Diversity & Evolution 22(2): 285–315. https://doi.org/10.1007/s13127-021-00536-6
- Copilaș-Ciocianu D, Rewicz T, Sands AF, Palatov D, Marin I, Arbačiauskas K, Hebert PDN, Grabowski M, Audzijonyte A (2022) A DNA barcode reference library for endemic Ponto-Caspian amphipods. Scientific Reports 12(1): e11332. https://doi.org/10.1038/s41598-022-15442-w
- Copilaș-Ciocianu D, Sidorov D, Šidagytė-Copilas E (2023a) Global distribution and diversity of alien Ponto-Caspian amphipods. Biological Invasions 25(1): 179–195. https://doi. org/10.1007/s10530-022-02908-1
- Copilaş-Ciocianu D, Palatov D, Rewicz T, Sands AF, Arbačiauskas K, van Haaren T, Hebert PN, Grabowski M, Marin I (2023b) A widespread Ponto-Caspian invader with a mistaken identity: integrative taxonomy elucidates the confusing taxonomy of *Trichogammarus trichiatus* (= *Echinogammarus*) (Crustacea: Amphipoda). Zoological Journal of the Linnean Society 198(3): 821–846. https://doi.org/10.1093/zoolinnean/zlad010
- Cristescu MEA, Hebert PDN (2005) The "Crustacean Seas"– An evolutionary perspective on the Ponto-Caspian peracarids. Canadian Journal of Fisheries and Aquatic Sciences 62(3): 505–517. https://doi.org/10.1139/f04-210
- Csabai Z, Borza P, Rewicz T, Pernecker B, Berta BJ, Móra A (2020) Mass appearance of the Ponto-Caspian invader *Pontogammarus robustoides* in the River Tisza catchment: Bypass in the southern invasion corridor? Knowledge and Management of Aquatic Ecosystems 421(421): 1–9. https://doi.org/10.1051/kmae/2020003
- Cuthbert RN, Diagne C, Hudgins EJ, Turbelin A, Ahmed DA, Albert C, Bodey TW, Briski E, Essl F, Haubrock PJ, Gozlan RE, Kirichenko N, Kourantidou M, Kramer AM, Courchamp F (2021a) Biological invasion costs reveal insufficient proactive management worldwide. The Science of the Total Environment 819: e153404. https://doi.org/10.1016/j.scitotenv.2022.153404
- Cuthbert RN, Pattison Z, Taylor NG, Verbrugge I, Diagne C, Ahmed DA, Leroy B, Angulo E, Briski E, Capinha C, Catford JA, Dalu T, Ells F, Gozlan RE, Haubrock PJ, Kourantidou M, Kramer AM, Renault D, Wasserman RJ, Courchamp F (2021b) Global economic costs of aquatic invasive species. The Science of the Total Environment 775: e145238. https:// doi.org/10.1016/j.scitotenv.2021.145238
- De Ventura L, Wissert N, Tobias R, Kopp K, Jokela J (2016) Overland transport of recreational boats as a spreading vector of zebra mussel *Dreissena polymorpha*. Biological Invasions 18(5): 1451–1466. https://doi.org/10.1007/s10530-016-1094-5
- Dermott R, Witt J, Um YM, González M (1998) Distribution of the Ponto-Caspian Amphipod *Echinogammarus ischnus* in the Great Lakes and Replacement of Native *Gammarus fasciatus*. Journal of Great Lakes Research 24(2): 442–452. https://doi.org/10.1016/S0380-1330(98)70834-2
- Dick JTA, Platvoet D (1996) Intraguild predation and species exclusions in amphipods: The interaction of behaviour, physiology and environment. Freshwater Biology 36(2): 375–383. https://doi.org/10.1046/j.1365-2427.1996.00106.x

- Dick JTA, Platvoet D (2000) Invading predatory crustacean *Dikerogammarus villosus* eliminates both native and exotic species. Proceedings of the Royal Society B, Biological Sciences 267(1447): 977–983. https://doi.org/10.1098/rspb.2000.1099
- Dudgeon D, Arthington AH, Gessner MO, Kawabata ZI, Knowler DJ, Lévêque C, Naiman RJ, Prieur-Richard AH, Soto D, Stiassny MLJ, Sullivan CA (2006) Freshwater biodiversity: Importance, threats, status and conservation challenges. Biological Reviews of the Cambridge Philosophical Society 81(2): 163–182. https://doi.org/10.1017/S1464793105006950
- Eggers TO, Martens A (2001) Bestimmungsschlüssel der Süßwasser-Amphipoda (Crustacea) Deutschlands. Lauterbornia 42: 1–68. [In German]
- Ejsmont-Karabin J, Kalinowska K, Karpowicz M (2020) Structure of Ciliate, Rotifer, and Crustacean Communities in Lake Systems of Northeastern Poland. In: Korzeniewska E, Harnisz M (Eds) Polish River Basins and Lakes – Part II. The Handbook of Environmental Chemistry 87. Springer, Cham, 77–101. https://doi.org/10.1007/978-3-030-12139-6_4
- European Environment Agency (2000) Water Framework Directive (WFD) 2000. https:// www.eea.europa.eu/policy-documents/water-framework-directive-wfd-2000
- European Environment Agency (2018) State of Water report 7/2018. https://www.eea.europa.eu/publications/state-of-water
- Fee EJ (1979) A relation between lake morphometry and primary productivity and its use in interpreting whole-lake eutrophication experiments. Limnology and Oceanography 24(3): 401–416. https://doi.org/10.4319/lo.1979.24.3.0401
- Feld CK, Birk S, Eme D, Gerisch M, Hering D, Kernan M, Maileht K, Mischke U, Ott I, Pletterbauer F, Poikane S, Salgado J, Sayer CD, Van Wichelen J, Malard F (2016) Disentangling the effects of land use and geo-climatic factors on diversity in European freshwater ecosystems. Ecological Indicators 60: 71–83. https://doi.org/10.1016/j.ecolind.2015.06.024
- Fernández-Rodríguez I, López-Alonso R, Sánchez O, Suárez-Turienzo I, Gutiérrez-Martínez R, Arias A (2022) Detection and prevention of biological invasions in marinas and ports: Epibionts and associated fauna of *Mytilus galloprovincialis* revisited. Estuarine, Coastal and Shelf Science 274: e107943. https://doi.org/10.1016/j.ecss.2022.107943
- Foley JA, DeFries R, Asner GP, Barford C, Bonan G, Carpenter SR, Chapin FS, Coe MT, Daily GC, Gibbs HK, Helkowski JH, Holloway T, Howard EA, Kucharik CJ, Monfreda C, Patz JA, Prentice IC, Ramankutty N, Snyder PK (2005) Global consequences of land use. Science 309(5734): 570–574. https://doi.org/10.1126/science.1111772
- Galil BS, Nehring S, Panov V (2008) Waterways as Invasion Highways Impact of Climate Change and Globalization. In: Nentwig W (Ed.) Biological Invasions. Ecological Studies (Vol. 193). Springer, Berlin. https://doi.org/10.1007/978-3-540-36920-2_5
- Gerhardt A, Bloor M, Lloyd Mills C (2011) Gammarus: Important Taxon in Freshwater and Marine Changing Environments. International Journal of Zoology 2011: 524276. https:// doi.org/10.1155/2011/524276
- Glasby TM, Connell SD, Holloway MG, Hewitt CL (2007) Nonindigenous biota on artificial structures: Could habitat creation facilitate biological invasions? Marine Biology 151(3): 887–895. https://doi.org/10.1007/s00227-006-0552-5
- Glisson WJ, Wagner CK, Verhoeven R, Muthukrishnan R, Contreras-Rangel R, Larkin DJ (2020) Desiccation tolerance of the invasive alga starry stonewort (*Nitellopsis obtusa*) as an indicator of overland spread risk. Journal of Aquatic Plant Management 58: 7–18.

- Grabowski M, Konopacka A, Jażdżewski K, Janowska E (2006) Invasions of alien gammarid species and retreat of natives in the Vistula Lagoon (Baltic Sea, Poland). Helgoland Marine Research 60(2): 90–97. https://doi.org/10.1007/s10152-006-0025-8
- Grabowski M, Jażdzewski K, Konopacka A (2007) Alien crustacea in Polish waters–Amphipoda. Aquatic Invasions 2(1): 25–38. https://doi.org/10.3391/ai.2007.2.1.3
- Grabowski M, Bącela K, Konopacka A, Jażdżewski K (2009) Salinity-related distribution of alien amphipods in rivers provides refugia for native species. Biological Invasions 11(9): 2107–2117. https://doi.org/10.1007/s10530-009-9502-8
- Grigorovich IA, MacIsaac HJ, Shadrin NV, Mills EL (2002) Patterns and mechanisms of aquatic invertebrate introductions in the Ponto-Caspian region. Canadian Journal of Fisheries and Aquatic Sciences 59(7): 1189–1208. https://doi.org/10.1139/f02-088
- Grigorovich IA, Kang M, Ciborowski JJH (2005) Colonization of the Laurentian Great Lakes by the amphipod *Gammarus tigrinus*, a native of the North American Atlantic Coast. Journal of Great Lakes Research 31(3): 333–342. https://doi.org/10.1016/S0380-1330(05)70264-1
- Gruszka P (1999) The river Odra Estuary as a gateway for alien species immigration to the Baltic Sea basin. Acta Hydrochimica et Hydrobiologica 27(5): 374–382. https://doi.org/10.1002/(SICI)1521-401X(199911)27:5<374::AID-AHEH374>3.0.CO;2-V
- Hall MC, Härkönen T (2006) Lake Tourism: An Integrated Approach to Lacustrine Tourism Systems. Channel View Publications, Bristol, 256 pp. https://doi. org/10.21832/9781845410421
- Hartig F (2022) DHARMa: Residual Diagnostics for Hierarchical (Multi-Level / Mixed) Regression Models. R package version 0.4.6.
- Haubrock PJ, Pilotto F, Innocenti G, Cianfanelli S, Haase P (2021) Two centuries for an almost complete community turnover from native to non-native species in a riverine ecosystem. Global Change Biology 27(3): 606–623. https://doi.org/10.1111/gcb.15442
- Hesselschwerdt J, Meeker J, Wantzen KM (2008) Gammarids in Lake Constance: Habitat segregation between the invasive *Dikerogammarus villosus* and the indigenous *Gammarus roeselii*. Fundamental and Applied Limnology 173(3): 177–186. https://doi. org/10.1127/1863-9135/2008/0173-0177
- Hewitt CL, Gollasch S, Minchin D (2009) The vessel as a vector Biofouling, ballast water and sediments. In: Rilov G, Crooks JA (Eds) Biological Invasions in Marine Ecosystems. Ecological Studies (Vol. 204). Springer, Berlin, 117–131. https://doi.org/10.1007/978-3-540-79236-9_6
- Hillbricht-Ilkowska A, Rybak J, Rzepecki M (2000) Ecohydrological research of lake watershed relations in diversified landscape (Masurian Lakeland, Poland). Ecological Engineering 16(1): 91–98. https://doi.org/10.1016/S0925-8574(00)00093-8
- Iqbal MF, Feng Y-L, Feng W-W, Liu M-C, Lu X-R (2021) Ecological impacts of the invasive plant *Xanthium strumarium* and the impacts of three aboveground herbivores on the invader. Ecological Indicators 131: e108140. https://doi.org/10.1016/j.ecolind.2021.108140
- Jażdżewska A, Jażdżewski K (2008) *Pontogammarus robustoides* (G.O. Sars, 1894) (Crustacea, Amphipoda), a new Ponto-Caspian invader in Great Masurian Lakes (NE Poland). Fragmenta Faunistica 51(1): 1–7. https://doi.org/10.3161/00159301FF2008.51.1.001

