NeoBiota 82: 145–161 (2023)
doi: 10.3897/neobiota.82.97881
https://neobiota.pensoft.net

SHORT COMMUNICATION

A peer-reviewed open-access journal
NeoBiota
Advancing research on alien species and biological invasions

Genetic relationships among laboratory lines of the egg parasitoid *Trissolcus japonicus* from native and adventive populations

Paul K. Abram[1*], Tyler D. Nelson[2*], Valerie Marshall[2], Tara D. Gariepy[3], Tim Haye[4,5], Jinping Zhang[5], Tracy Hueppelsheuser[6], Susanna Acheampong[7], Chandra E. Moffat[2]

1 *Agriculture and Agri-Food Canada, Agassiz Research and Development Centre, 6947 Lougheed Hwy., Agassiz, BC, Canada* **2** *Agriculture and Agri-Food Canada, Summerland Research and Development Centre, 4200 BC-97, Summerland, BC, Canada* **3** *Agriculture and Agri-Food Canada, London Research and Development Centre, 1391 Sandford St, London, ON, Canada* **4** *CABI Switzerland, Rue des Grillions 1, CH-2800, Delemont, Switzerland* **5** *MARA-CABI Joint Laboratory for Bio-safety, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China* **6** *British Columbia Ministry of Agriculture and Food, 1767 Angus Campbell Road, Abbotsford, BC, Canada* **7** *British Columbia Ministry of Agriculture and Food, 1690 Powick Road, Kelowna, BC, Canada*

Corresponding author: Paul K. Abram (paul.abram@agr.gc.ca)

Academic editor: Richard Shaw | Received 23 November 2022 | Accepted 7 February 2023 | Published 27 February 2023

Citation: Abram PK, Nelson TD, Marshall V, Gariepy TD, Haye T, Zhang J, Hueppelsheuser T, Acheampong S, Moffat CE (2023) Genetic relationships among laboratory lines of the egg parasitoid *Trissolcus japonicus* from native and adventive populations. NeoBiota 82: 145–161. https://doi.org/10.3897/neobiota.82.97881

Abstract

Candidate biological control agents of invasive insect pests are increasingly being found in new geographic regions as a result of unintentional introductions. However, testing the degree of genetic differentiation among adventive and native-range populations of these agents is rarely done. We used reduced-representation sequencing of genomic DNA to investigate the relationships among laboratory lines of *Trissolcus japonicus* (Ashmead) (Hymenoptera, Scelionidae), an egg parasitoid and biological control agent of the brown marmorated stink bug, *Halyomorpha halys* (Stål) (Hemiptera, Pentatomidae). We compared sequences from multiple adventive populations in North America (Canada, USA) and Europe (Switzerland) with populations sourced from part of its native range in China. We found considerably more genetic variation among lines sourced from adventive populations than among those within native populations. In the Pacific Northwest of North America (British Columbia, Canada and Washington State, USA), we found preliminary evidence of three distinct genetic clusters, two of which were highly dissimilar from all

* These authors contributed equally to this work.

other lines we genotyped. In contrast, we found that other adventive lines with close geographic proximity (two from Ontario, Canada, three from Switzerland) had limited genetic variation. These findings provide a basis for testing biological differences among lines that will inform their use as biological control agents, and provide evidence to support a hypothesis of several independent introductions of *T. japonicus* in western North America from different source areas.

Keywords
classical biological control, ddRAD, *Halyomorpha halys*, Scelionidae, unintentional biological control

Introduction

There are now numerous documented instances where natural enemies of invasive insects have been discovered following the establishment of their host or prey species, presumably as a result of unintentional introductions (Roy et al. 2011; Weber et al. 2021). Several unintentional introductions are high-profile cases wherein natural enemies were discovered in the insect's invaded range while being evaluated for intentional biological control releases (e.g., Servick 2018; Abram et al. 2020). While molecular techniques have been used to confirm species-level identification of numerous invasive insect pests, determine their invasion history, their geographic origin(s), and possible genetic admixture among them (reviewed in Garnas et al. 2016), these techniques have less often been employed for their natural enemies (but see Lombaert et al. 2014; McCulloch et al. 2022).

In addition to determining invasion histories, molecular techniques could determine relationships among laboratory cultures of adventive natural enemy populations that have been sourced from different regions. When applied to these 'living genetic resources', genetic analyses could identify distinct populations that may differ in biological attributes (e.g., host range, life history, climate tolerance) that affect establishment success and suitability as biological control agents, and could inform future introductions (e.g., to increase genetic diversity of unintentionally introduced populations) or redistributions within new geographic areas that aim to improve biological control outcomes (Abram and Moffat 2018). A range of molecular markers are potentially available for such analyses, however, reduced-representation sequencing (RRS) has not been employed to genotype adventive natural enemies under consideration for biological control programs (Rius et al. 2015; McCartney et al. 2019; Leung et al. 2020). High-resolution genotypes derived from RRS methods could improve our ability to match lab-reared natural enemies with individual populations of invasive species, but these genotypes must first be characterised to permit proper assessment of their safety (potential non-target impacts) and efficacy (host specificity, foraging, and reproductive behaviours) as biocontrol agents.

Trissolcus japonicus (Ashmead) (Hymenoptera, Scelionidae) is an egg parasitoid of the brown marmorated stink bug *Halyomorpha halys* (Stål) (Hemiptera, Pentatomidae) whose presumed native range includes China, southeastern Russia, South Korea, Ja-

pan, and Taiwan (Yonow et al. 2021). In 2014, while *T. japonicus* was being evaluated as a candidate biological control agent, unintentionally introduced populations were detected in the northeastern USA (Talamas et al. 2015a). These findings were followed by detections in the western USA in 2015 (Milnes et al. 2016), Switzerland and eastern Canada in 2017 (Gariepy and Talamas 2019; Stahl et al. 2019), western Canada and Italy in 2018 (Peverieri et al. 2018; Abram et al. 2019), and several other locations throughout Europe and North America thereafter (e.g., Dieckhoff et al. 2021; reviewed in Conti et al. 2021). In the meantime, extensive research has been done on the parasitoid's host range and basic biology (reviewed in Conti et al. 2021; Abram et al. 2022), and intentional introductions of laboratory lines and redistributions of adventive lines are ongoing in Italy and the United States, respectively (Lowenstein et al. 2019; Conti et al. 2021). Stahl et al. (2019) did a preliminary haplotype analysis of the DNA barcode region of the mitochondrial cytochrome *c* oxidase I (COI) gene among populations of *T. japonicus* from Switzerland, Japan, and China, finding five haplotypes among Japanese specimens (one of which was the same as the single haplotype found in Switzerland) and a single base-pair difference between Swiss and Chinese specimens. However, single-marker analyses can be insufficient for detecting species- or population-level genetic structuring (Roe and Sperling 2007; Dupuis et al. 2012; Roe et al. 2017). Thus, in the Pacific Northwest of North America (British Columbia and Washington State), it is unclear whether recently discovered *T. japonicus* are one contiguous population or several distinct introductions. Here, we employed a RRS method, double digest restriction-site associated DNA sequencing (ddRADseq), to: (i) characterise the genetic relationships among laboratory lines established from populations in the native and adventive ranges of this parasitoid; and (ii) begin to evaluate the invasion history of *T. japonicus* in the Pacific Northwest.