- Jażdżewska AM, Rewicz T, Mamos T, Wattier R, Bącela-Spychalska K, Grabowski M (2020) Cryptic diversity and mtDNA phylogeography of the invasive demon shrimp, *Dikero-gammarus haemobaphes* (Eichwald, 1841), in Europe. NeoBiota 57: 53–86. https://doi. org/10.3897/neobiota.57.46699
- Jażdżewski K (1975) Morfologia, Taksonomia i Występowanie w Polsce Kiełży z Rodzajów Gammarus Fabr. i Chaetogammarus MART. (Crustacea, Amphipoda). Acta Universitatis Lodziensis, Łódź, 185 pp. [In Polish]
- Jażdżewski K (1980) Range Extensions of Some Gammaridean Species in European Inland Waters Caused by Human Activity. Crustaceana 6: 84–107.
- Jażdżewski K (2003) An invasive Ponto-Caspian amphipod *Dikerogammarus haemobaphes* (Eichwald, 1841)–conquers Great Masurian Lakes, north-eastern Poland. Fragmenta Faunistica 46(1): 19–25. https://doi.org/10.3161/00159301FF2003.46.1.019
- Jażdżewski K, Konopacka A (1995) Malacostraca prócz Oniscoidea. In: Mroczkowski M (Ed.) Katalog Fauny Polskiej 13(1): 1–165. [In Polish]
- Jażdżewski K, Konopacka A (2000) Immigration history and present distribution of alien crustaceans in Polish waters. In: von Vaupel Klein JC, Schram FR (Eds) The Biodiversity Crisis and Crustacea, Proceedings of 4th International Crustacean Congress, Brill, Leiden. Crustacean Issues 12: 55–64.
- Jażdżewski K, Konopacka A (2002) Invasive ponto-caspian species in waters of the vistula and oder basins and the Southern Baltic Sea. In: Leppäkoski E, Gollasch S, Olenin S (Eds) Invasive Aquatic Species of Europe. Distribution, Impacts and Management. Springer, Dordrecht, 384–398. https://doi.org/10.1007/978-94-015-9956-6_39
- Jażdżewski K, Konopacka A, Grabowski M (2002) Four Ponto-Caspian and one American gammarid species (Crustacea, Amphipoda) recently invading Polish waters. Contributions to Zoology 71(4): 115–122. https://doi.org/10.1163/18759866-07104001
- Jażdżewski K, Konopacka A, Grabowski M (2004) Recent drastic changes in the gammarid fauna (Crustacea, Amphipoda) of the Vistula River deltaic system in Poland caused by alien invaders. Diversity & Distributions 10(2): 81–87. https://doi.org/10.1111/j.1366-9516.2004.00062.x
- Jażdżewski K, Konopacka A, Grabowski M (2005) Native and alien Malacostracan Crustacea along the Polish Baltic Sea coast in the twentieth century Oceanological and Hydrobiological Studies (Vol. XXXIV, Supplement 1). Oceanological and Hydrobiological Studies 34: 175–193.
- Johnson LE, Padilla DK (1996) Geographic spread of exotic species: Ecological lessons and opportunities from the invasion of the zebra mussel *Dreissena polymorpha*. Biological Conservation 78(1–2): 23–33. https://doi.org/10.1016/0006-3207(96)00015-8
- Jones CG, Lawton JH, Shachak M (1994) Organisms as ecosystem engineers. Oikos 69(3): 373–386. https://doi.org/10.2307/3545850
- Jones CG, Lawton JH, Shachak M (1997) Positive and negative effects of organisms as physical ecosystem engineers. Ecology 78(7): 1946–1957. https://doi.org/10.1890/0012-9658(1997)078[1946:PANEOO]2.0.CO;2
- Keramidas I, Dimarchopoulou D, Pardalou A, Tsikliras AC (2018) Estimating recreational fishing fleet using satellite data in the Aegean and Ionian Seas (Mediterranean Sea). Fisheries Research 208: 1–6. https://doi.org/10.1016/j.fishres.2018.07.001

- Kettunen M, Genovesi P, Gollasch S, Pagad S, Starfinger U, Ten Brink P, Shine C (2009) Technical support to EU strategy on invasive species (IAS)– Assessment of the impacts of IAS in Europe and the EU (final module report for the European Commission). Institute for European Environmental Policy (IEEP), Brussels, Belgium. 44 pp. [+ Annexes]
- Kistowski M, Śleszyński P (2010) Presja turystyczna na tle walorów turystycznych Polski. Prace Komisji Krajobrazu Kulturowego Nr 14. [In Polish]
- Kobak J, Rachalewski M, Bącela-Spychalska K (2016) Conquerors or exiles? Impact of interference competition among invasive Ponto-Caspian gammarideans on their dispersal rates. Biological Invasions 18(7): 1953–1965. https://doi.org/10.1007/s10530-016-1140-3
- Kondracki J (2002) Geografia regionalna Polski, Warszawa, PWN, [ISBN 83-01-13897-1]. [In Polish]
- Konopacka A (1998) Nowy dla Polski gatunek kiełża, Dikerogammarus haemobaphes (EICH-WALD, 1841) (Crustacea, Amphipoda) oraz dwa inne rzadkie gatunki skorupiaków obunogich w Wiśle. Przegląd Zoologiczny 3–4: 211–218. [in Polish]
- Konopacka A, Jażdżewski K (2002) Obesogammarus crassus (G.O. Sars, 1894) one more Ponto-Caspian gammarid species in Polish waters. Fragmenta Faunistica 45(1): 19–26. https:// doi.org/10.3161/00159301FF2002.45.1.019
- Kouba A, Oficialdegui FJ, Cuthbert RN, Kourantidou M, South J, Tricarico E, Gozlan RE, Courchamp F, Haubrock PJ (2022) Identifying economic costs and knowledge gaps of invasive aquatic crustaceans. The Science of the Total Environment 813: e152325. https:// doi.org/10.1016/j.scitotenv.2021.152325
- Kulczyk S, Derek M, Woźniak E (2016) Zagospodarowanie turystyczne strefy brzegowej jezior na potrzeby żeglarstwa–przykład wielkich jezior mazurskich. Prace i Studia Geograficzne 61(3): 27–49. [In Polish]
- Kurashov EA, Barbashova MA, Barkov DV, Rusanov AG, Lavrova MS (2012) Invasive amphipods as a factor of transformation of Lake Ladoga ecosystems. Russian Journal of Biological Invasions 3(3): 202–212. https://doi.org/10.1134/S2075111712030058
- Lambertini M, Leape J, Marton-Lefevre J, Mittermeier R, Rose M, Robinson J, Stuart S, Waldman B, Genovesi P (2011) Invasives: A major conservation threat. Science 333(6041): 404–405. https://doi.org/10.1126/science.333.6041.404-b
- Legendre P, Anderson MJ (1999) Distance-based redundancy analysis: Testing multispecies responses in multifactorial ecological experiments. Ecological Monographs 69(1): 1–24. https://doi.org/10.1890/0012-9615(1999)069[0001:DBRATM]2.0.CO;2
- Lipták B, Veselý L, Ercoli F, Bláha M, Buřič M, Ruokonen T, Kouba A (2019) Trophic role of marbled crayfish in a lentic freshwater ecosystem. Aquatic Invasions 14(2): 299–309. https://doi.org/10.3391/ai.2019.14.2.09
- Lobato TC, Hauser-Davis RA, Oliveira TF, Silveira AM, Silva HAN, Tavares MRM, Saraiva ACF (2015) Construction of a novel water quality index and quality indicator for reservoir water quality evaluation: A case study in the Amazon region. Journal of Hydrology 522: 674–683. https://doi.org/10.1016/j.jhydrol.2015.01.021
- Mačić V, Albano PG, Almpanidou V, Claudet J, Corrales X, Essl F, Evagelopoulos A, Giovos I, Jimenez C, Kark S, Marković O, Mazaris AD, Ólafsdóttir G, Panayotova M, Petović S, Rabitsch W, Ramdani M, Rilov G, Tricarico E, Fernández TV, Sini M,

Trygonis V, Katsanevakis S (2018) Biological invasions in conservation planning: A global systematic review. Frontiers in Marine Science 5: e178. https://doi.org/10.3389/fmars.2018.00178

- Mamos T, Grabowski M, Rewicz T, Bojko J, Strapagiel D, Burzyński A (2021) Mitochondrial genomes, phylogenetic associations, and SNP recovery for the key invasive Ponto-Caspian amphipods in Europe. International Journal of Molecular Sciences 22(19): e10300. https:// doi.org/10.3390/ijms221910300
- Martens A, Grabow K (2008) Das risiko der verschleppung neozoischer Amphipoda beim überlandtransport von yachten. Lauterbornia 62: 41–44. https://doi.org/10.3391/ai.2007.2.1.4 [in German]
- Martinez Arbizu P (2020) pairwiseAdonis: Pairwise multilevel comparison using adonis. R package version 0.4 mazury24.eu. https://mazury24.eu [in Polish]
- Matafonov D, Bazova N (2014) Decline of *Gammarus lacustris* Sars (Crustacea: Amphipoda) population in the delta of the Selenga River. Biology Bulletin of the Russian Academy of Sciences 41(2): 168–175. https://doi.org/10.1134/S1062359014020071
- Mathers KL, Clinton K, Constable D, Gerrard C, Patel C, Wood PJ (2023) Invasion dynamics of Ponto-Caspian amphipods leads to changes in invertebrate community structure and function. Freshwater ecology 14: 1–14. https://doi.org/10.1002/ecs2.4593
- Meßner U, Zettler MB (2021) Drastic changes of the amphipod fauna in northern Germany and the displacement of *Gammarus lacustris* G.O. Sars, 1864 to relict habitats/ status. Knowledge and Management of Aquatic Ecosystems 422(17): 1–8. https://doi. org/10.1051/kmae/2021016
- Minchin D, Arbačiauskas K, Daunys D, Ezhova E, Grudule N, Kotta J, Molchanova N, Olenin S, Višinskienė G, Strake S (2019) Rapid expansion and facilitating factors of the Ponto-Caspian invader *Dikerogammarus villosus* within the Eastern Baltic sea. Aquatic Invasions 14(2): 165–181. https://doi.org/10.3391/ai.2019.14.2.02
- Mishra M, Singhal A, Srinivas R (2023) Effect of urbanization on the urban lake water quality by using water quality index (WQI). Materials Today: Proceedings: 1–9. https://doi. org/10.1016/j.matpr.2023.06.357
- Mohit S, Johnson TB, Arnott SE (2021) Recreational watercraft decontamination: Can current recommendations reduce aquatic invasive species spread? Management of Biological Invasions : International Journal of Applied Research on Biological Invasions 12(1): 148–164. https://doi.org/10.3391/mbi.2021.12.1.10
- Mordukhai-Boltovskoi FD (1964) Caspian fauna beyond the Caspian Sea. Internationale Revue der Gesamten Hydrobiologie 49(1): 139–176. https://doi.org/10.1002/iroh.19640490105
- Nalepa TF, Schloesser DW, Pothoven SA, Hondorp DW, Fanslow DL, Tuchman ML, Fleisher GW (2001) First finding of the amphipod *Echinogammarus ischnus* and the mussel *Dreissena bugensis* in Lake Michigan. Journal of Great Lakes Research 27(3): 384–391. https:// doi.org/10.1016/S0380-1330(01)70653-3
- Nalepa TF, Fanslow DL, Lang GA (2009) Transformation of the offshore benthic community in Lake Michigan: Recent shift from the native amphipod *Diporeia* spp. to the invasive mussel *Dreissena rostriformis bugensis*. Freshwater Biology 54(3): 466–479. https://doi. org/10.1111/j.1365-2427.2008.02123.x

- Nehring S (2005) International shipping–a risk for aquatic biodiversity in Germany. Biological invasions–from ecology to control. NeoBiota 6: 125–143.
- Okland KA (1969) On the distribution and ecology of *Gammarus lacustris* G.O. Sars in Norway, with notes on its morphology and biology. Norvegian journal of zoology 17: 111–152.
- Oksanen J, Simpson G, Blanchet F, Kindt R, Legendre P, Minchin P, Hara R, Solymos P, Stevens M, Szoecs E, Wagner H, Barbour M, Bedward M, Bolker B, Borcard D, Carvalho G, Chirico M, De Caceres M, Durand S, Evangelista H, FitzJohn R, Friendly M, Furneaux B, Hannigan G, Hill M, Lahti L, McGlinn D, Ouellette M, Ribeiro Cunha E, Smith T, Stier A, Ter Braak C, Weedon J (2022) vegan: Community Ecology Package. R package version 2.6-4. https://CRAN.R-project.org/package=vegan
- Panlasigui S, Davis AJS, Mangiante MJ, Darling JA (2018) Assessing threats of non-native species to native freshwater biodiversity: Conservation priorities for the United States. Biological Conservation 224: 199–208. https://doi.org/10.1016/j.biocon.2018.05.019
- Panov VE, Alexandrov B, Arbačiauskas K, Binimelis R, Copp GH, Grabowski M, Lucy F, Leuven RS, Nehring S, Paunović M, Semenchenko V, Son MO (2009) Assessing the risks of aquatic species invasions via European inland waterways: From concepts to environmental indicators. Integrated Environmental Assessment and Management 5(1): 110–126. https://doi.org/10.1897/IEAM_2008-034.1
- Pinkster S, Scheepmaker M, Platvoet D, Broodbakker N (1992) Drastic changes in the amphipod fauna (Crustacea) of Dutch inland waters during the last 25 years. Bijdragen tot de Dierkunde 61(4): 193–204. https://doi.org/10.1163/26660644-06104001
- Platvoet D, Song Y, Shuqiang L, Van der Velde G (2007) Description of the lateral line organ of *Dikerogammarus villosus* (Sowinsky, 1894), with discussion on its function (Peracarida, Amphipoda). Crustaceana 80(11): 1373–1392. https://doi. org/10.1163/156854007782605619
- Pöckl M, Grabowski M, Grabowska J, Bącela-Spychalska K, Wittmann KJ (2011) Large European rivers as biological invasion highways. In: Habersack H, Schober B, Walling D (Eds) International Conference on the Status and Future of the World's Large Rivers, 11–14 April 2011. Conference Abstract Book, Vienna, 215 pp.
- Poznańska M, Kakareko T, Krzyżyński M, Kobak J (2013) Effect of substratum drying on the survival and migrations of Ponto-Caspian and native gammarids (Crustacea: Amphipoda). Hydrobiologia 700(1): 47–59. https://doi.org/10.1007/s10750-012-1218-6
- Poznańska-Kakareko M, Lis M, Kakareko T, Augustyniak M, Kłosiński P, Kobak J (2021) Nearshore distribution of alien Ponto-Caspian amphipods in a European dam reservoir in relations to substratum type and occurrence of macroinvertebrate taxa. Knowledge and Management of Aquatic Ecosystems 422(5): 1–12. https://doi.org/10.1051/kmae/2021005
- Pyšek P, Richardson DM (2010) Invasive species, environmental change and management, and health. Annual Review of Environment and Resources 35(1): 25–55. https://doi. org/10.1146/annurev-environ-033009-095548
- QGIS Development Team (2020) QGIS Geographic Information System. Open Source Geospatial Foundation Project. https://qgis.org/en/site/about/index.html
- R Core Team (2023) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna. https://www.R-project.org/