Materials and methods

Insect collection and rearing

Between 2017 and 2020, we established 19 laboratory lines of *T. japonicus* in a containment facility certified by the Canadian Food Inspection Agency at Agriculture and Agri-Food Canada's Agassiz Research and Development Centre (Agassiz, British Columbia, Canada). We originally collected progenitors of these lines from wild populations in Switzerland, China, the USA, and Canada between 2009 and 2021 (Fig. 1; Suppl. material 1: table S1). We established 16 iso-female lines by taking a single female from a field-collected parasitized pentatomid egg mass and rearing at least 10 generations of its offspring. We established three additional lines using multiple individuals and reared each as mixed laboratory lines (Suppl. material 1: table S1). For each country of collection, we established two to eight lines. We reared all lines on *H. halys* egg masses following Wong et al. (2021). Elijah Talamas (Florida Department of Agriculture and Consumer Services) or Francesco Tortorici (Department of Agricultural,

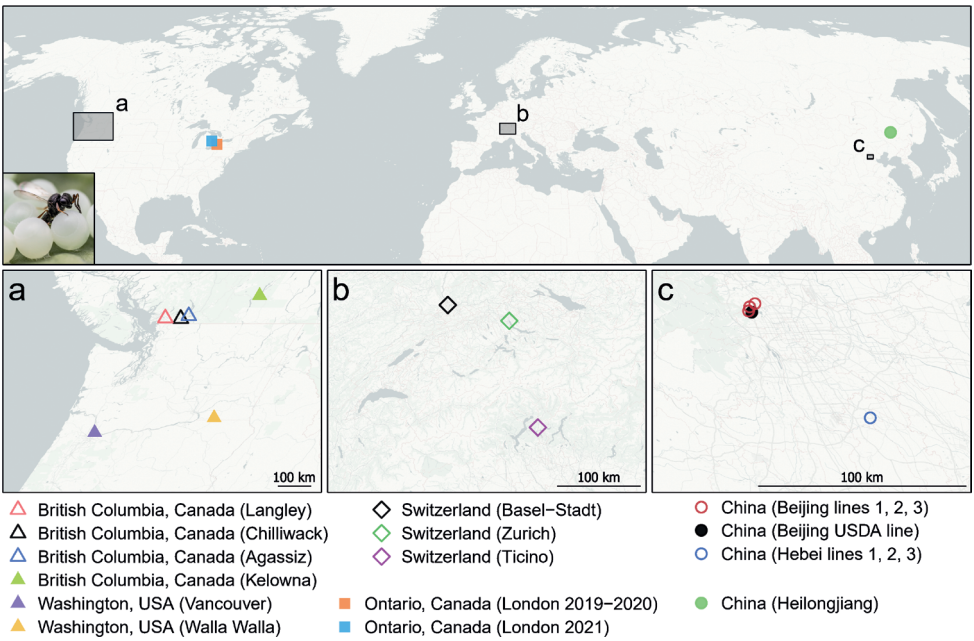

Figure 1. Collection locations for 19 laboratory lines of *Trissolcus japonicus*. Inset maps depict sampling regions **a** the Pacific Northwest of North America **b** Switzerland, and **c** Beijing and Hebei provinces, China. Symbol shape depicts geographic area of collection: diamond = Switzerland, triangle = Pacific Northwest of North America, circle = China, square = Ontario, Canada. Square symbols for the two locations in London, Ontario, Canada have been jittered for visibility.

Forest and Food Sciences, University of Torino) identified vouchers of each line to species-level (Talamas et al. 2015b; Talamas et al. 2017). We deposited representative vouchers in the Florida State Collection of Arthropods; the Canadian National Collection of Insects, Arachnids, and Nematodes; the Summerland Research and Development Centre arthropod collection; and the Royal British Columbia Museum.

DNA extraction and sequencing

We extracted genomic DNA using DNeasy Blood and Tissue DNA kits (QIAGEN, Hilden, Germany) by following the manufacturer's protocol, but we added a bovine ribonuclease A treatment (RNaseA, 4 uL at 100 mg/mL, QIAGEN) to digest RNA. We eluted DNA into 2× 50 uL of 56 °C Buffer AE to increase DNA concentration and yield, then we stored DNA at -20 °C until ddRAD library preparation. *PstI-MspI* library preparation and sequencing were performed by sequencing facility staff as outlined in MacDonald et al. (2020) on an Illumina NextSeq 500 at the University of Alberta (Edmonton, Alberta, Canada). We generated single-end 75 base-pair reads in two separate sequencing runs; we sequenced two samples twice to assess run effects.

Obtaining high-concentration, high-quality DNA from small-bodied organisms is an inherent challenge when preparing DNA libraries (e.g., Andersen et al. 2016; Paspati et al. 2019; Campbell et al. 2020), particularly for hymenopteran parasitoids (Cruaud et al. 2018; Gebiola et al. 2019; Ferguson et al. 2020). However, pooling individuals from female iso-lines has become standard practice in RRS and whole genome analyses of hymenopterans (Leung et al. 2020). We determined that extractions from individual *T. japonicus* did not yield enough DNA for successful ddRADseq (<<200 ng, TDN pers. obs.), and as such, we pooled 10 diploid female *T. japonicus* from individual lines for each DNA extraction (hereafter referred to as 'pools') to meet the minimum required concentration of DNA. Comparison of pooled close relatives may artificially inflate the perceived 'true' genetic distance between wild *T. japonicus* populations (e.g., Rodríguez-Ramilo and Wang 2012), however, our primary objective was to determine the genetic relatedness among laboratory lines. Because Bayesian model-based clustering can be more easily influenced by close relatives than other methods (Waples and Anderson 2017; O'Connell et al. 2019), we employed both Bayesian and non-Bayesian analysis to assess genetic structuring. We prepared a minimum of five pools per line (Table 1), however, we could only prepare four pools from the Ontario, Canada 2021 line due to low specimen availability when sampling. Each pool acted as one unit in analyses.