- Rachalewski M, Grabowski M, Konopacka A, Bacela-Spychalska K (2013) *Echinogammarus trichiatus* (Martynov, 1932)' A new Ponto-Caspian amphipod invader in Poland with remarks on other alien amphipods from the Oder river. Crustaceana 86(10): 1224–1233. https://doi.org/10.1163/15685403-00003228
- Rewicz T, Grabowski M, Macneil C, Bącela-Spychalska K (2014) The profile of a 'perfect' invader – the case of killer shrimp, *Dikerogammarus villosus*. Aquatic Invasions 9(3): 267– 288. https://doi.org/10.3391/ai.2014.9.3.04
- Rewicz T, Wattier R, Rigaud T, Grabowski M, Mamos T, Bącela-Spychalska K (2017) The killer shrimp, *Dikerogammarus villosus*, invading European Alpine Lakes: A single main source but independent founder events with an overall loss of genetic diversity. Freshwater Biology 62(6): 1036–1051. https://doi.org/10.1111/fwb.12923
- Ricciardi A (2006) Patterns of Invasion in the Laurentian Great Lakes in Relation to Changes in Vector Activity. Diversity & Distributions 12(4): 425–433. https://doi.org/10.1111/ j.1366-9516.2006.00262.x
- Ricciardi A, Kipp R (2008) Predicting the number of ecologically harmful exotic species in an aquatic system. Diversity & Distributions 14(2): 374–380. https://doi.org/10.1111/j.1472-4642.2007.00451.x
- Ricciardi A, MacIsaac HJ (2000) Recent mass invasion of the North American Great lakes by Ponto-Caspian species. Trends in Ecology & Evolution 15(2): 62–65. https://doi. org/10.1016/S0169-5347(99)01745-0
- Rodríguez-Rey M, Consuegra S, Börger L, Garcia de Leaniz C (2021) Boat ramps facilitate the dispersal of the highly invasive zebra mussel (*Dreissena polymorpha*). Biological Invasions 23(5): 1487–1496. https://doi.org/10.1007/s10530-020-02453-9
- Ros M, Vázquez-Luis M, Guerra-García JM (2013) The role of marinas and recreational boating in the occurrence and distribution of exotic caprellids (Crustacea: Amphipoda) in the Western Mediterranean: Mallorca Island as a case study. Journal of Sea Research 83: 94– 103. https://doi.org/10.1016/j.seares.2013.04.004
- Sala OE, Chapin FS, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, Huber-Sanwald E, Huenneke LF, Jackson RB, Kinzig A, Leemans R, Lodge DM, Mooney HA, Oesterheld M, Poff NL, Sykes MT, Walker BH, Walker M, Wall DH (2000) Global biodiversity scenarios for the year 2100. Science 287(5459): 1770–1774. https://doi.org/10.1126/science.287.5459.1770
- Sánchez E, Colmenarejo MF, Vicente J, Rubio A, García MG, Travieso L, Borja R (2007) Use of the water quality index and dissolved oxygen deficit as simple indicators of watersheds pollution. Ecological Indicators 7(2): 315–328. https://doi.org/10.1016/j.ecolind.2006.02.005
- Sardain A, Sardain E, Leung B (2019) Global forecasts of shipping traffic and biological invasions to 2050. Nature Sustainability 2(4): 274–282. https://doi.org/10.1038/s41893-019-0245-y
- Smith ERC, Bennion H, Sayer CD, Aldridge DC, Owen M (2020) Recreational angling as a pathway for invasive non-native species spread: Awareness of biosecurity and the risk of long distance movement into Great Britain. Biological Invasions 22(3): 1135–1159. https://doi.org/10.1007/s10530-019-02169-5

- Smith ERC, Heal R, Wood LE (2023) Understanding and improving biosecurity among recreational anglers in Great Britain. Journal of Fish Biology 102(5): 1177–1190. https://doi. org/10.1111/jfb.15345
- Soszka H, Kolada A, Pasztaleniec A, Ochocka A, Kutyła S, Bielczyńska A (2016) Opracowanie zbiorczej oceny stanu jezior. Institute of Environmental Protection – National Research Institute. [In Polish]
- Strayer DL, Dudgeon D (2010) Freshwater biodiversity conservation: Recent progress and future challenges. Journal of the North American Benthological Society 29(1): 344–358. https://doi.org/10.1899/08-171.1
- UIA World Countries Boundaries (2023) UIA World Countries Boundaries. https://hub.arcgis.com/datasets/UIA::uia-world-countries-boundaries
- Ulikowski D, Traczuk P, Kapusta A, Kalinowska K (2021) New records of alien monkey goby, *Neogobius fluviatilis* (Pallas, 1814), in the waters of the Great Masurian Lakes system (northeastern Poland). BioInvasions Records 10(4): 924–933. https://doi.org/10.3391/ bir.2021.10.4.17
- Väinölä R, Witt JDS, Grabowski M, Bradbury JH, Jażdżewski K, Sket B (2008) Freshwater Animal Diversity Assessment. Hydrobiologia 595: 241–255.
- Van den Brink FWB, Van der Velde G, Bij de Vaate A (1993) Ecological aspects, explosive range extension and impact of a mass invader, *Corophium curvispinum* Sars, 1895 (Crustacea: Amphipoda), in the Lower Rhine (The Netherlands). Oecologia 93(2): 224–232. https://doi.org/10.1007/BF00317675
- Van der Velde G, Rajagopal S, Musko I, Bij de Vaate A (2000) Ecological impact of crustacean invaders. General considerations and examples from the Rhine River. In: von Vaupel Klein JC, Schram FR (Eds) The biodiversity crisis and Crustacea, Proceedings of 4th International Crustacean Congress, Brill, Leiden, Crustacean Issues 12: 3–33.
- Van der Velde G, Leuven RSEW, Platvoet D, Bącela K, Huijbregts MAJ, Hendriks HWM, Kruijt D (2009) Environmental and morphological factors influencing predatory behaviour by invasive non-indigenous gammaridean species. Biological Invasions 11(9): 2043– 2054. https://doi.org/10.1007/s10530-009-9500-x
- Vander Zanden MJ, Olden JD (2008) A management framework for preventing the secondary spread of aquatic invasive species. Canadian Journal of Fisheries and Aquatic Sciences 65(7): 1512–1522. https://doi.org/10.1139/F08-099
- Vantarová KH, Eliáš Jr P, Jiménez-Ruiz J, Tokarska-Guzik B, Cires E (2023) Biological invasions in the twenty-first century: A global risk. Biologia 78(5): 1211–1218. https://doi. org/10.1007/s11756-023-01394-7
- Venohr M, Langhans SD, Peters O, Hölker F, Arlinghaus R, Mitchell L, Wolter C (2018) The underestimated dynamics and impacts of water-based recreational activities on freshwater ecosystems. Environmental Reviews 26(2): 199–213. https://doi.org/10.1139/er-2017-0024
- Witt JDS, Hebert PDN, Morton WB (1997) Echinogammarus ischnus: Another crustacean invader in the Laurentian Great Lakes basin. Canadian Journal of Fisheries and Aquatic Sciences 54(2): 264–268. https://doi.org/10.1139/f96-292
- Yang Q, Weigelt P, Fristoe TS, Zhang Z, Kreft H, Stein A, Seebens H, Dawson W, Essl F, König C, Lenzner B, Pergl J, Pouteau R, Pyšek P, Winter M, Ebel AL, Fuentes N,

Giehl ELH, Kartesz J, Krestov P, Kukk T, Nishino M, Kupriyanov A, Villaseñor JL, Wieringa JJ, Zeddam A, Zykova E, van Kleunen M (2021) The global loss of floristic uniqueness. Nature Communications 12(1): e7290. https://doi.org/10.1038/s41467-021-27603-y

- Zhulidov AV, Kozhara AV, van der Velde G, Leuven RSEW, Son MO, Gurtovaya TY, Zhulidov DA, Nalepa TF, Santiago-Fandino VJR, Chuikov YS (2018) Status of the invasive brackish water bivalve *Mytilopsis leucophaeata* (Conrad, 1831) (Dreissenidae) in the Ponto-Caspian region. BioInvasions Records 7(2): 111–120. https://doi.org/10.3391/ bir.2018.7.2.02
- Żmudziński L (1995) Retreat of *Pallasiola quadrispinosa* [G.O.Sars] and *Monoporeia affinis* [Lindstrom] from the Polish lakes. Polskie Archiwum Hydrobiologii 42(4): 401–407.
- Żytkowicz J, Kobak J (2008) Relationships between the occurrence of *Chaetogammarus ischnus* (Stebbing, 1898) and *Pontogammarus robustoides* (G. O. Sars, 1894) (Crustacea, Amphipoda) in a lowland Polish dam reservoir. Oceanological and Hydrobiological Studies 37(2): 39–50. https://doi.org/10.2478/v10009-007-0050-1

Supplementary material I

Metadata for each sampling site and date

Authors: Krzysztof Podwysocki, Andrea Desiderato, Tomasz Mamos, Tomasz Rewicz, Michał Grabowski, Alicja Konopacka, Karolina Bącela-Spychalska

Data type: xlsx

- Explanation note: Sampling sites between the years 2001–2016 with a number of individuals (or +/-) for the presence/absence) of each recorded Amphipoda species (names of invasive species have been underlined). Symbols for lakes provide twoletter acronyms used on the figures. Symbols for rivers and canals provide threeletter acronyms used on the figures. Water QS: Water quality status (Soszka et al. 2016); A/V ratio: Surface area to volume ratio (Soszka et al. 2016); Shoreline development: Shoreline length to surface area ratio (https://mojemazury.pl); Density of boats: number of boats per ha of lake surface (https://mazury24.eu; https:// skorupki.mazury.info.pl).
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.109221.suppl1

Supplementary material 2

Summary of the best-fitting Bernoulli GLMM for the presence of native gammarid – *Gammarus lacustris*

Authors: Krzysztof Podwysocki, Andrea Desiderato, Tomasz Mamos, Tomasz Rewicz, Michał Grabowski, Alicja Konopacka, Karolina Bącela-Spychalska

Data type: docx

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.109221.suppl2

Supplementary material 3

Results of PERMANCOVA test using 9999 permutations

Authors: Krzysztof Podwysocki, Andrea Desiderato, Tomasz Mamos, Tomasz Rewicz, Michał Grabowski, Alicja Konopacka, Karolina Bącela-Spychalska

Data type: docx

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.109221.suppl3

Supplementary material 4

Supplementary image

Authors: Krzysztof Podwysocki, Andrea Desiderato, Tomasz Mamos, Tomasz Rewicz, Michał Grabowski, Alicja Konopacka, Karolina Bącela-Spychalska

Data type: jpeg

Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.109221.suppl4

RESEARCH ARTICLE



Competitive interaction in headwaters: slow upstream migration leads to trophic competition between native and non-native amphipods

Lars Pelikan^{1,2,3}, Eglė Šidagytė-Copilas², Andrius Garbaras⁴, Jonas Jourdan¹, Denis Copilaș-Ciocianu²

I Goethe University Frankfurt, Department Aquatic Ecotoxicology, Max-von-Laue-Straße 13, D-60438 Frankfurt am Main, Germany 2 Nature Research Centre, Laboratory of Evolutionary Ecology of Hydrobionts, Akademijos St. 2, LT-08412 Vilnius, Lithuania 3 University of Turku, Department of Biology, Vesilinnantie 5, FI-20014 Turku, Finland 4 Centre for Physical Sciences and Technology, Saulètekio Ave. 3, LT-10257 Vilnius, Lithuania

Corresponding author: Lars Pelikan (lars.pelikan@utu.fi)

Academic editor: Jaimie T. A. Dick | Received 6 October 2023 | Accepted 2 January 2024 | Published 1 February 2024

Citation: Pelikan L, Šidagytė-Copilas E, Garbaras A, Jourdan J, Copilaș-Ciocianu D (2024) Competitive interaction in headwaters: slow upstream migration leads to trophic competition between native and non-native amphipods. NeoBiota 90: 193–216. https://doi.org/10.3897/neobiota.90.112383

Abstract

The spread of non-native species is one of the outcomes of global change, threatening many native communities through predation and competition. Freshwater ecosystems are particularly affected by species turnover with non-native species. One species that has been established in Central Europe for many decades - or even a few centuries - is the amphipod crustacean Gammarus roeselii. Although G. roeselii is nowadays widespread in major river systems, there have been recent reports of its spread into smaller streams that are typically inhabited by the native species Gammarus fossarum. Due to their leaf shredding ability, G. fossarum takes up a key position in headwater streams. This raises the important question, to what extent G. roeselii can equivalently take over this function. To answer this question, we collected both species from nine different sites in a mid-mountain river system (Kinzig catchment, Hesse, Germany) and investigated their functional similarity using a combination of stable isotope analysis, gut content and functional morphology. The species hardly differed in morphological characteristics, only females showed differences in some traits. Gut content analysis indicated a broad dietary overlap, while stable isotopes showed a higher trophic position of G. roeselii. The observed functional overlap could intensify interspecific competition and allow the larger and more predaceous G. roeselii to replace G. fossarum in the future as a headwater keystone species. However, the differentiation in the stable isotopes also shows that co-existence can occur by occupying different trophic niches. Moreover, the wide range of inhabited sites and exploited resources demonstrate the omnivorous lifestyle of G. roeselii, which is likely to help the species succeed in rapidly changing environments.