Table 1. The number of individual pools of 10 female wasps from each *Trissolcus japonicus* laboratory line included in each of the three analyses of population genetic structure. See Suppl. material 1: table S1 for additional information about each *T. japonicus* line.

Laboratory line	Number of pools included in dataset		
	full	geographic	Pacific Northwest
British Columbia, Canada (Langley)	7	–	7
British Columbia, Canada (Chilliwack)	5	–	5
British Columbia, Canada (Agassiz)	7	7	7
British Columbia, Canada (Kelowna)	7	7	7
Washington, USA (Vancouver)	5	4	4
Washington, USA (Walla Walla)	5	–	3
Switzerland (Basel-Stadt)	4	–	–
Switzerland (Zurich)	5	–	–
Switzerland (Ticino)	5	5	–
Ontario, Canada (London, 2019–2020)	4	4	–
Ontario, Canada (London, 2021)	3	–	–
China (Beijing line 1)	5	–	–
China (Beijing line 2)	5	–	–
China (Beijing line 3)	5	5	–
China (Beijing USDA line)	4	–	–
China (Hebei line 1)	5	–	–
China (Hebei line 2)	4	–	–
China (Hebei line 3)	5	–	–
China (Heilongjang)	5	4	–

Bioinformatics

We used Stacks 2 version 2.55 (Rochette et al. 2019) to demultiplex and process raw DNA reads. We removed reads if they 1) contained Phred scores below 20 over 15% of their length, 2) failed the Illumina chastity filter, or 3) had uncalled bases. We used the 'barcode rescue' option to retain reads with one mismatched base in its 8 base adaptor sequence, then we removed all adaptor sequences. Due to some sequencing error in the *PstI* restriction site, we removed an additional 5 bases from the 5' end of each read using Cutadapt version 3.4 (Martin 2011), resulting in final lengths of 62 bases. We used the denovo_map pipeline in Stacks 2 to call single nucleotide polymorphisms (SNPs), specifying one 'population' in the popmap and a minor allele frequency of 0.05. In accordance with Paris et al. (2017), we retained SNPs that were present across at least 80% of the pools. We completed final filtering using VCFtools version 0.1.16 (Danecek et al. 2011), retaining SNPs with a minimum read depth of five and discarding SNPs and pools that had more than 10% missing data, and we used the thin option to keep only one SNP per stack.

We assessed population genetic structuring for three datasets: one containing all 19 laboratory lines ('full dataset'), one with seven lines chosen for proportional representation of potential geographic clusters ('geographic dataset'), and one with all six lines from the Pacific Northwest of North America ('Pacific Northwest dataset') (Table 1). We used principal component analysis (PCA) and Bayesian model-based clustering for assessment. We performed PCA in adegenet version 2.1.5 (Jombart and Ahmed 2011) and we visualised results in ggplot version 3.3.5 (Wickham 2016) using R version 4.1.2 (R Core Team, 2021). We implemented Bayesian model-based clustering in structure version 2.3.4 (Pritchard et al. 2000) using the admixture model. For structure analysis, we randomly sub-selected pools from lines to ensure equal sample size from each (Puechmaille 2016), then ran a burn-in period of 100,000 sweeps followed by 1,000,000 sweeps for each of 10 replications for each potential subpopulation (K) between 1–20 (full dataset) or 1–15 (geographic and Pacific Northwest datasets). We defined each laboratory line in a given dataset as a 'location' prior to better resolving the genetic structure (Porras-Hurtado et al. 2013). We set the alpha prior of the full and geographic datasets to 1/7, and that of the Pacific Northwest dataset to 1/3, reflecting the expected results of K=7 or K=3 (Wang 2017). We assessed statistical support of each K value using LnP(K) (Pritchard et al. 2000) and ΔK (Evanno et al. 2005) in StructureSelector (Li and Liu 2018). Finally, we generated Q-matrices from the 10 replicates of each K for each dataset in CLUMPAK version 1.1 (Kopelman et al. 2015).

Data availability

DNA sequences are available as fastq files in the National Center for Biotechnology Information Sequence Read Archive (NCBI SRA) as BioProject PRJNA933214.

Results and discussion

Sequencing outcomes and geographic structure

In total, we sequenced 109 pools of *T. japonicus* (1090 individuals) from the 19 laboratory lines, resulting in 109,473,577 raw Illumina reads. In a preliminary PCA, two pools had unexpected behaviour. They may have been contaminated during sample preparation or DNA extraction: neither clustered with any other pool, and both were placed intermediate to other pools extracted from the same line and pools from other lines. We removed both before all subsequent analyses. After our final filtering of raw reads, the full dataset contained 1,889 SNPs across 95 pools with a mean SNP read depth of 64.9×, 11,360,339 filtered reads, and 3.82% total missing data. We used VCFtools to re-filter this dataset before running structure, using the same filtering parameters but sub-selecting near-equal sample sizes for pools from 18 lines in accordance with Puechmaille (2016). We did not include the 2021 line from Ontario, Canada in the full structure dataset because it only had two pools after filtering, leaving a total of 18 lines in the full structure dataset. In 16 of these 18 lines, we subsampled four pools; the other 2 lines (Walla Walla and Beijing-United States Department of Agriculture [USDA]) each had only three pools that passed our filters. Our final full structure dataset had 1,860 SNPs across 70 pools with a mean SNP read depth of 66.0×, 8,469,428 filtered reads, and 1.64% total missing data. In PCA, we identified 6–7 genetic clusters in the full dataset, and we found greatest statistical support for K=9 (LnP(K) method) and K=4 (ΔK method) in the full structure dataset (Fig. 2; Suppl. material 1: figs S1, S2).

In the geographic dataset, we retained 36 pools across the 7 laboratory lines after filtering raw reads. The dataset contained 2,896 SNPs with a mean SNP read depth of 72.9×, 7,498,995 filtered reads, and 2.03% total missing data. We used VCFtools to sub-select four pools per line before running structure, ensuring equal sample size. Our final geographic structure dataset had a mean SNP read depth of 67.4×, 5,386,771 filtered reads, and 2.45% total missing data across 28 pools. We found greatest statistical support for K=11 (LnP(K) method) and K=10 (ΔK method) in this dataset (Fig. 3, Suppl. material 1: figs S3, S4, K=11 structure plot not shown due to similarity with K=10).

In the Pacific Northwest dataset, we retained 33 pools across the 6 laboratory lines after filtering raw reads. This dataset contained 1,976 SNPs with a mean SNP read depth of 78.8×, 5,069,119 filtered reads, and 2.15% total missing data. We used VCFtools to sub-select four pools per line before running structure, ensuring equal sample size; however we could only select three high quality pools from the Walla Walla laboratory line. Our final Pacific Northwest structure dataset had a mean SNP read depth of 71.3×, 3,186,750 filtered reads, and 2.83% total missing data across 23 pools. We found greatest statistical support for K=12 (LnP(K) method) and K=3 (ΔK method) in this dataset (Fig. 4; Suppl. material 1: figs S5, S6).

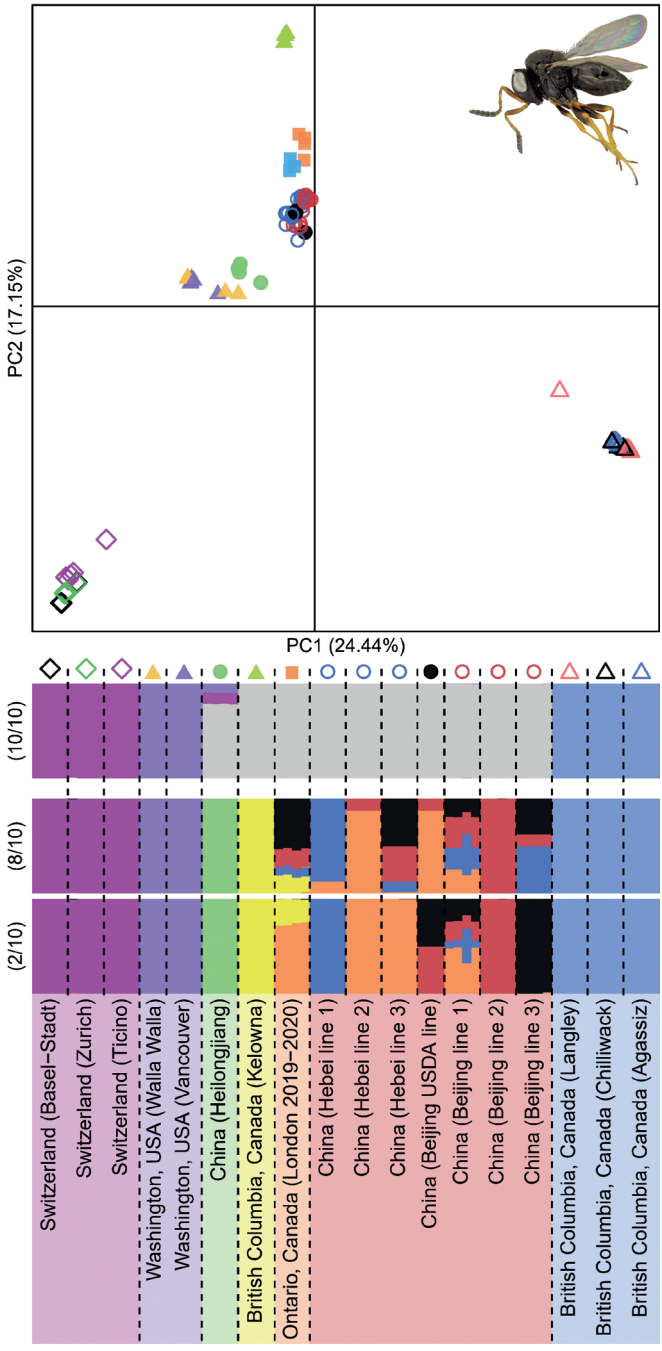

Figure 2. Principal component and structure analyses of SNP data from the full datasets comprising 18–19 *Trissolcus japonicus* laboratory lines. We present structure results with greatest LnP(*K*) and Δ*K* statistical support. Symbol shape depicts geographic area of collection: diamond = Switzerland, triangle = Pacific Northwest of North America, circle = China, square = Ontario, Canada (blue square = London, Ontario, Canada 2021 line). Colours behind laboratory line names correspond with geographic genetic clusters (Fig. 3). We present both modes of *K*=9 across its 10 replicate runs.

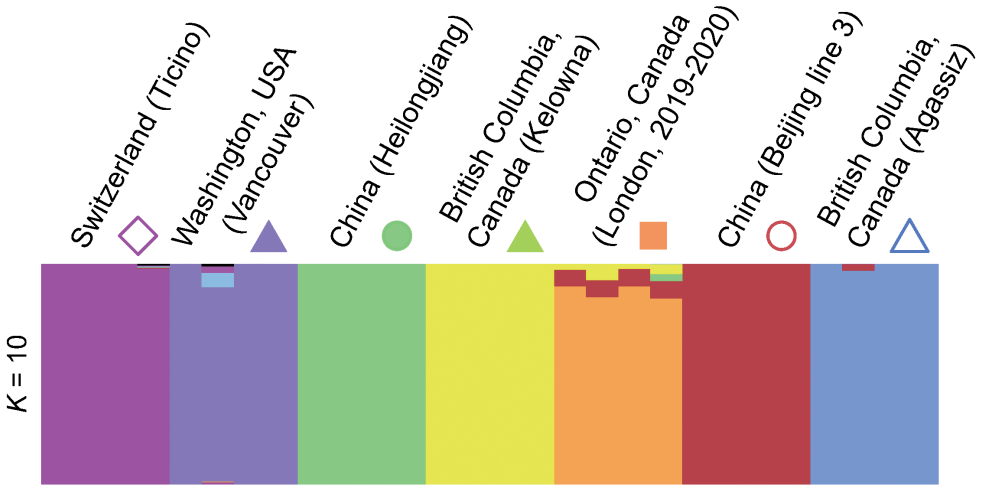

Figure 3. Structure analysis of SNP data from the geographic structure dataset comprising seven *Trissolcus japonicus* laboratory lines. We present structure results with greatest Δ*K* statistical support. Symbol shape depicts geographic area of collection: diamond = Switzerland, triangle = Pacific Northwest of North America, circle = China, square = Ontario, Canada.

Genetic similarities and differences among laboratory lines

Overall, we found *T. japonicus* lines had the greatest genetic similarity when collected in close geographic proximity (Figs 1, 2), but some lines were exceptional (Figs 2, 3). In both PCA and structure analyses, lines collected across Switzerland were more closely related to each other than lines from other countries, as were those from Beijing and Hebei provinces of China (Fig. 2), but lines from Switzerland and Beijing/Hebei were genetically different from one another (Fig. 2). At *K*=4, the Heilongjiang line was included in the same genetic cluster as Beijing and Hebei, however, at *K*=9 it belonged to its own cluster. This could be evidence of isolation by distance across the native range of *T. japonicus* and evidence of one introduction event in Switzerland that did not originate from the areas of China we sampled. In contrast, Stahl et al. (2019) found that the Ticino, Switzerland and Beijing lines had only one base substitution difference in their partial COI sequences (i.e., high apparent similarity) and inferred that the Swiss populations could have originated from the Beijing area. However, our results suggest that neither population-level differentiation nor patterns of invasion history of *T. japonicus* can be evaluated using the mitochondrial genome and thus require a nuclear genome-wide investigation.