Copyright Lars Pelikan et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Keywords

Freshwater ecosystems, functional morphology, *Gammarus*, gut content analysis, stable isotope analysis, trophic niche

Introduction

The introduction and spread of non-native species pose a threat to native communities globally (Gallardo et al. 2016). Due to their interconnection, streams are particularly affected by invasive species (Bij de Vaate et al. 2002; Leuven et al. 2009), which can cause a decrease of native species through predation and competition (Van der Velde et al. 2000). This change is not only expressed on a taxonomic level (Van der Velde et al. 2009), but probably also leads to changes in functional characteristics of ecosystems (Rosenfeld 2002). Non-native species might possess traits and behaviour that enable them to fulfil similar ecological functions as the native species they displace. However, the non-native species did not evolve within the recipient ecosystem and might lack natural predators, competitors or parasites that would otherwise regulate their populations in their native range (Torchin et al. 2003; Sih et al. 2010). Consequently, this unregulated population growth can lead to changes in resource availability and disrupting ecological interactions. However, post-invasion transformations of community structure and ecosystem function can differ immensely, are context-dependent and, often, poorly understood (Parker et al. 1999; Strayer et al. 2006; Kenis et al. 2009; Jeschke et al. 2014; Kumschick et al. 2015; Bellard et al. 2016).

A key group that is currently enormously affected by species-turnover is the taxonomic group of amphipods (Jażdżewski 1980; Leuven et al. 2009). Native Gammarus spp. are key species in freshwater ecosystems, due to their role in the decomposition of organic matter - an essential process in the headwaters of stream ecosystems (Cummins and Klug 1979; Vannote et al. 1980; Gessner et al. 1999; Graça 2001). However, the species turnover that is currently attracting attention is taking place in larger rivers (Leuven et al. 2009; Jourdan et al. 2016), while small headwaters – which are often not monitored within the Water Framework Directive - receive far less attention. While Gammarus fossarum Koch, 1836, is a typical headwater species, Gammarus roeselii Gervais, 1835, mainly inhabits the downstream parts of the rivers (Pöckl and Humpesch 1990; Pöckl et al. 2003). The exact origin and classification (native or non-native) of G. roeselii is still under debate, but the reduced genetic diversity (Csapó et al. 2020) and the main distribution on the Balkan Peninsula (Grabowski et al. 2017; Kabus et al. 2023) suggest that G. roeselii is probably an older invader in northern and western river systems outside the Danube system (Jażdżewski and Roux 1988). The species has been described near Paris in 1835 (Jażdżewski and Roux 1988), probably from a non-native population and has been spreading in Western Europe in the past centuries (Jażdżewski 1980; Jażdżewski and Roux 1988; Csapó et al. 2020). Nowadays, we observe an increasing spread of G. roeselii into smaller tributaries and upstream regions (Jourdan et al. 2019). In the Kinzig catchment (Hesse, Germany), some first order streams are

already colonised by *G. roeselii*, while others are still inhabited by *G. fossarum*. Interestingly, there are also streams where both occur syntopically. The similar phenotypic adaptation along environmental gradients in both species (Jourdan et al. 2019; Grethlein et al. 2022) now raises the question to which extent they share a similar trophic niche.

Investigating the trophic ecology of invasive species is necessary to gain a better understanding of the community-wide effects of invasions (Tillberg et al. 2007). Studies have shown that invaders across the animal kingdom often have a flexible and generalist diet (Garton et al. 2005; Tillberg et al. 2007; Caut et al. 2008; Zhang et al. 2010; Grey and Jackson 2012; Jackson et al. 2017; Mothapo and Wossler 2017). Especially in changing environments, omnivorous species or species with a broad trophic niche are more successful than those with a more specialised diet, so that omnivorous invasive species may prey on or compete with native species (Barbosa and Castellanos 2005; Simberloff 2010; Jackson et al. 2017).

The ecological niche of an organism is connected with its functional morphology (Bock and von Wahlert 1965). Therefore, morphological traits can provide additional important ecological insights (Premate et al. 2021). Characterising such traits – like body size and mouthparts – is suitable to evaluate the relationship between morphology and trophic ecology, which, in turn, can be tested against further results of stable isotope and gut content analyses (Premate et al. 2021). Morphological analysis combined with stable isotope analysis can be used to elucidate potential relationships between morphology and function (Hutchins et al. 2014), since trophic levels characterise the functional role of organisms (Hairston and Hairston 1993). So far, the relationship between morphology and ecology is not yet fully understood in many invertebrate groups and only now starting to become clearer in amphipods (Copilaş-Ciocianu et al. 2021).

Stable isotope analyses are useful for answering general questions about trophic structure (Vander Zanden et al. 1999; Post et al. 2000). However, this analysis is not able to distinguish very well between closely-related food sources with a similar isotopic value (McCutchan et al. 2003; Hood-Nowotny and Knols 2007; Bowes and Thorp 2015). Therefore, to gain qualitative information necessary to interpret the isotopic results, stable isotopes should be used together with other information, such as the analysis of the gut content (Inger and Bearhop 2008). This analysis has, in previous studies, been demonstrated to have a strong correlation with stable isotope data and, thus, can be considered a reliable reflection of diet in amphipods (Bacela-Spychalska and Van Der Velde 2013; Aumack et al. 2017). Copilaş-Ciocianu et al. (2021) identified a strong relationship between morphological traits and the proportion of food items found in the gut. These correlations can provide an important understanding of the functional morphology of freshwater amphipods (Copilaş-Ciocianu et al. 2021).

Stable isotope analysis is a common method used to reveal the trophic position of organisms in the field (Peterson et al. 1986; Cabana and Rasmussen 1996; Vander Zanden and Rasmussen 1999; Vander Zanden et al. 1999; Post et al. 2000; Post 2002; Layman et al. 2007). This analysis is an important tool to study food webs and has already been successfully used to detect the impact of biological invasions on trophic

structures (Vander Zanden et al. 1999; Van Riel et al. 2006; Mancinelli and Vizzini 2015; McCue et al. 2020). For nitrogen, isotope ratios ($^{15}N/^{14}N$; $\delta^{15}N$) of a consumer are on average 3 to 5‰ higher than of dietary items and increase with successive trophic levels (DeNiro and Epstein 1981; Minagawa and Wada 1984; Peterson and Fry 1987; Vander Zanden and Rasmussen 1999; Layman et al. 2007). On the contrary, the ratios of carbon isotopes ($^{13}C/^{12}C$; $\delta^{13}C$) show only minor changes since carbon moves through the food web with little alteration (DeNiro and Epstein 1978; Rounick and Winterbourn 1986; Peterson and Fry 1987; Layman et al. 2007).

We hypothesise that, in the focal study area, *G. fossarum* and *G. roeselii* share a similar trophic niche. This equivalence is suggested by a laboratory experiment where *G. roeselii* showed the same leaf consumption rate as *G. fossarum* (Jourdan et al. 2016). Moreover, we expect a high level of competition amongst ecologically similar species, which should lead to a niche shift in co-occurrence scenarios. Finally, *G. roeselii* is found both in large lowland rivers, but also occasionally in upper stretches, while *G. fossarum* is restricted to upper stretches (Pöckl and Humpesch 1990; Pöckl et al. 2003). Given that headwaters mainly receive energy input through terrestrial organic material, resulting in simpler food webs, while lower stretches exhibit more complexity due to increased nutrient availability and primary production (Vannote et al. 1980), we hypothesise that *G. roeselii* has a broader trophic niche compared to *G. fossarum*.

Materials and methods

Fieldwork

Gammarus fossarum and *G. roeselii* were collected with the kick-sampling method on 10 and 11 August 2021 at nine sampling sites in the Kinzig catchment in Hesse, Germany (Table 1; Fig. 1). The sampling sites and their site IDs correspond with the sites from Weigand et al. (2020). Adult and juvenile individuals, as well as submerged leaves of *Alnus* sp. or *Corylus* sp. (depending on availability), were collected at each sampling site. Gammarids displaying visible parasitism, such as acanthocephalans (Médoc et al. 2011;

Site ID	Stream	Species	GPS
1	Gründau	G. fossarum + G. roeselii	50°14.93'N, 9°9.33'E
5	Bracht	G. fossarum + G. roeselii	50°22.62'N, 9°16.22'E
6	Bracht	G. fossarum + G. roeselii	50°26.22'N, 9°16.43'E
7	Salz	G. roeselii	50°25.00'N, 9°21.80'E
10	Ulmbach	G. fossarum + G. roeselii	50°20.18'N, 9°25.70'E
20	Schwarzbach	G. fossarum + G. roeselii	50°21.53'N, 9°33.11'E
33	Kinzig headwater	G. fossarum + G. roeselii	50°18.87'N, 9°36.98'E
37	Haselsbach	G. fossarum	50°13.87'N, 9°22.21'E
105	Riedbach, Kinzig	G. fossarum	50°22.62'N, 9°31.58'E

Table 1. The nine sampling sites within the Kinzig catchment. The site ID (according to Weigand et al. (2020), name of the stream, the collected species and the GPS coordinates are given for each sampling site.



Figure 1. The nine sampling sites within the Kinzig catchment.

Kochmann et al. 2023), were omitted from the analysis due to their potential to impact the feeding behaviour and metabolic processes of the organisms.

Morphological and gut content analysis

The methodology largely followed Copilas-Ciocianu et al. (2021). The animals were first soaked overnight in a 2% lactic acid solution and transferred to a 1:1 solution of 70% ethanol and glycerine. Dissections we performed in glycerine with the help of fine needles and microsurgical scissors. Appendages were glycerine-mounted on microscope slides. Photographs were taken using a Nikon DS-Fi2 camera attached to a Nikon Eclipse Ci-L microscope or a Nikon SMZ1000 stereomicroscope. Measurements were taken with Digimizer software (https//www.digimizer.com/), based on the photographs. A total of 15 female G. fossarum, 12 male G. fossarum, 14 female G. roeselii and 12 male G. roeselii were used for this analysis. We measured 35 functional morphological traits that mainly reflect the diet (body length, mouthparts, stomach and shape of gnathopods). Additionally, traits, indirectly related to diet that reflect sensory function (antennae) and locomotion (pereiopods), were measured as well. For an overview of landmarks and function of these traits, see Copilas-Ciocianu et al. (2021). The same individuals used for the morphological analysis were used for the gut content analysis. The gut content analysis followed the protocol of Copilas-Ciocianu et al. (2021), using a Nikon Eclipse Ci-L microscope. In short, the gut was emptied out and evenly spread on a glass slide with a square grid (24×24 mm) containing 10×10

smaller squares. The gut content was classified into six categories (alga, arthropod, detritus, fungus, plant, sand) and the number of squares on which a particular food item occurred were counted to calculate proportions.

Stable isotope analysis

Two gammarids within each 2 mm size class were used for the analysis. Amphipods with a body size lower than 7 mm were considered juvenile. In total, 26 juveniles, 18 females and 24 males of *G. fossarum* and 19 juveniles, 36 females and 28 males of *G. roeselii* were used. Three leaf replicates per site were used as baseline for trophic position estimates. All samples were sorted, washed with distilled water and dried for 48 h at 60 °C. Afterwards, they were ground to a fine powder with a pestle and mortar. The powder for each amphipod sample was aimed to be 1 mg. In case juveniles did not have a high enough body mass, a composite sample of more individuals of the same species, sex and size class was used. The aimed weight for the leaves was between 3 mg and 4 mg powder for each sample. Carbon and nitrogen stable isotope ratios were measured at the Isotopic Research Laboratory of the Centre for Physical Sciences and Technology in Vilnius, Lithuania. Here, an elemental analyser coupled to the isotope ratio mass spectrometer (EA-IRMS, Flash EA1112–Thermo V Advantage) via the ConFlo III interface was used for the measurement.

In our amphipod samples, the C:N mass ratio varied over 3.5 and in our leaf samples the C exceeded 40%; thus, we corrected the δ^{13} C values for lipid content using the relevant formulae for aquatic animals and plants from Post et al. (2007). We referenced the trophic position of amphipods to Δ C and Δ N in each site by subtracting the corresponding mean values of the leaves. We further divided the Δ N by the conventional trophic fractionation factor value of 3.4‰ (Post 2002) and added one (baseline trophic level) to obtain the amphipod trophic level (TL) estimates.