Among adventive *T. japonicus* populations in Canada, the lines from London, Ontario and Kelowna, British Columbia were the most similar to populations from China, the parasitoid's native range, suggesting that these two populations may have originated from an area in proximity to our sampled Chinese populations. The two lines from London, Ontario were more closely related to the Beijing and Hebei cluster than was the Heilongjiang line in the full dataset (PCA and structure plots, Fig. 2), but the Ontario, Beijing/Hebei, and Heilongjiang lines each formed their own genetic cluster when proportionally

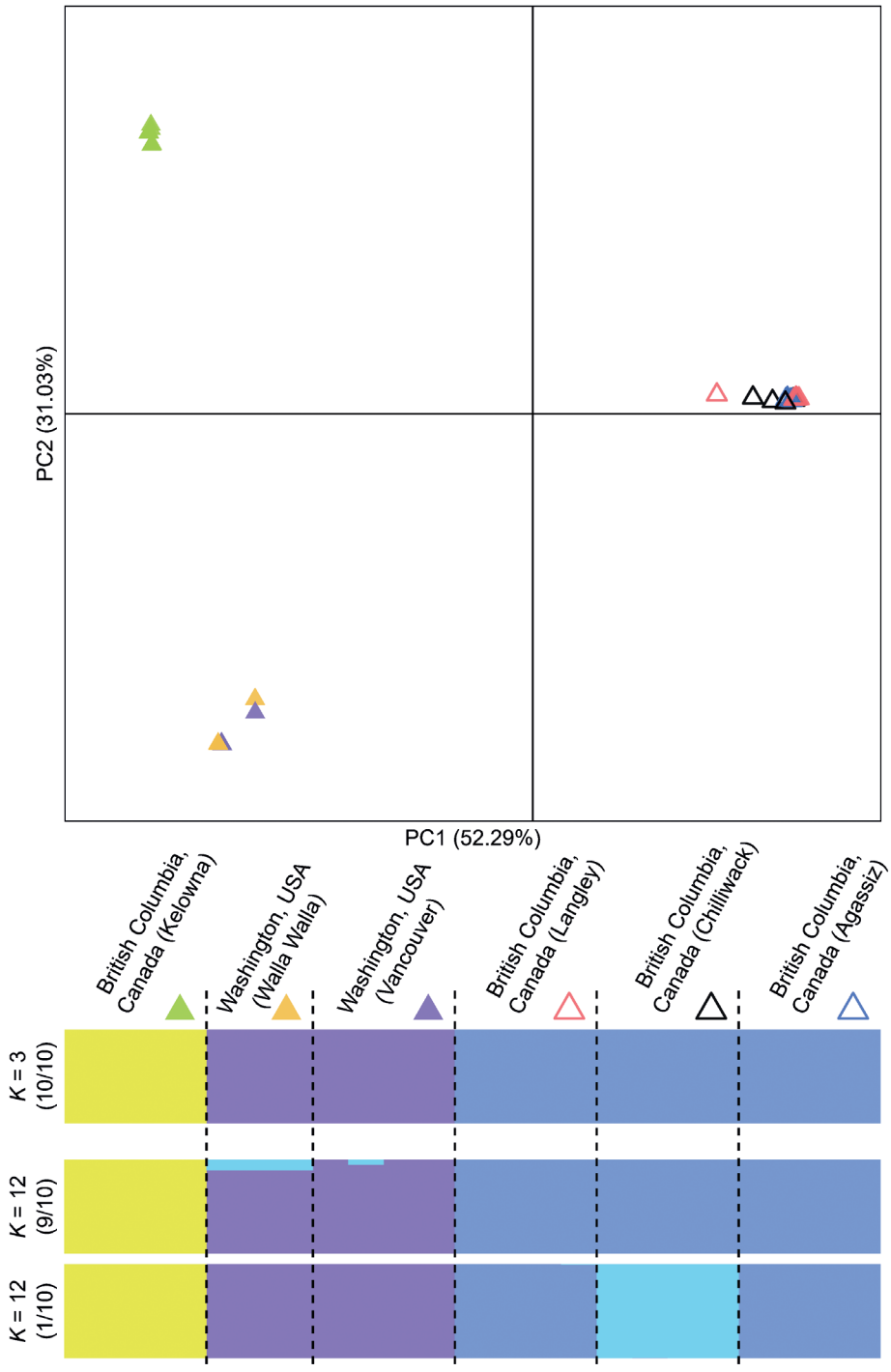

Figure 4. Principal component and structure analyses of SNP data from six *Trissolcus japonicus* laboratory lines collected across the Pacific Northwest of North America. We present the structure results with greatest ΔK and LnP(K) statistical support. We present both modes of K=12 across its 10 replicate runs.

represented in the geographic structure analysis (Fig. 3). It is likely that the seven Beijing/Hebei lines influenced the full dataset by over-representing the central China genotype, as each line is a genetic pseudoreplicate from the same population (see Puechmaille 2016). Nonetheless, this analysis suggests that the Ontario population could have originated from central China. Similarly, the Kelowna line was closely related to the Beijing and Hebei cluster at K=4 but formed its own cluster at K=9, suggesting it too may have been introduced from central China. Further genotyping of *T. japonicus* in its native range will be required to confirm the provenance of adventive populations.

We had expected that westernmost lines from Canada and the USA (Langley, Chilliwack, Agassiz, and Vancouver, WA) would be most closely related, as would those from the interior of BC and WA (Kelowna, BC and Walla Walla, WA). Instead, both lines from Washington State are members of the same cluster despite being separated by more than 350 kilometres, providing good evidence that *T. japonicus* in Walla Walla and Vancouver are either 1) descendants of a single introduction event in Washington or 2) two separate introduction events from the same region (Fig. 4). Likewise, the three lines from western British Columbia (Langley, Chilliwack, and Agassiz) form their own cluster that is genetically dissimilar from those collected in Washington and does not cluster near any other population we sampled. In addition, the newest detection in the Pacific Northwest, Kelowna, forms its own genetic cluster independent of both western BC and Washington State populations, indicating the population in interior British Columbia is likely an independent adventive introduction of unknown provenance and not a dispersion of the other adventive populations of *T. japonicus* in the Pacific Northwest. Strikingly, the amount of genetic variation within the Pacific Northwest lines alone is greater than populations separated by more than 1000 km in the native Chinese range. This suggests that there have been at least three distinct introductions of *T. japonicus* into the Pacific Northwest of North America, and that unintentional introductions of *T. japonicus* from different source areas may be happening with relative frequency. This may be consistent with the introduction history of its host, *H. halys*, in the Pacific Northwest, which appears to have resulted from multiple introduction events as suggested by the occurrence of at least three mitochondrial haplotypes (Abram et al. 2017). However, more extensive analyses using the same genotyping methods would be needed to adequately compare the invasion histories of *H. halys* and *T. japonicus*. In any case, future surveys for genetic and phenotypic variation in *T. japonicus* should not assume that populations in geographically proximal regions are necessarily the result of spread from adjacent regions.

Conclusions

Our study demonstrates that there is relatively strong population genetic structuring between *T. japonicus* laboratory lines collected at relatively small geographic scales, such as the Pacific Northwest of North America. One caveat of these analyses is the relatively low level of biological replication in certain genetic clusters. Nonetheless, clusters with more replicates of independently collected lines (n≥3: Switzerland; Beijing/

Hebei, China; and western British Columbia) did tend to have high genetic similarity relative to the much larger between-cluster variation. Because the geographic limits of these clusters are not yet known, it may be difficult to increase biological replication of the adventive populations. Several regions of the native and adventive ranges of *T. japonicus* are missing from the analyses (e.g., Japan, Italy, Eastern and Central USA), so more work is required to comprehensively describe the worldwide population genetic structure of this species. Secondly, the analyses compared inbred laboratory lines, possibly leading to greater perceived genetic differences between lines than the 'true' wild relationships due to high genetic similarity of each individual in a pool. However, the lines show little evidence of genetic drift towards a common 'lab genotype', and lines that have been in culture for many generations are still genetically similar to more recently established lines from the same genetic cluster, strongly suggesting that these living genetic resources are maintaining their individual integrity and are a close representation of the wild genotypic relationships. To build on this study and clarify the genetic relationships among these laboratory lines, we recommend further research comparing behavioural and life history attributes of each line to inform their use for biological control of *H. halys*. In addition, we suggest that for investigating patterns of invasion history for adventive or invasive species of parasitoids, data from RRS or other genome-wide methods be used, as inferences from single-gene sequencing can over-estimate genetic relatedness among disjunct populations.

Acknowledgements

We thank Josh Milnes, Betsy Beers, and Kim Hoelmer for laboratory lines of *Trissolcus japonicus* collected in 2016–2017 from Washington State and the strain collected in 2009 from Beijing, respectively. For ddRAD sequencing, we thank Sophie Dang and staff at the Molecular Biology Service Unit at the University of Alberta. We also thank Warren Wong, Jade Sherwood, Peggy Clarke, Chris Hou, Caitlyn MacDonald, Emily Grove, Laura Keery, Allison Briun, and Jason Thiessen for insect collection and rearing support. We again thank Warren Wong and Jason Thiessen for photos of *Trissolcus japonicus*. Finally, we thank one anonymous reviewer for their thoughtful comments on the manuscript. Funding to PKA and CEM is from Agriculture and Agri-Food Canada, ABASE #2955 and APMS #4609. Funding for TH and SA is from the Canadian Agricultural Partnership, a federal provincial territorial initiative.

References

Abram PK, Moffat CE (2018) Rethinking biological control programs as planned invasions. Current Opinion in Insect Science 27: 9–15. https://doi.org/10.1016/j.cois.2018.01.011

Abram PK, Hueppelsheuser T, Acheampong S, Clarke P, Douglas H, Gariepy TD (2017) Evidence of established brown marmorated stink bug populations in British Columbia, Canada. Journal of the Entomological Society of British Columbia 114: 83–86.

Abram PK, Talamas EJ, Acheampong S, Mason PG, Gariepy TD (2019) First detection of the samurai wasp, *Trissolcus japonicus* (Ashmead)(Hymenoptera, Scelionidae) in Canada. Journal of Hymenoptera Research 68: 29–36. https://doi.org/10.3897/jhr.68.32203

Abram PK, McPherson AE, Kula R, Hueppelsheuser T, Thiessen J, Perlman SJ, Curtis CI, Fraser JL, Tam J, Carrillo J, Gates M, Scheffer S, Lewis M, Buffington M (2020) New records of *Leptopilina*, *Ganaspis*, and *Asobara* species associated with *Drosophila suzukii* in North America, including detections of *L. japonica* and *G. brasiliensis*. Journal of Hymenoptera Research 78: 1–17. https://doi.org/10.3897/jhr.78.55026

Abram PK, Haye T, Clarke P, Grove E, Thiessen J, Gariepy TD (2022) Partial refuges from biological control due to intraspecific variation in protective host traits. Ecological Applications. https://doi.org/10.1002/eap.2796

Andersen JC, Bourchier RS, Grevstad FS, Van Driesche R, Mills NJ (2016) Development and verification of SNP arrays to monitor hybridization between two host-associated strains of knotweed psyllid, Aphalara itadori. Biological Control 93: 49–55. https://doi.org/10.1016/j.biocontrol.2015.11.007

Campbell EO, Dupuis JR, Holowachuk J, Hladun S, Vankosky MA, Mori BA (2020) Disjunction between canola distribution and the genetic structure of its recently described pest, the canola flower midge (*Contarinia brassicola*). Ecology and Evolution 10(23): 13284–13296. https://doi.org/10.1002/ece3.6927

Conti E, Avila G, Barratt B, Cingolani F, Colazza S, Guarino S, Hoelmer K, Laumann RA, Maistrello L, Martel G, Peri E, Rodriguez-Saona C, Rondoni G, Rostás M, Roversi PF, Sforza RFH, Tavella L, Wajnberg E (2021) Biological control of invasive stink bugs: Review of global state and future prospects. Entomologia Experimentalis et Applicata 169(1): 28–51. https://doi.org/10.1111/eea.12967

Cruaud A, Groussier G, Genson G, Sauné L, Polaszek A, Rasplus JY (2018) Pushing the limits of whole genome amplification: Successful sequencing of RADseq library from a single microhymenopteran (Chalcidoidea, Trichogramma). PeerJ 6: e5640. https://doi.org/10.7717/peerj.5640

Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R (2011) The variant call format and VCFtools. Bioinformatics 27(15): 2156–2158. https://doi.org/10.1093/bioinformatics/btr330