Statistical analysis

To correct for body size, the morphological measurements were first regressed against the body length and the residuals across all sampling sites were pooled into four species by sex groups (female *G. roeselii*, male *G. roeselii*, female *G. fossarum*, and male *G. fossarum*). The mean value of residuals was used in case of missing values. The gut content data of all sampling sites were grouped in the same manner. Subsequently, principal component analyses (PCA) were conducted in PAST 4 (version 4.08; Hammer et al. 2001). They were computed with a correlation matrix. To test for dietary or morphological differences amongst groups, a one-way permutational multivariate analysis of variance (PERMANOVA) with 9999 permutations and Euclidean similarity index was conducted in PAST. The PERMANOVA was performed either on the gut content data or all morphological traits combined or separately on four trait-complexes reflecting sensorial function (antennae, six measurements), food grasping and manipulation (gnathopods, eight measurements), food processing and digestion (mouthparts and stomach, four measurements) and locomotion (pereiopods along with their accompanying bases and coxae, 15 measurements). To test for differences between gammarids occurring alone compared to co-occurrence, PERMANOVA was performed on gut content and all morphological traits between those sites. Bonferroni correction was applied for further multiple comparisons between group pairs.

We analysed the difference in trophic position between species by building linear mixed-effects models (LMEMs) for each of the referenced metrics (ΔC and TL). In these models, we considered the interacting fixed effects of size, species and the syntopy (or co-occurrence) factor, while site was included as a random factor. The continuous size variable was centred around the global mean of 8.25 mm for more proper effect testing (but back-transformations were applied for the provided visuals). The effects were tested using type III analysis of variance with Satterthwaite's approximation for denominator degrees of freedom. These analyses were conducted by employing the R packages *lme4* v. 1.1-32 and *lmerTest* v. 3.1-3 (Kuznetsova et al. 2017) and visualised using the package visreg v. 2.7.0 (Breheny and Burchett 2017). We further conducted the *post hoc* group comparisons at minimum and maximum amphipod sizes with the Sidak p-value adjustment for eight tests using the package emmeans v. 1.8.8 (Lenth 2023). To test for the differences in ΔC and TL between sexes, we extracted the residuals from the LMEMs for the adult observations only and applied simple linear models with backward stepwise removal of terms (based on Akaike Information Criterion), starting with fully interacting effects of species, sex and syntopy.

In the bivariate stable isotope space, we approximated the population isotopic niches as ellipses containing 95% of the data with their area estimates (BEA_{95%}). We also estimated the overlaps between the species in the six syntopic sites and standardised them as proportions of the sum of the non-overlapping ellipse areas (0 – no overlap, 1 – complete overlap). For this, we used the Bayesian estimation available in R package *SIBER* v. 1.2.7 (Jackson et al. 2011) and provided the estimates as modes with 95% credible intervals of the posterior distributions (400 draws). Using these draws, within each of the six syntopic sites, we tested if: (1) the BEA_{95%} of *G. roeselii* is wider than the ellipse of *G. fossarum*, (2) if the overlap is larger than 0 and (3) if the overlap is larger than 60% which is the commonly considered threshold for an ecologically significant overlap (Jackson et al. 2011). Provided probabilities were translated to be treated conventionally – significant effects when p < 0.05.

Results

Morphological analysis

Results from omnibus PERMANOVA testing for all traits (F = 12.7, p = 0.001) revealed a significant morphological differentiation between sexes within *G. roeselii* and within *G. fossarum* and between females of both species, but not males (see Suppl. material 1: table S1 for pairwise comparisons). This could be confirmed by the PCA, since the convex hulls of the males of both species are mainly overlapping, whereas the females are more separated from each other (Fig. 2a). Pairwise comparisons (Suppl. material 1: table S1, Fig. 2b–e) further showed that females differed significantly between species only with regard to gnathopods and pereiopod+coxae trait complex (Fig. 2c, d). The pairwise comparisons also indicated that the sexual dimorphism within both species was reflected by significant differences in all tested trait complexes. Regarding the comparison between gammarids occurring alone compared to co-occurring, the omnibus PERMANOVA testing for all traits (*G. fossarum* alone/co-occurring: F = 0.54, p = 0.64; *G. roeselii* alone/co-occurring: F = 0.96, p = 0.38) revealed no significant morphological differentiation.



Figure 2. Morphological differentiation between studied male and female gammarid species according to principal component analyses (PCAs) of all traits (**a**), antennae (**b**), gnathopods (**c**), pereiopods+coxae (**d**) and mouthparts (**e**).

Gut content analysis

The PCA of the gut content indicated that the most important differentiation between specimens was amongst the detritus, sand vs. plant axis, explaining 29.9% of variation (Fig. 3). The second axis of differentiation was between detritus and sand and it explained 21.4% of the variance (Fig. 3). The PERMANOVA revealed no significant differentiation between species or sexes when only considering co-occurrences (F = 1.3, p = 0.24). All groups overlapped to a great extent (Fig. 3). Results from PERMANO-VA testing for gut content between syntopic and non-syntopic populations did reveal a significant differentiation between *G. fossarum* alone/co-occurring with *G. roeselii* (F = 3.6, p = 0.032), but not between *G. roeselii* alone/co-occurring with *G. fossarum* (F = 2.6, p = 0.065, Fig. 3b).



Figure 3. A principal component analysis (PCA) scatterplot depicting dietary differentiation with respect to the proportion of various food items of species by **a** sex and **b** by non-syntopic and syntopic occurrence (sexes combined).

Stable isotope analysis

Isotopic position

The results of stable isotope analysis revealed pronounced niche differentiation between *G. fossarum* and *G. roeselii*, with *G. roeselii* generally occupying a higher trophic level (Fig. 4). At most sites, ΔC values varied indicating ¹³C-enrichment of amphipods relative to the tree-leaf detritus. However, we observed a notable separation of site 7, where *G. roeselii* was exceptionally ¹³C-depleted relative to the leaves (coincidentally, *Alnus* sp. leaves were missing at the site, thus *Corylus* sp. were used). Trophic level of amphipods mostly fell within the normal range between herbivory (TL = 2) and complete carnivory (TL = 3), with the extremes of 1.8 in *G. fossarum* juveniles from site 37 and slightly above 3.0 in the largest adults of *G. roeselii* from site 6 and site 10. The trend of increasing TL with size was also reflected in the data. Estimates of trophic



Figure 4. Referenced stable isotope biplot of studied amphipods showing trophic niches by site (means \pm SD). Point size reflects animal body size. Green labels at the means correspond to site IDs from Table 1.

position for different amphipod size groups derived from stable isotope analysis are provided in Suppl. material 1: table S2.

Both LMEMs of ΔC and TL (Table 2) returned a significant second order interaction of amphipod size, species and the syntopy factor, indicating variable steepness of the ontogenetic slopes of gammarids. Judging by the various modelled cases (Fig. 5), there was always an increasing ΔC and TL trend with amphipod size and *G. roeselii* generally tended to exhibit lower ΔC and higher TL values than *G. fossarum*. Regarding both metrics, the interspecific differences in the overall position, as well as the slopes, were more evident across the non-syntopic sites, while, when co-occurring, the two species tended to overlap more and to exhibit very similar ontogenetic slopes. Across the syntopic sites, both amphipods appeared to have high ΔC values, similar as in *G. fossarum*-only sites and their juveniles converged more to intermediate TL values. The ΔC slope of *G. roeselii* and TL slope of *G. fossarum* were visibly steeper across the non-syntopic sites in comparison to the syntopic ones, suggesting wider ontogenetic niches in respective metrics when a single species is found. (It is important to note that, to some extent, the observed patterns may have been biased by the single *G. roeselii*only site coinciding with a more degraded habitat.)



Figure 5. Effects of amphipod size, species and their syntopic occurrence on isotopic metrics of trophic position within the linear mixed-effects models of **a**, **b** referenced δ^{13} C (Δ C) and **c**, **d** of trophic level (TL) by **a**, **c** non-syntopic vs. **b**, **d** syntopic sites. See Table 2 for effect tests. Grey asterisks between species' lines indicate significant (p < 0.05) interspecific differences at corresponding size extremes according to *post hoc* analysis. Not shown here, but this analysis also indicated significant Δ C differences within the smallest and largest specimens of *G. roeselii* occurring in non-syntopic vs. syntopic sites.

Table 2. Results of analysis of variance (type III decomposition) from the linear mixed-effects models of isotopic metrics of trophic position – referenced $\delta^{13}C$ (ΔC) and trophic level (TL) – testing for the interacting effects of amphipod size, species (*Gammarus fossarum* vs. *G. roeselii*) and their syntopic occurrence. See Fig. 5 for effect plots.

Tested term	df	ΔC model		TL model			
		$df_{\rm denominator}$	F	p	$df_{ m denominator}$	F	p
Size	1	142.6	35.2	< 0.001	142.1	43.4	< 0.001
Species	1	10.6	125.1	< 0.001	9.4	7.0	0.026
Syntopy	1	9.7	114.8	< 0.001	9.1	0.6	0.459
Size : Species	1	142.7	6.3	0.013	142.1	5.0	0.027
Size : Syntopy	1	142.6	14.7	< 0.001	142.1	0.6	0.444
Species : Syntopy	1	10.6	87.8	< 0.001	9.4	1.9	0.202
Size : Species : Syntopy	1	142.7	7.1	0.008	142.1	4.3	0.039

Regarding the effect of sex across the adult dataset, the stepwise procedure removed all the effects apart from sex from the linear model of ΔC and indicated a model without predictors for TL (although sex was removed last). Thus, we ended up applying simple *t*-tests using only the sex factor. These indicated a marginally higher female ΔC ($t_{104} = 1.9$, p = 0.061), but no effect of sex on TL ($t_{104} = 0.4$, p = 0.69).

Isotopic niche width and overlaps

Isotopic niche widths of the populations and the overlaps between species are provided in Table 3. Although the mode estimates of $\text{BEA}_{95\%}$ were often larger in *G. roeselii* than in *G. fossarum*, the Bayesian testing indicated such a pattern significantly only in site 10 (p = 0.030 as opposed to $p \ge 0.84$ in other cases). All the six overlaps were larger than 0 ($p \le 0.037$), but none of them exceeded 60% ($p \ge 0.96$), indicating a relatively low trophic niche overlap.

Table 3. Populat	ion isotopic niche wid	ths as ellipse areas	(BEA _{95%}) and	d their absolute and	d relative over-
laps by study site.	The Bayesian estimate	es are provided as n	nodes and 95	% credible interval	S.

Site ID	G. fossarum BEA _{95%}	G. roeselii BEA _{95%}	Overlap	%Overlap
37	3.47 (1.56-6.59)	-	-	-
105	5.20 (3.01-9.21)	-	-	-
6	3.51 (2.10-6.79)	3.05 (1.96-5.37)	0.02 (0.00-1.44)	0.00 (0.00-0.20)
10	2.10 (0.99-4.07)	4.76 (3.09-9.63)	1.61 (0.46-2.88)	0.26 (0.08-0.49)
20	1.31 (0.66–2.94)	1.93 (1.34–3.67)	0.70 (0.00-1.47)	0.27 (0.00-0.55)
33	3.53 (2.09-8.38)	4.85 (3.10-8.40)	1.73 (0.15-3.17)	0.24 (0.05-0.39)
1	3.63 (1.56-9.25)	4.78 (1.79–11.19)	0.04 (0.00-3.30)	0.00 (0.00-0.32)
5	3.84 (1.94–10.98)	2.39 (1.42-4.85)	1.19 (0.00-2.50)	0.02 (0.00-0.40)
7	-	6.75 (3.66–12.41)	-	-

Discussion

Our study revealed a strong overlap in morphology and gut content between the native *G. fossarum* and non-native *G. roeselii* in headwater streams. However, stable isotopes indicated a stronger dietary differentiation between the two species when occurring alone and a more similar trophic niche when occurring together, with *G. roeselii*, however, generally occupying a higher trophic level. In addition, the gut content analysis confirmed that *G. fossarum* appears to have a different diet when occurring alone. This indicates that, despite their apparent functional morphological equivalence, the two species exploit different food resources. Below, we expand on the significance of these findings.

Overall, morphological differentiation occurs between the sexes rather than between species when looking at the combined data, pooled over all sampling sites. Only in some traits of the females (gnathopods, pereiopods and coxae) could we see differences between the species. This differentiation between females of both species could be explained by different reproduction characteristics of the species, such as different thermal optima for maximum fecundity (Pöckl 1993). As reproduction involves a metabolic cost (Sutcliffe 1992), a different reproduction effort could mean that different energy levels are available for growth and, thus, translating into different sizes of morphological features. Besides, the size of males is significantly affected by sexual selection, while females are significantly affected by natural selection (Ward 1988). Sexual selection of a trait is induced by competition over mates (Andersson 1994), while natural selection is induced by trait variation amongst individuals (Endler 1986). The competition between males of both species might be similar throughout the sampling sites and, therefore, the males show similar morphological traits. On the other hand, females of both species experiencing natural selection might lead to morphological trait differentiation. Similar morphological features of males of both species point to an absence of morpho-functional differentiation, the occupation of a similar trophic niche and ecological similarity (Cothran et al. 2013; Fišer et al. 2015; Copilaș-Ciocianu et al. 2021). However, morphological similarity does not always mean ecological equivalence. Fišer et al. (2015) found ecological differentiation between four morphologically similar Niphargus species, which could potentially lead to different ecological roles in the ecosystem. Moreover, Premate et al. (2023) discovered that morphological traits are influenced not only by trophic position, but also by the specific habitat type. This observation could also explain the similar morphology of our study species occurring in the same headwater habitat.