Dieckhoff C, Wenz S, Renninger M, Reißig A, Rauleder H, Zebitz CPW, Reetz J, Zimmermann O (2021) Add Germany to the list – adventive population of *Trissolcus japonicus* (Ashmead) (Hymenoptera: Scelionidae) emerges in Germany. Insects 12(5): e414. https://doi.org/10.3390/insects12050414

Dupuis JR, Roe AD, Sperling FAH (2012) Multi-locus species delimitation in closely related animals and fungi: One marker is not enough. Molecular Ecology 21(18): 4422–4436. https://doi.org/10.1111/j.1365-294X.2012.05642.x

Evanno G, Regnaut S, Goudet J (2005) Detecting the number of clusters of individuals using the software STRUCTURE: A simulation study. Molecular Ecology 14(8): 2611–2620. https://doi.org/10.1111/j.1365-294X.2005.02553.x

Ferguson KB, Kursch-Metz T, Verhulst EC, Pannebakker BA (2020) Hybrid genome assembly and evidence-based annotation of the egg parasitoid and biological control agent *Trichogramma brassicae*. G3 Genes/Genomes/Genetics 10: 3533–3540. https://doi.org/10.1534/g3.120.401344

Gariepy TD, Talamas EJ (2019) Discovery of *Trissolcus japonicus* (Hymenoptera: Scelionidae) in Ontario, Canada. Canadian Entomologist 151(6): 824–826. https://doi.org/10.4039/tce.2019.58

Garnas RJ, Auger-Rozenberg MA, Roques A, Bertelsmeier C, Wingfield MJ, Saccaggi DL, Roy HE, Slippers B (2016) Complex patterns of global spread in invasive insects: Eco-evolutionary and management consequences. Biological Invasions 18: 935–952. https://doi.org/10.1007/s10530-016-1082-9

Gebiola M, Streicher JW, Rugman-Jones PF, Morse JG, Stouthamer R (2019) Genome-wide analyses of single nucleotide polymorphisms reveal the consequences of traditional mass-rearing on genetic variation in *Aphytis melinus* (Hymenoptera: Aphelinidae): the danger of putting all eggs in one basket. Pest Management Science 75(11): 3102–3112. https://doi.org/10.1002/ps.5427

Jombart T, Ahmed I (2011) adegenet 1.3-1: New tools for the analysis of genome-wide SNP data. Bioinformatics 27(21): 3070–3071. https://doi.org/10.1093/bioinformatics/btr521

Kopelman NM, Mayzel J, Jakobsson M, Rosenberg NA, Mayrose I (2015) CLUMPAK: A program for identifying clustering modes and packaging population structure inferences across K. Molecular Ecology Resources 15(5): 1179–1191. https://doi.org/10.1111/1755-0998.12387

Leung K, Ras E, Ferguson KB, Ariëns S, Babendreier D, Bijma P, Bourtzis K, Brodeur J, Bruins MA, Centurión A, Chattington SR, Chinchilla-Ramírez M, Dicke M, Fatouros NE, González-Cabrera J, Groot TVM, Haye T, Knapp M, Koskinioti P, Le Hesran S, Lyrakis M, Paspati A, Pérez-Hedo M, Plouvier WN, Schlötterer C, Stahl JM, Thiel A, Urbaneja A, Zande L, Verhulst EC, Vet LEM, Visser S, Werren JH, Xia S, Zwaan BJ, Magalhães S, Beukeboom LW, Pannebakker BA (2020) Next-generation biological control: The need for integrating genetics and genomics. Biological Reviews of the Cambridge Philosophical Society 95(6): 1838–1854. https://doi.org/10.1111/brv.12641

Li YL, Liu JX (2018) StructureSelector: A web based software to select and visualize the optimal number of clusters using multiple methods. Molecular Ecology Resources 18(1): 176–177. https://doi.org/10.1111/1755-0998.12719

Lombaert E, Guillemaud T, Lundgren J, Koch R, Facon B, Grez A, Loomans A, Malausa T, Nedved O, Rhule E, Staverlokk A, Steenberg T, Estoup A (2014) Complementarity of statistical treatments to reconstruct worldwide routes of invasion: The case of the Asian ladybird *Harmonia axyridis*. Molecular Ecology 23(24): 5979–5997. https://doi.org/10.1111/mec.12989

Lowenstein DM, Andrews H, Hilton RJ, Kaiser C, Wiman NG (2019) Establishment in an introduced range: Dispersal capacity and winter survival of *Trissolcus japonicus*, an adventive egg parasitoid. Insects 10(12): e443. https://doi.org/10.3390/insects10120443

MacDonald ZG, Dupuis JR, Davis CS, Acorn JH, Nielsen SE, Sperling FAH (2020) Gene flow and climate-associated genetic variation in a vagile habitat specialist. Molecular Ecology 29(20): 3889–3906. https://doi.org/10.1111/mec.15604

Martin M (2011) Cutadapt removes adapter sequences from high-throughput sequencing reads. EMBnet.journal 17(1): 10–12. https://doi.org/10.14806/ej.17.1.200

McCartney MA, Mallez S, Gohl DM (2019) Genome projects in invasion biology. Conservation Genetics 20(6): 1201–1222. https://doi.org/10.1007/s10592-019-01224-x

McCulloch GA, Makinson JR, Purcell MF, Raghu S, Walter GH (2022) A prospective and iterative approach to finding safe weed biological control agents – testing ecological and evolutionary hypotheses with molecular evidence. Biological Control 169: e104887. https://doi.org/10.1016/j.biocontrol.2022.104887

Milnes JM, Wiman NG, Talamas EJ, Brunner JF, Hoelmer KA, Buffington ML, Beers EH (2016) Discovery of an exotic egg parasitoid of the brown marmorated stink bug, *Halyomorpha halys* (Stål) in the Pacific Northwest. Proceedings of the Entomological Society of Washington 118(3): 466–470. https://doi.org/10.4289/0013-8797.118.3.466

O'Connell KA, Muler KP, Maldonado J, Currie KL, Ferraro DM (2019) Sampling related individuals within ponds biases estimates of population structure in a pond-breeding amphibian. Ecology and Evolution 9(6): 3620–3636. https://doi.org/10.1002/ece3.4994

Paris JR, Stevens JR, Catchen JM (2017) Lost in parameter space: A road map for stacks. Methods in Ecology and Evolution 8(10): 1360–1373. https://doi.org/10.1111/2041-210X.12775

Paspati A, Ferguson KB, Verhulst EC, Urbaneja A, González-Cabrera J, Pannebakker BA (2019) Effect of mass rearing on the genetic diversity of the predatory mite *Amblyseius swirskii*. Entomologia Experimentalis et Applicata 167(7): 670–681. https://doi.org/10.1111/eea.12811