The gut content analysis showed a strong dietary overlap between the species when pooled over all sampling sites. Our results thus indicate that the foraging on the same food sources might lead to competition between the two species. In principle, a strategy adopted by different amphipods to reduce competition for limited resources could be to utilise resources in different ways, in different microhabitats or at different times (Piscart et al. 2010, 2011; Mauchart et al. 2017; Premate et al. 2021). Apart from that, our species differed significantly regarding the δ^{13} C level, which has also been observed in other co-occurring freshwater amphipods (Premate et al. 2021). Such partitioning of food resources is expected amongst co-occurring species (Schoener 1974; Chesson 2000), facilitating a stable co-existence (Chesson 2000). Our analysis showed a significant differentiation in gut content for G. fossarum when occurring alone compared to co-occurring with G. roeselii and close to significance for G. roeselii when occurring alone compared to co-occurring with G. fossarum. This could be explained by their, in general, separated occurrence in different river sections (Pöckl and Humpesch 1990; Pöckl et al. 2003) and the different availability of resources in these sections (Vannote et al. 1980). On the other hand, the absence of dietary separation for co-occurring populations might be a consequence of high abundance of food during the study period and could indicate potential for interspecific competition (Piscart et al. 2011; Rothhaupt et al. 2014). Interspecific competition, in turn, can reduce intraspecific specialisation, therefore, reducing the niche breadth of individuals (Araújo et al. 2011), leading to a dietary overlap. In addition, dietary separations or overlaps can change frequently in populations depending on spatio-temporal availability of food items and on the abundance of competing species (Rothhaupt et al. 2014). Thus, stable isotope analysis is useful for studying the diet on a longer time-scale.

The stable isotope analysis revealed that both species had more similar trophic niches when they co-occurred. Specifically, *G. fossarum* underwent a trophic level increase while *G. roeselii* a decrease which was also accompanied by a shift in the carbon source. Although this result was unexpected and non-intuitive at a first glance, it could be explained by reciprocal predation on juveniles or recently moulted individuals of the other species, as observed in another native and non-native amphipod species pair (Dick et al. 1993; Dick 1996; Dick and Platvoet 1996). Moreover, other researchers have discovered niche segregation in co-occurring amphipods in order to facilitate co-existence and avoid competition (Piscart et al. 2011; Premate et al. 2021).

Nevertheless, our stable isotope analysis revealed that, even though both species experience a niche shift, G. roeselii still has generally a higher trophic position compared to G. fossarum when both species occur together, contradicting our first hypothesis of a shared trophic niche. One reason why we have not found this more carnivorous lifestyle in the gut content could be that animal material can be digested more quickly (Guerra-García et al. 2014), but the stable isotopes of animal prey are incorporated in the tissues of the amphipod and, thus, influence the stable isotope analysis. The higher trophic position of the non-native G. roeselii observed in our stable isotope analysis could be an advantage compared to the native G. fossarum. Animal material, a high quality and easily-assimilated food item, can improve the growth of Gammarus sp. (Anderson and Cummins 1979; Crenier et al. 2017). The acceleration of growth, maturation and reproduction progress when being fed animal matter has been observed for several Gammarus species (Vassallo and Steele 1980; Smith 1988; Delong et al. 1993). In addition, it has been observed for G. fasciatus that, with increasing size, the amount of animal matter found in the gut increased proportionally (Summers et al. 1997). The present study showed that larger individuals occupy a higher trophic niche, confirming an ontogenetic niche shift. Thus, since G. roeselii usually has a larger body length, it will presumably consume more animal material than the smaller G. fossarum (Delong et al. 1993). In return, individuals of G. fossarum have a lower trophic position than G. roeselii. Since G. fossarum also has a smaller body size and smaller gnathopods than G. roeselii, it may be mainly limited to detritus because their smaller gnathopods may not be large enough to handle larger food items, such as animal matter (Summers et al. 1997). This is supported by a laboratory experiment of Delong et al. (1993), which showed that small individuals of G. fasciatus showed a delayed growth response when fed animal matter, but a normal growth when fed only leaf litter. However, G. fossarum is generally also known to be predatory and able to exploit animal food resources (Stoffels et al. 2011; Georgievová et al. 2020; Syrovátka et al. 2020) resulting in gammarids generally being classified as rather omnivorous (MacNeil et al. 1997).

Overall, our results indicate that *G. roeselii* exhibits a broader trophic niche than *G. fossarum* confirming our second hypothesis. It has been shown that, when *G. roeselii* co-occurs with *G. fossarum*, it significantly affects their micro-distribution (Mauchart et al. 2017). *G. roeselii* chooses habitats with macrophytes, (dead) wood or deposits of CPOM (coarse particulate organic matter), whereas *G. fossarum* prefers coarse gravel and cobbles (Mauchart et al. 2017). This suggest different sources of algal or terrestrial

 δ^{13} C (Finlay 2001, 2004) for the diet of *G. roeselii* and could be an explanation for its broader trophic niche. Moreover, in our stable isotope analysis, population from site 7 appears to be an outlier with lower δ^{13} C values. Site 7, unlike the other sites, was highly morphologically degraded, canalised and without significant riparian vegetation. Unlike G. fossarum, G. roeselii is able to occur in many anthropogenically shaped habitats (Mauchart et al. 2014; Enns et al. 2023; Kochmann et al. 2023). Apparently, in such habitats, G. roeselii is able to exploit resources from different food chains, such as algae. On the other hand, the larger size of *G. roeselii* could be an explanation for its broader trophic niche, because the larger size can offer competitive advantage (Young 2004). This advantage arises from the potential for a wider dietary range as size increases, potentially resulting in accelerated growth rates and shorter reproduction times (Summers et al. 1997). Moreover, it enables the exploitation of seasonal changes of food items (Delong et al. 1993). High trophic diversity is one of the most important factors responsible for the dispersal success of amphipods (Legeżyńska et al. 2012), because invasive species must be able to gain food material in heterogeneous habitats (Rothhaupt et al. 2014). Nevertheless, in a future study, it needs to be elucidated whether G. roeselii has a similar or different niche in their native range on the Balkan Peninsula. Presumably, G. roeselii could become even more competitive through depending on leaf litter as constant adaptation, effectively exploiting leaf litter, and also higher quality food sources and continuing to colonise tributaries and headwaters (Jourdan et al. 2016).

The omnivorous diet of both amphipods, the higher trophic niche of G. roeselii seen in our stable isotope analysis and the observed microhabitat partitioning in the field (Mauchart et al. 2017) can facilitate and explain the co-existence of both species. However, unlike G. fossarum, G. roeselii possesses dorsal pleosomal spines. An experimental study identified these as an effective defence mechanism against predatory fish (Bollache et al. 2006). The protection through spines can probably not only facilitate its spatial distribution, but may also favour the colonisation of new niches, hence increasing its invasion potential (Copilas-Ciocianu et al. 2020). The low predation of G. roeselii may promote its establishment in communities with native species, such as G. fossarum (Bollache et al. 2006). Moreover, headwater sections of streams are particularly impacted by anthropogenic pollution and a subsequent species turnover (Betz-Koch et al. 2023; Enns et al. 2023). This ongoing pollution can facilitate the spread and establishment of *G. roeselii* into headwaters (Mauchart et al. 2014; Jourdan et al. 2024) and can lead to the decline of the more sensitive G. fossarum (Enns et al. 2023). In addition, temperature records for the herein studied sites show water temperatures between 14.4 °C and 21.5 °C, which are congruent with an earlier study of some of the sites (Grethlein et al. 2022). G. roeselii reaches sexual maturity faster, has a shorter brood development and a higher reproductive success than G. fossarum at temperatures above 12 °C (Pöckl et al. 2003). Furthermore, these characteristics of G. roeselii are enhancing with increasing temperature (Pöckl 1992). These life-history traits could facilitate invasion success and alter the community structure of invaded habitats (Grabowski et al. 2007; Rothhaupt et al. 2014). Thus, due to increasing water

temperatures associated with climate change and ongoing pollution through anthropogenic impact, *G. roeselii* might outcompete *G. fossarum* in headwaters in the future (Pöckl et al. 2003; Enns et al. 2023; Jourdan et al. 2024).

Conclusions

Our study revealed that the non-native *G. roeselii* is morphologically similar to the native *G. fossarum* in headwater streams. We also found similar food items in the gut content, which showed the generally omnivorous lifestyle of both species. However, stable isotopes indicated that the trophic niches of both species differ substantially, with *G. roeselii* being more predaceous and generally having a broader niche. This indicates that, despite their shared morphological characteristics and omnivorous tendencies, there is a noticeable niche differentiation in *G. roeselii*, consequently influencing the headwater food web. In situations of ample resource availability, co-existence between both species may be possible. However, in cases of resource scarcity, we expect *G. roeselii* to be competitively superior, particularly given its ability to exploit a broader range of food resources, regardless of their quality.

Acknowledgements

We thank Erik Aschenbrenner and Jana Kabus for fieldwork assistance. We thank an anonymous reviewer for constructive comments that helped to improve the manuscript. Lars Pelikan thanks the Laboratory of Evolutionary Ecology of Hydrobionts at the Nature Research Centre in Vilnius, Lithuania, for their kind hospitality during his stay.

Lars Pelikan was funded by an Erasmus+ scholarship. The open access publication of this article was funded by the Open Access Publication Fund of Goethe University Frankfurt am Main.

References

- Anderson NH, Cummins KW (1979) Influences of diet on the life histories of aquatic insects. Journal of the Fisheries Research Board of Canada 36(3): 335–342. https://doi. org/10.1139/f79-052
- Andersson MB (1994) Sexual Selection. Princeton University Press, Princeton, 599 pp.
- Araújo MS, Bolnick DI, Layman CA (2011) The ecological causes of individual specialisation. Ecology Letters 14(9): 948–958. https://doi.org/10.1111/j.1461-0248.2011.01662.x
- Aumack CF, Lowe AT, Amsler CD, Amsler MO, McClintock JB, Baker BJ (2017) Gut content, fatty acid, and stable isotope analyses reveal dietary sources of macroalgal-associated amphipods along the western Antarctic Peninsula. Polar Biology 40(7): 1371–1384. https://doi.org/10.1007/s00300-016-2061-4
- Bacela-Spychalska K, Van Der Velde G (2013) There is more than one 'killer shrimp': trophic positions and predatory abilities of invasive amphipods of Ponto-Caspian origin: Trophic

position and predatory abilities of amphipods. Freshwater Biology 58(4): 730–741. https://doi.org/10.1111/fwb.12078

- Barbosa P, Castellanos I [Eds] (2005) Ecology of Predator-Prey Interactions. Oxford University Press, Oxford/New York, 394 pp. https://doi.org/10.1093/oso/9780195171204.001.0001
- Bellard C, Leroy B, Thuiller W, Rysman J-F, Courchamp F (2016) Major drivers of invasion risks throughout the world. Ecosphere 7(3): e01241. https://doi.org/10.1002/ecs2.1241
- Betz-Koch S, Jacobs B, Oehlmann J, Ratz D, Reutter C, Wick A, Oetken M (2023) Pesticide dynamics in three small agricultural creeks in Hesse, Germany. PeerJ 11: e15650. https:// doi.org/10.7717/peerj.15650
- Bij de Vaate A, Jazdzewski K, Ketelaars HAM, Gollasch S, Van der Velde G (2002) Geographical patterns in range extension of Ponto-Caspian macroinvertebrate species in Europe. Canadian Journal of Fisheries and Aquatic Sciences 59(7): 1159–1174. https://doi.org/10.1139/f02-098
- Bock WJ, von Wahlert G (1965) Adaptation and the Form-Function Complex. Evolution 19(3): 269–299. https://doi.org/10.2307/2406439
- Bollache L, Kaldonski N, Troussard J-P, Lagrue C, Rigaud T (2006) Spines and behaviour as defences against fish predators in an invasive freshwater amphipod. Animal Behaviour 72(3): 627–633. https://doi.org/10.1016/j.anbehav.2005.11.020
- Bowes RE, Thorp JH (2015) Consequences of employing amino acid vs. bulk-tissue, stable isotope analysis: A laboratory trophic position experiment. Ecosphere 6(1): art14. https:// doi.org/10.1890/ES14-00423.1
- Breheny P, Burchett W (2017) Visualization of Regression Models Using visreg. The R Journal 9(2): 1–56. https://doi.org/10.32614/RJ-2017-046
- Cabana G, Rasmussen JB (1996) Comparison of aquatic food chains using nitrogen isotopes. Proceedings of the National Academy of Sciences of the United States of America 93(20): 10844–10847. https://doi.org/10.1073/pnas.93.20.10844
- Caut S, Angulo E, Courchamp F (2008) Dietary shift of an invasive predator: Rats, seabirds and sea turtles. Journal of Applied Ecology 45(2): 428–437. https://doi.org/10.1111/j.1365-2664.2007.01438.x
- Chesson P (2000) Mechanisms of maintenance of species diversity. Annual Review of Ecology and Systematics 31(1): 343–366. https://doi.org/10.1146/annurev.ecolsys.31.1.343
- Copilaș-Ciocianu D, Borza P, Petrusek A (2020) Extensive variation in the morphological antipredator defense mechanism of *Gammarus roeselii* Gervais, 1835 (Crustacea: Amphipoda). Freshwater Science 39(1): 47–55. https://doi.org/10.1086/707259
- Copilaș-Ciocianu D, Boros B, Šidagytė-Copilas E (2021) Morphology mirrors trophic niche in a freshwater amphipod community. Freshwater Biology 66(10): 1968–1979. https://doi. org/10.1111/fwb.13804
- Cothran RD, Henderson KA, Schmidenberg D, Relyea RA (2013) Phenotypically similar but ecologically distinct: Differences in competitive ability and predation risk among amphipods. Oikos 122(10): 1429–1440. https://doi.org/10.1111/j.1600-0706.2013.00294.x
- Crenier C, Arce-Funck J, Bec A, Billoir E, Perrière F, Leflaive J, Guérold F, Felten V, Danger M (2017) Minor food sources can play a major role in secondary production in detritus-based ecosystems. Freshwater Biology 62(7): 1155–1167. https://doi.org/10.1111/fwb.12933
- Csapó H, Krzywoźniak P, Grabowski M, Wattier R, Bącela-Spychalska K, Mamos T, Jelić M, Rewicz T (2020) Successful post-glacial colonization of Europe by single lineage of fresh-

water amphipod from its Pannonian Plio-Pleistocene diversification hotspot. Scientific Reports 10(1): e18695. https://doi.org/10.1038/s41598-020-75568-7