Peverieri GS, Talamas E, Bon M-C, Marianelli L, Bernardinelli I, Malossini G, Benvenuto L, Roversi PF, Hoelmer K (2018) Two Asian egg parasitoids of *Halyomorpha halys* (Stål)(Hemiptera, Pentatomidae) emerge in northern italy: *Trissolcus mitsukurii* (Ashmead) and *Trissolcus japonicus* (Ashmead) (Hymenoptera, Scelionidae). Journal of Hymenoptera Research 67: 37–53. https://doi.org/10.3897/jhr.67.30883

Porras-Hurtado L, Ruiz Y, Santos C, Phillips C, Carracedo Á, Lareu MV (2013) An overview of STRUCTURE: Applications, parameter settings, and supporting software. Frontiers in Genetics 4: 1–13. https://doi.org/10.3389/fgene.2013.00098

Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. Genetics 155(2): 945–959. https://doi.org/10.1093/genetics/155.2.945

Puechmaille SJ (2016) The program structure does not reliably recover the correct population structure when sampling is uneven: Subsampling and new estimators alleviate the problem. Molecular Ecology Resources 16(3): 608–627. https://doi.org/10.1111/1755-0998.12512

R Core Team (2021) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna. https://www.R-project.org

Rius M, Bourne S, Hornsby HG, Chapman MA (2015) Applications of next-generation sequencing to the study of biological invasions. Current Zoology 61(3): 488–504. https://doi.org/10.1093/czoolo/61.3.488

Rochette NC, Rivera-Colón AG, Catchen JM (2019) Stacks 2: Analytical methods for paired-end sequencing improve RADseq-based population genomics. Molecular Ecology 28(21): 4737–4754. https://doi.org/10.1111/mec.15253

Rodríguez-Ramilo ST, Wang J (2012) The effect of close relatives on unsupervised Bayesian clustering algorithms in population genetic structure analysis. Molecular Ecology Resources 12(5): 873–884. https://doi.org/10.1111/j.1755-0998.2012.03156.x

Roe AD, Sperling FAH (2007) Patterns of evolution of mitochondrial cytochrome c oxidase I and II DNA and implications for DNA barcoding. Molecular Phylogenetics and Evolution 1(1): 325–345. https://doi.org/10.1016/j.ympev.2006.12.005

Roe A, Dupuis J, Sperling FAH (2017) Molecular dimensions of insect taxonomy in the genomics era. In: Foottit RG, Adler PH (Eds) Insect Biodiversity, chapter 16. https://doi.org/10.1002/9781118945568.ch16

Roy HE, De Clercq P, Lawson Handley LJ, Poland RL, Sloggett JJ, Wajnberg E (2011) Alien arthropod predators and parasitoids: An ecological approach. BioControl 56(4): 375–382. https://doi.org/10.1007/s10526-011-9388-0

Servick K (2018) Control freaks. Science 361(6402): 542–545. https://doi.org/10.1126/science.361.6402.542

Stahl J, Tortorici F, Pontini M, Bon MC, Hoelmer K, Marazzi C, Tavella L, Haye T (2019) First discovery of adventive populations of *Trissolcus japonicus* in Europe. Journal of Pest Science 92(2): 371–379. https://doi.org/10.1007/s10340-018-1061-2

Talamas EJ, Herlihy MV, Dieckhoff C, Hoelmer KA, Buffington ML, Bon MC, Weber DC (2015a) *Trissolcus japonicus* (Ashmead) (Hymenoptera, Scelionidae) emerges in North America. Journal of Hymenoptera Research 43: 119–128. https://doi.org/10.3897/JHR.43.4661

Talamas EJ, Johnson NF, Buffington M (2015b) Key to Nearctic species of *Trissolcus* Ashmead (Hymenoptera, Scelionidae), natural enemies of native and invasive stink bugs (Hemiptera, Pentatomidae). Journal of Hymenoptera Research 43: 45–110. https://doi.org/10.3897/JHR.43.8560

Talamas EJ, Buffington ML, Hoelmer K (2017) Revision of Palearctic *Trissolcus* Ashmead (Hymenoptera, Scelionidae). Journal of Hymenoptera Research 56: 3–185. https://doi.org/10.3897/jhr.56.10158

Wang J (2017) The computer program structure for assigning individuals to populations: easy to use but easier to misuse. Molecular Ecology Resources 17: 981–990. https://doi.org/10.1111/1755-0998.12650

Waples RS, Anderson EC (2017) Purging putative siblings from population genetic data sets: A cautionary view. Molecular Ecology 25(5): 1211–1224. https://doi.org/10.1111/mec.14022

Weber D, Hajek AE, Hoelmer KA, Schaffner U, Mason P, Stouthamer R, Talamas EJ, Buffington M, Hoddle MS, Haye T (2021) Unintentional biological control. In: Mason PG (Ed.) Biological Control: Global Impacts, Challenges and Future Directions of Pest Management. CSIRO Publishing, Australia, 110–140.

Wickham H (2016) ggplot2: Elegant Graphics for Data Analysis. https://ggplot2.tidyverse.org

Yonow T, Kriticos DJ, Ota N, Avila GA, Hoelmer KA, Chen H, Caron V (2021) Modelling the potential geographic distribution of two *Trissolcus* species for the brown marmorated stink bug, *Halyomorpha halys*. Insects 12(6): e491. https://doi.org/10.3390/insects12060491

Wong WHL, Walz MA, Oscienny AB, Sherwood JL, Abram PK (2021) An effective cold storage method for stockpiling *Halyomorpha halys* (Hemiptera: Pentatomidae) eggs for field surveys and laboratory rearing of *Trissolcus japonicus* (Hymenoptera: Scelionidae). Journal of Economic Entomology 114(2): 571–581. https://doi.org/10.1093/jee/toaa307

Yonow T, Kriticos DJ, Ota N, Avila GA, Hoelmer KA, Chen H, Caron V (2021) Modelling the potential geographic distribution of two *Trissolcus* species for the brown marmorated stink bug, *Halyomorpha halys*. Insects 12: e491. https://doi.org/10.3390/insects12060491

Supplementary material 1

Supplementary information

Authors: Paul K. Abram, Tyler D. Nelson, Valerie Marshall, Tara D. Gariepy, Tim Haye, Jinping Zhang, Tracy Hueppelsheuser, Susanna Acheampong, Chandra E. Moffat

Data type: table and figures (word document)

Link: https://doi.org/10.3897/neobiota.82.97881.suppl1