- Cummins KW, Klug MJ (1979) Feeding ecology of stream invertebrates. Annual Review of Ecology and Systematics 10(1): 147–172. https://doi.org/10.1146/annurev.es.10.110179.001051
- Delong MD, Summers RB, Thorp JH (1993) Influence of food type on the growth of a riverine amphipod, *Gammarus fasciatus*. Canadian Journal of Fisheries and Aquatic Sciences 50(9): 1891–1896. https://doi.org/10.1139/f93-211
- DeNiro MJ, Epstein S (1978) Influence of diet on the distribution of carbon isotopes in animals. Geochimica et Cosmochimica Acta 42(5): 495–506. https://doi.org/10.1016/0016-7037(78)90199-0
- DeNiro MJ, Epstein S (1981) Influence of diet on the distribution of nitrogen isotopes in animals. Geochimica et Cosmochimica Acta 45(3): 341–351. https://doi.org/10.1016/0016-7037(81)90244-1
- Dick JTA (1996) Post-invasion amphipod communities of lough neagh, Northern Ireland: Influences of habitat selection and mutual predation. Journal of Animal Ecology 65(6): 756–767. https://doi.org/10.2307/5674
- Dick JTA, Platvoet D (1996) Intraguild predation and species exclusions in amphipods: The interaction of behaviour, physiology and environment. Freshwater Biology 36(2): 375–383. https://doi.org/10.1046/j.1365-2427.1996.00106.x
- Dick JTA, Montgomery I, Elwood RW (1993) Replacement of the indigenous amphipod Gammarus duebeni celticus by the Introduced G. pulex: Differential cannibalism and mutual predation. Journal of Animal Ecology 62(1): 79–88. https://doi.org/10.2307/5484

Endler JA (1986) Natural Selection in the Wild. Princeton University Press, Princeton, 336 pp.

- Enns D, Cunze S, Baker NJ, Oehlmann J, Jourdan J (2023) Flushing away the future: The effects of wastewater treatment plants on aquatic invertebrates. Water Research 243: e120388. https://doi.org/10.1016/j.watres.2023.120388
- Finlay JC (2001) Stable-carbon-isotope ratios of river biota: Implications for energy flow in lotic food webs. Ecology 82(4): 1052–1064. https://doi.org/10.2307/2679902
- Finlay JC (2004) Patterns and controls of lotic algal stable carbon isotope ratios. Limnology and Oceanography 49(3): 850–861. https://doi.org/10.4319/lo.2004.49.3.0850
- Fišer Ž, Altermatt F, Zakšek V, Knapič T, Fišer C (2015) Morphologically cryptic amphipod species are "Ecological clones" at regional but not at local scale: A case study of four *Niphargus* species. PLOS ONE 10: e0134384. https://doi.org/10.1371/journal.pone.0134384
- Gallardo B, Clavero M, Sánchez MI, Vilà M (2016) Global ecological impacts of invasive species in aquatic ecosystems. Global Change Biology 22(1): 151–163. https://doi.org/10.1111/gcb.13004
- Garton DW, Payne CD, Montoya JP (2005) Flexible diet and trophic position of dreissenid mussels as inferred from stable isotopes of carbon and nitrogen. Canadian Journal of Fisheries and Aquatic Sciences 62(5): 1119–1129. https://doi.org/10.1139/f05-025
- Georgievová B, Zhai M, Bojková J, Šorfová V, Syrovátka V, Polášková V, Schenková J, Horsák M (2020) Does predation by the omnivorous *Gammarus fossarum* affect small-scale distribution of macroinvertebrates? A case study from a calcareous spring fen. International Review of Hydrobiology 105(5–6): 162–170. https://doi.org/10.1002/iroh.202002046

- Gessner MO, Chauvet E, Dobson M (1999) A perspective on leaf litter breakdown in streams. Oikos 85(2): 377–384. https://doi.org/10.2307/3546505
- Grabowski M, Bacela K, Konopacka A (2007) How to be an invasive gammarid (Amphipoda: Gammaroidea)–comparison of life history traits. Hydrobiologia 590(1): 75–84. https://doi.org/10.1007/s10750-007-0759-6
- Grabowski M, Mamos T, Bącela-Spychalska K, Rewicz T, Wattier RA (2017) Neogene paleogeography provides context for understanding the origin and spatial distribution of cryptic diversity in a widespread Balkan freshwater amphipod. PeerJ 5: e3016. https://doi.org/10.7717/peerj.3016
- Graça MAS (2001) The role of invertebrates on leaf litter decomposition in streams a review. International Review of Hydrobiology 86(4–5): 383–393. https://doi.org/10.1002/1522-2632(200107)86:4/5<383::AID-IROH383>3.0.CO;2-D
- Grethlein M, Pelikan L, Dombrowski A, Kabus J, Oehlmann J, Weigand A, Jourdan J (2022) Small-scale population structuring results in differential susceptibility to pesticide exposure. Environmental Sciences Europe 34(1): e113. https://doi.org/10.1186/s12302-022-00690-4
- Grey J, Jackson MC (2012) 'Leaves and eats shoots': Direct terrestrial feeding can supplement invasive red swamp crayfish in times of need. PLOS ONE 7: e42575. https://doi. org/10.1371/journal.pone.0042575
- Guerra-García JM, Tierno de Figueroa JM, Navarro-Barranco C, Ros M, Sánchez-Moyano JE, Moreira J (2014) Dietary analysis of the marine Amphipoda (Crustacea: Peracarida) from the Iberian Peninsula. Journal of Sea Research 85: 508–517. https://doi.org/10.1016/j. seares.2013.08.006
- Hairston Jr NG, Hairston Sr NG (1993) Cause-effect relationships in energy flow, trophic structure, and interspecific interactions. American Naturalist 142(3): 379–411. https:// doi.org/10.1086/285546
- Hammer Ø, Harper DAT, Ryan PD (2001) PAST: Paleontological statistics software package for education and data analysis. Palaeontologia Electronica 4: 1–9.
- Hood-Nowotny R, Knols BGJ (2007) Stable isotope methods in biological and ecological studies of arthropods. Entomologia Experimentalis et Applicata 124(1): 3–16. https://doi. org/10.1111/j.1570-7458.2007.00572.x
- Hutchins BT, Schwartz BF, Nowlin WH (2014) Morphological and trophic specialization in a subterranean amphipod assemblage. Freshwater Biology 59(12): 2447–2461. https://doi. org/10.1111/fwb.12440
- Inger R, Bearhop S (2008) Applications of stable isotope analyses to avian ecology: Avian stable isotope analysis. The Ibis 150(3): 447–461. https://doi.org/10.1111/j.1474-919X.2008.00839.x
- Jackson AL, Inger R, Parnell AC, Bearhop S (2011) Comparing isotopic niche widths among and within communities: SIBER – Stable Isotope Bayesian Ellipses in R: Bayesian isotopic niche metrics. Journal of Animal Ecology 80(3): 595–602. https://doi.org/10.1111/ j.1365-2656.2011.01806.x
- Jackson MC, Wasserman RJ, Grey J, Ricciardi A, Dick JTA, Alexander ME (2017) Novel and disrupted trophic links following invasion in freshwater ecosystems. Advances in ecological research 57: 55–97. https://doi.org/10.1016/bs.aecr.2016.10.006
- Jażdżewski K (1980) Range extensions of some Gammaridean species in european inland waters caused by human activity. Crustaceana (Supplement): 84–107.

- Jażdżewski K, Roux A-L (1988) Biogéographie de *Gammarus roeselii* Gervais en Europe, en particulier répartition en France et en Pologne. Crustaceana (Supplement): 272–277.
- Jeschke JM, Bacher S, Blackburn TM, Dick JTA, Essl F, Evans T, Gaertner M, Hulme PE, Kühn I, Mrugała A, Pergl J, Pyšek P, Rabitsch W, Ricciardi A, Richardson DM, Sendek A, Vilà M, Winter M, Kumschick S (2014) Defining the Impact of Non-Native Species. Conservation Biology 28(5): 1188–1194. https://doi.org/10.1111/cobi.12299
- Jourdan J, Westerwald B, Kiechle A, Chen W, Streit B, Klaus S, Oetken M, Plath M (2016) Pronounced species turnover, but no functional equivalence in leaf consumption of invasive amphipods in the river Rhine. Biological Invasions 18(3): 763–774. https://doi. org/10.1007/s10530-015-1046-5
- Jourdan J, Piro K, Weigand A, Plath M (2019) Small-scale phenotypic differentiation along complex stream gradients in a non-native amphipod. Frontiers in Zoology 16(1): 1–29. https://doi.org/10.1186/s12983-019-0327-8
- Jourdan J, El Toum Abdel Fadil S, Oehlmann J, Hupało K (2024) Rapid development of increased neonicotinoid tolerance in non-target freshwater amphipods. Environment International 183: e108368. https://doi.org/10.1016/j.envint.2023.108368
- Kabus J, Cunze S, Dombrowski A, Karaouzas I, Shumka S, Jourdan J (2023) Uncovering the Grinnellian niche space of the cryptic species complex *Gammarus roeselii*. PeerJ 11: e15800. https://doi.org/10.7717/peerj.15800
- Kenis M, Auger-Rozenberg M-A, Roques A, Timms L, Péré C, Cock MJW, Settele J, Augustin S, Lopez-Vaamonde C (2009) Ecological effects of invasive alien insects. Biological Invasions 11(1): 21–45. https://doi.org/10.1007/s10530-008-9318-y
- Kochmann J, Laier M, Klimpel S, Wick A, Kunkel U, Oehlmann J, Jourdan J (2023) Infection with acanthocephalans increases tolerance of *Gammarus roeselii* (Crustacea: Amphipoda) to pyrethroid insecticide deltamethrin. Environmental Science and Pollution Research International 30(19): 55582–55595. https://doi.org/10.1007/s11356-023-26193-0
- Kumschick S, Gaertner M, Vilà M, Essl F, Jeschke JM, Pyšek P, Ricciardi A, Bacher S, Blackburn TM, Dick JTA, Evans T, Hulme PE, Kühn I, Mrugała A, Pergl J, Rabitsch W, Richardson DM, Sendek A, Winter M (2015) Ecological impacts of alien species: Quantification, scope, caveats, and recommendations. Bioscience 65(1): 55–63. https://doi.org/10.1093/biosci/biu193
- Kuznetsova A, Brockhoff PB, Christensen RHB (2017) Imertest package: Tests in linear mixed effects models. Journal of Statistical Software 82(13): 1–26. https://doi.org/10.18637/jss.v082.i13
- Layman CA, Arrington DA, Montaña CG, Post DM (2007) Can stable isotope ratios provide for community-wide measures of trophic structure? Ecology 88(1): 42–48. https://doi. org/10.1890/0012-9658(2007)88[42:CSIRPF]2.0.CO;2
- Legeżyńska J, Kędra M, Walkusz W (2012) When season does not matter: Summer and winter trophic ecology of Arctic amphipods. Hydrobiologia 684(1): 189–214. https://doi. org/10.1007/s10750-011-0982-z
- Lenth R (2023) Emmeans: Estimated marginal means, aka least-squares means. R package version 1.8.8. https://CRAN.R-project.org/package=emmeans
- Leuven RSEW, van der Velde G, Baijens I, Snijders J, van der Zwart C, Lenders HJR, bij de Vaate A (2009) The river Rhine: A global highway for dispersal of aquatic invasive species. Biological Invasions 11(9): e1989. https://doi.org/10.1007/s10530-009-9491-7

- MacNeil C, Dick JTA, Elwood RW (1997) The trophic ecology of freshwater *Gammarus* spp. (Crustacea: Amphipoda): problems and perspectives concerning the functional feeding group concept. Biological Reviews of the Cambridge Philosophical Society 72(3): 349– 364. https://doi.org/10.1017/S0006323196005038
- Mancinelli G, Vizzini S (2015) Assessing anthropogenic pressures on coastal marine ecosystems using stable CNS isotopes: State of the art, knowledge gaps, and community-scale perspectives. Estuarine, Coastal and Shelf Science 156: 195–204. https://doi.org/10.1016/j. ecss.2014.11.030
- Mauchart P, Bereczki C, Ortmann-Ajkai A, Csabai Z, Szivák I (2014) Niche segregation between two closely similar gammarids (Peracarida, Amphipoda) – Native vs. naturalized nonnative species. Crustaceana 87(11–12): 1296–1314. https://doi.org/10.1163/15685403-00003355
- Mauchart P, Czirok A, Horvai V, Herczeg R, Móra A, Csabai Z (2017) Effects of meso- and microhabitat characteristics on the coexistence of two native gammarid species (Crustacea, Gammaridae). International Review of Hydrobiology 102(1–2): 38–46. https://doi. org/10.1002/iroh.201601855
- McCue MD, Javal M, Clusella-Trullas S, Le Roux JJ, Jackson MC, Ellis AG, Richardson DM, Valentine AJ, Terblanche JS (2020) Using stable isotope analysis to answer fundamental questions in invasion ecology: Progress and prospects. Methods in Ecology and Evolution 11: 196–214. https://doi.org/10.1111/2041-210X.13327
- McCutchan Jr JH, Lewis Jr WM, Kendall C, McGrath CC (2003) Variation in trophic shift for stable isotope ratios of carbon, nitrogen, and sulfur. Oikos 102(2): 378–390. https://doi. org/10.1034/j.1600-0706.2003.12098.x
- Médoc V, Piscart C, Maazouzi C, Simon L, Beisel J-N (2011) Parasite-induced changes in the diet of a freshwater amphipod: Field and laboratory evidence. Parasitology 138(4): 537–546. https://doi.org/10.1017/S0031182010001617
- Minagawa M, Wada E (1984) Stepwise enrichment of 15N along food chains: Further evidence and the relation between δ15N and animal age. Geochimica et Cosmochimica Acta 48(5): 1135–1140. https://doi.org/10.1016/0016-7037(84)90204-7
- Mothapo NP, Wossler TC (2017) Patterns of floral resource use by two dominant ant species in a biodiversity hotspot. Biological Invasions 19(3): 955–969. https://doi.org/10.1007/ s10530-016-1336-6
- Parker IM, Simberloff D, Lonsdale WM, Goodell K, Wonham M, Kareiva PM, Williamson MH, Von Holle B, Moyle PB, Byers JE, Goldwasser L (1999) Impact: Toward a framework for understanding the ecological effects of invaders. Biological Invasions 1(1): 3–19. https://doi.org/10.1023/A:1010034312781
- Peterson BJ, Fry B (1987) Stable isotopes in ecosystem studies. Annual Review of Ecology and Systematics 18(1): 293–320. https://doi.org/10.1146/annurev.es.18.110187.001453
- Peterson BJ, Howarth RW, Garritt RH (1986) Sulfur and carbon isotopes as tracers of saltmarsh organic matter flow. Ecology 67(4): 865–874. https://doi.org/10.2307/1939809
- Piscart C, Bergerot B, Laffaille P, Marmonier P (2010) Are amphipod invaders a threat to regional biodiversity? Biological Invasions 12(4): 853–863. https://doi.org/10.1007/ s10530-009-9506-4

- Piscart C, Roussel J-M, Dick JTA, Grosbois G, Marmonier P (2011) Effects of coexistence on habitat use and trophic ecology of interacting native and invasive amphipods: Diet shift and invasive species. Freshwater Biology 56(2): 325–334. https://doi.org/10.1111/j.1365-2427.2010.02500.x
- Pöckl M (1992) Effects of temperature, age and body size on moulting and growth in the freshwater amphipods *Gammarus fossarum* and *G. roeselii*. Freshwater Biology 27(2): 211–225. https://doi.org/10.1111/j.1365-2427.1992.tb00534.x
- Pöckl M (1993) Reproductive potential and lifetime potential fecundity of the freshwater amphipods *Gammarus fossarum* and *G. roeselii* in Austrian streams and rivers. Freshwater Biology 30(1): 73–91. https://doi.org/10.1111/j.1365-2427.1993.tb00790.x
- Pöckl M, Humpesch UH (1990) Intra- and inter-specific variations in egg survival and brood development time for Austrian populations of *Gammarus fossarum* and *G. ro-eselii* (Crustacea: Amphipoda). Freshwater Biology 23(3): 441–455. https://doi. org/10.1111/j.1365-2427.1990.tb00286.x
- Pöckl M, Webb BW, Sutcliffe DW (2003) Life history and reproductive capacity of Gammarus fossarum and G. roeselii (Crustacea: Amphipoda) under naturally fluctuating water temperatures: a simulation study: Natural thermal regimes and reproductive capacity of Gammarus spp. Freshwater Biology 48(1): 53–66. https://doi.org/10.1046/j.1365-2427.2003.00967.x
- Post DM (2002) Using stable isotopes to estimate trophic position: Models, methods, and assumptions. Ecology 83(3): 703–718. https://doi.org/10.1890/0012-9658(2002)083[0703:USI TET]2.0.CO;2
- Post DM, Pace ML, Hairston Jr NG (2000) Ecosystem size determines food-chain length in lakes. Nature 405(6790): 1047–1049. https://doi.org/10.1038/35016565
- Post DM, Layman CA, Arrington DA, Takimoto G, Quattrochi J, Montaña CG (2007) Getting to the fat of the matter: Models, methods and assumptions for dealing with lipids in stable isotope analyses. Oecologia 152(1): 179–189. https://doi.org/10.1007/s00442-006-0630-x
- Premate E, Borko Š, Delić T, Malard F, Simon L, Fišer C (2021) Cave amphipods reveal co-variation between morphology and trophic niche in a low-productivity environment. Freshwater Biology 66(10): 1876–1888. https://doi.org/10.1111/fwb.13797
- Premate E, Borko Š, Altermatt F, Fišer C (2023) Context-dependent evolution of high trophic position drives functional disparity in subterranean crustaceans. Functional Ecology 37(9): 2523–2534. https://doi.org/10.1111/1365-2435.14407
- Rosenfeld JS (2002) Functional redundancy in ecology and conservation. Oikos 98(1): 156–162. https://doi.org/10.1034/j.1600-0706.2002.980116.x
- Rothhaupt K-O, Hanselmann AJ, Yohannes E (2014) Niche differentiation between sympatric alien aquatic crustaceans: An isotopic evidence. Basic and Applied Ecology 15(5): 453– 463. https://doi.org/10.1016/j.baae.2014.07.002
- Rounick JS, Winterbourn MJ (1986) Stable carbon isotopes and carbon flow in ecosystems. Bioscience 36(3): 171–177. https://doi.org/10.2307/1310304
- Schoener TW (1974) Resource partitioning in ecological communities: Research on how similar species divide resources helps reveal the natural regulation of species diversity. Science 185(4145): 27–39. https://doi.org/10.1126/science.185.4145.27

- Sih A, Bolnick DI, Luttbeg B, Orrock JL, Peacor SD, Pintor LM, Preisser E, Rehage JS, Vonesh JR (2010) Predator-prey naïveté, antipredator behavior, and the ecology of predator invasions. Oikos 119(4): 610–621. https://doi.org/10.1111/j.1600-0706.2009.18039.x
- Simberloff D (2010) Invasive Species. Conservation Biology for All. Oxford University Press, Oxford. https://doi.org/10.1093/acprof:oso/9780199554232.003.0008
- Smith CL [Ed.] (1988) Fisheries research in the Hudson River food habits of the amphipod Gammarus tigrinus in the Hudson River and the effects of diet upon its growth and reproduction. State University of New York Press, Albany, 407 pp.
- Stoffels BEMW, Tummers JS, Van Der Velde GV, Platvoet D, Hendriks HWM, Leuven RSEW (2011) Assessment of predatory ability of native and non-native freshwater gammaridean species: A rapid test with water fleas as prey. Current Zoology 57(6): 836–843. https://doi. org/10.1093/czoolo/57.6.836
- Strayer DL, Eviner VT, Jeschke JM, Pace ML (2006) Understanding the long-term effects of species invasions. Trends in Ecology & Evolution 21(11): 645–651. https://doi. org/10.1016/j.tree.2006.07.007
- Summers RB, Delong MD, Thorp JH (1997) Ontogenetic and Temporal Shifts in the Diet of the Amphipod Gammarus fasciatus, in the Ohio River. American Midland Naturalist 137(2): 329–336. https://doi.org/10.2307/2426852
- Sutcliffe DW (1992) Reproduction in *Gammarus* (Crustacea, Amphipoda): basic processes. http://hdl.handle.net/1834/22140
- Syrovátka V, Zhai M, Bojková J, Šorfová V, Horsák M (2020) Native Gammarus fossarum affects species composition of macroinvertebrate communities: Evidence from laboratory, field enclosures, and natural habitat. Aquatic Ecology 54(2): 505–518. https://doi. org/10.1007/s10452-020-09756-y
- Tillberg CV, Holway DA, LeBrun EG, Suarez AV (2007) Trophic ecology of invasive Argentine ants in their native and introduced ranges. Proceedings of the National Academy of Sciences of the United States of America 104(52): 20856–20861. https://doi.org/10.1073/pnas.0706903105
- Torchin ME, Lafferty KD, Dobson AP, McKenzie VJ, Kuris AM (2003) Introduced species and their missing parasites. Nature 421(6923): 628–630. https://doi.org/10.1038/nature01346
- Van der Velde G, Rajagopal S, Kelleher B, Musko IB, Bij de Vaate A (2000) Ecological impact of crustacean invaders: General considerations and examples from the River Rhine. Crustacean Issues 12: 3–34.
- Van der Velde G, Leuven RSEW, Platvoet D, Bacela K, Huijbregts MAJ, Hendriks HWM, Kruijt D (2009) Environmental and morphological factors influencing predatory behaviour by invasive non-indigenous gammaridean species. Biological Invasions 11(9): 2043– 2054. https://doi.org/10.1007/s10530-009-9500-x
- Van Riel MC, Velde G, van der, Rajagopal S, Marguillier S, Dehairs F (2006) Trophic relationships in the Rhine Food Web during Invasion and after establishment of the ponto-caspian invader *Dikerogammarus villosus*. Hydrobiologia 565(1): 39–58. https://doi.org/10.1007/ s10750-005-1904-8
- Vander Zanden MJ, Rasmussen JB (1999) Primary consumer δ 13C and δ 15N and the trophic position of aquatic consumers. Ecology 80(4): 1395–1404. https://doi.org/10.1890/0012-9658(1999)080[1395:PCCANA]2.0.CO;2

- Vander Zanden MJ, Casselman JM, Rasmussen JB (1999) Stable isotope evidence for the food web consequences of species invasions in lakes. Nature 401(6752): 464–467. https://doi. org/10.1038/46762
- Vannote RL, Minshall GW, Cummins KW, Sedell JR, Cushing CE (1980) The river continuum concept. Canadian Journal of Fisheries and Aquatic Sciences 37(1): 130–137. https:// doi.org/10.1139/f80-017
- Vassallo L, Steele DH (1980) Survival and growth of young *Gammarus lawrencianus* Bousfield, 1956, on different diets. Crustaceana (Supplement): 118–125.
- Ward PI (1988) Sexual selection, natural selection, and body size in *Gammarus pulex* (Amphipoda). American Naturalist 131(3): 348–359. https://doi.org/10.1086/284794
- Weigand AM, Michler-Kozma D, Kuemmerlen M, Jourdan J (2020) Substantial differences in genetic diversity and spatial structuring among (cryptic) amphipod species in a mountainous river basin. Freshwater Biology 65(9): 1641–1656. https://doi.org/10.1111/fwb.13529
- Young KA (2004) Asymmetric competition, habitat selection, and niche overlap in juvenile salmonids. Ecology 85(1): 134–149. https://doi.org/10.1890/02-0402
- Zhang W, Hendrix PF, Snyder BA, Molina M, Li J, Rao X, Siemann E, Fu S (2010) Dietary flexibility aids Asian earthworm invasion in North American forests. Ecology 91(7): 2070– 2079. https://doi.org/10.1890/09-0979.1

Supplementary material I

Supplementary information

Authors: Lars Pelikan, Eglė Šidagytė-Copilas, Andrius Garbaras, Jonas Jourdan, Denis Copilas-Ciocianu

Data type: docx

- Explanation note: table S1. Results (*p*-values) of the PERMANOVA for the morphological traits of GR-F (female *G. roeselii*), GR-M (male *G. roeselii*), GF-F (female *G. fossarum*) and GF-M (male *G. fossarum*). PERMANOVA was performed with 9999 permutations and Euclidean similarity index. Bonferroni correction was applied for multiple comparisons between group pairs. Significant *p*-values (< 0.05) are marked in bold. table S2. Trophic position metrics of different amphipod size groups by study site (derived from stable isotope analysis relative to tree-leaf detritus). Provided values are means with standard deviations.</p>
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/neobiota.90.112383.suppl1